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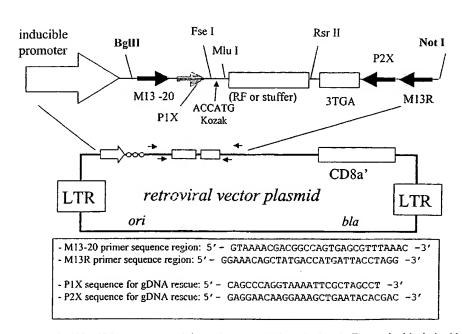
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#### (54) Title: TARGETS FOR CONTROLLING CELLULAR GROWTH AND FOR DIAGNOSTIC METHODS



(57) Abstract: A method of identifying a compound that induces apoptosis is disclosed. The method includes identifying compounds that inhibit the expression and/or activity of a target. Also disclosed are methods for inducing apoptosis by inhibiting one of the targets. The invention further includes methods for the diagnosis of a tumor that include determining the level of at least one of the targets as a biomarker in a patient sample, the level of the biomarker being indicative of the presence of tumor cells.

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

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# TARGETS FOR CONTROLLING CELLULAR GROWTH AND FOR DIAGNOSTIC METHODS

#### FIELD OF THE INVENTION

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The present invention relates to methods for inducing apoptosis in cells by inhibiting targets involved in the suppression of apoptosis, and to identifying compounds useful in such methods. The present invention also relates to methods for the diagnosis of cancer in a patient using the targets identified by the present invention as biomarkers.

#### 10 BACKGROUND OF THE INVENTION

The p53 tumor suppressor protein is an essential component in the regulation of the cell cycle, senescence, and programmed cell death (apoptosis). This protein regulates transcription of many genes in response to DNA damage and various transforming stimuli. The functional inactivation of p53 can occur through the action of viral oncoproteins, or through over-expression of the hdm2 (human) or mdm2 (murine) oncogene protein. Additional tumor suppressors, such as the p14<sup>ARF</sup> product of the INK4a gene, regulate the functional activity of p53. In the case of p14<sup>ARF</sup>, the suppressor interacts with hdm2 and thereby prevents the mentioned oncoprotein from inhibiting p53. An alternative translation product of the INK4a locus, p16INK4a, a cyclin-dependent kinase inhibitor, also contributes to normal growth control through its regulation of the Rb pathway.

When regulation of the cell cycle, senescence, and apoptosis is not functioning properly, uncontrolled cell growth and tumor formation occurs. Because of the complicated regulation of these cell functions, there are many potential points in a variety of regulatory pathways of a cell for intervention. By inhibiting the expression of genes important to cell growth and to suppression of apoptosis or the proteins encoded by them, it is possible to induce control cell growth and apoptosis in a cell, thereby preventing tumor formation. Once such genes or proteins are identified as targets, assays can be conducted for drug discovery to find inhibitors suitable for use as therapeutic agents. In addition, such genes or proteins are useful as markers of tumor formation.

There is an ongoing need to identify new targets and develop new assays for the identification of therapeutic compounds useful in the control of cell growth and tumor formation.

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#### SUMMARY OF THE INVENTION

This invention provides methods for identifying compounds that induce apoptosis by inhibiting target genes or gene products involved in the control of cell growth. The present invention also includes a method for inducing apoptosis in a cell by inhibiting such a target gene or gene product by, in one embodiment, contacting cells susceptible to uncontrolled growth with an inhibitory compound in an amount sufficient to inhibit said biochemical activity or expression. More particularly, targets of the present invention include any of the genes or gene products set forth in Table 1, which can also be identified as genes and gene products comprising SEQ ID NOs:1-80 (with odd numbered identifiers referring to nucleic acid sequences and even numbered identifiers referring to amino acid sequences).

In one embodiment, the present invention relates to a method of identifying a compound that induces apoptosis in a cell that includes contacting the cell with a putative apoptosis-inducing compound and determining whether the compound inhibits the expression and/or activity of a target selected from the group consisting of any of the targets listed in Table 1 (or comprising any of SEQ ID NOs:1-80). The target can have been validated as being involved in tumor cell growth, such as by a process of inhibiting the target in a cell by a method selected from gene knock-out, anti-sense oligonucleotide expression, use of RNAi molecules and GSE expression, or assaying the cell for the ability of the cell to grow. The cell can be a tumor cell line. The step of determining can be selected from assaying for reduced expression of the target and assaying for reduced activity of the target. The expression of the target can be measured by methods including, but not limited to, polymerase chain reaction or by using an antibody that specifically recognizes the target. The activity of the target can be measured by methods including, but not limited to, measuring the amount of a product generated in a biochemical reaction mediated by the target or by measuring the amount of a substrate consumed in a biochemical reaction mediated by the target. The inhibitor can be identified by methods including, but not limited to, determining the three-dimensional structure of the target or by determining the three-dimensional structure of an inhibitor by using computer software capable of modeling the interaction of the target and putative test compounds.

Another embodiment of the present invention is a method for inducing apoptosis in a cell by inhibiting a target selected from any of the genes or products encoded thereby

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listed in Table 1 (also represented herein as genes or gene products comprising any of SEQ ID NOs:1-80).

A further embodiment of the present invention is a method for the diagnosis of a tumor that includes determining the level of a biomarker selected from any of the genes or products encoded thereby listed in Table 1 (also represented herein as genes or gene products comprising any of SEQ ID NOs:1-80) in a patient test sample. In this method, the level of the biomarker is indicative of the presence of tumor cells. The presence of the biomarker at an increased level as compared to a normal baseline control is an indication of the presence a tumor, a possible predisposition to such tumor or a susceptibility to an anti-cancer therapeutic treatment. The level of the biomarker can be determined by conventional methods such as expression assays to determine the level of expression of the gene, by biochemical assays to determine the level of the gene product, or by immunoassays. In one embodiment of this method, the level of the biomarker can be determined by identifying the biomarker as a cell surface molecule in tissue or by detecting the biomarker in soluble form in a bodily fluid, such as serum, that can be immobilized. The biomarker level can be determined by contacting a patient test sample with an antibody, or a fragment thereof, that binds specifically to the biomarker and determining whether the anti-biomarker antibody or fragment has bound to the biomarker. The biomarker level can be determined by using a first monoclonal antibody that binds specifically to the biomarker and a second antibody that binds to the first antibody. This method can be used to determine the prognosis for cancer in the patient or to determine the susceptibility of the patient to a therapeutic treatment.

#### BRIEF DESCRIPTION OF THE DRAWINGS OF THE INVENTION

Fig. 1 illustrates a schematic of the features of the V98 vector.

Fig. 2 illustrates a schematic drawing of the construction of the V87 vector.

Fig. 3 illustrates a schematic drawing of the construction of the V98 vector

#### DETAILED DESCRIPTION OF THE INVENTION

The present invention includes methods for identifying protective compounds that control cell growth and induce apoptosis by using genes that encode products that are necessary for protecting cells from apoptosis as targets in the design of therapeutic agents. The invention further includes compounds for use in the treatment or prevention of tumor

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growth. Such compounds include chemical compounds and biological compounds. Chemical compounds or biological compounds include any chemical or biological compound that disrupts or inhibits one or more biological functions required for controlling cell growth. Preferred chemical compounds include small molecule inhibitor or substrate compounds, such as products of chemical combinatorial libraries. Preferred biological compounds include peptides, anti-sense molecules and antibodies.

The invention also includes methods for the diagnosis of cancer or for a prognosis of cancer or for determination of susceptibility to cancer treatments, by determining the level of expression of target genes and proteins of the present invention (also referred to as tumor antigens (TAGs)) in patient samples. The targets may originate from different parts of the cell and may be cell surface proteins, intracellular proteins or proteins that are secreted from the cell. There is a distinction between tissue, individual and speciesspecific cellular markers that may also be present physiologically as differentiation antigens on cells. There may be targets that are intermediate products released, over expressed or under expressed during the growth of a tumor cell type which can change upon further differentiation. The level of the target gene or protein can be determined by conventional methods such as expression assays to determine the level of expression of the gene, by biochemical assays to determine the level of the gene product, or by immunoassays. If appropriate, the marker can be identified as a cell surface molecule in tissue or in a bodily fluid, such as serum. These methods are described in detail below. The present invention provides much needed markers that permit an improved and more specific diagnosis of cancer, including the possible distinction between various tumor types, the prediction of tumor formation and the patient susceptibility to certain known cancer treatments.

The present invention is based, in part, on the present inventors' isolation of certain GSEs from human cells that prevent cell growth, and the discovery that such nucleic acid molecules correspond to fragments of certain genes. In that regard, any cellular phenotype or protein associated with cell growth can be used to select for such nucleic acid molecules or proteins encoded thereby.

More specifically, targets of the present invention have been identified as corresponding to genetic suppressor elements (GSEs) that control cell growth. The GSX<sup>TM</sup> System technology allows rapid screening for the inhibitors of gene function in the form of genetic suppressor elements. Briefly, a Genetic Suppressor Element (GSE), is

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a gene fragment, which, when expressed in cells, acts as a genetic inhibitor of the corresponding intact gene in those cells. A GSE can exert its effect through either an antisense, or a dominant negative peptide mechanism. GSEs are selected from libraries of DNA fragments, generated by random breakage of sets of test genes, cloned in a retroviral or other expression vector. The RFL clones are introduced into a population of test cells at approximately one test fragment per cell. Cells with a desired new phenotype, resulting from the expression of a GSE, are isolated on the basis of any selectable parameter. The GSEs are recovered from the selected cells and characterized by DNA sequence analysis and further functional assays.

GSEs having the ability to control cell growth can be functional in the sense orientation (and encode a peptide thereby), and can be functional in the antisense orientation (and encode antisense RNAs thereby). These GSEs are believed to downregulate the corresponding gene from which they were derived by different mechanisms. Such a corresponding gene is referred to herein as a "target gene" and its product (i.e., protein encoded by the coding region of the gene) is referred to as a "target product" or "target protein". As used herein, the term "target" alone can refer collectively to a target gene and its corresponding target product, or to useful portions thereof. Sense-oriented GSEs exert their effects as transdominant mutants or RNA decoys. Transdominant mutants are expressed proteins or peptides that competitively inhibit the normal function of a wild-type protein in a dominant fashion. RNA decoys are protein binding sites that titrate out these wild-type proteins. Anti-sense oriented GSEs exert their effects as antisense RNA molecules, i.e., nucleic acid molecules complementary to the mRNA of the target gene. These nucleic acid molecules bind to mRNA and block the translation of the mRNA. In addition, some antisense nucleic acid molecules can act directly at the DNA level to inhibit transcription.

Specific targets of the present invention are shown below in the Examples section in Table 1. The targets include the genes and products of the genes or any useful portion thereof. Methods of the present invention for identifying therapeutic compounds by identifying an inhibitor of a target include identifying an inhibitor of: a target gene from Table 1, as well as target products encoded by any of the foregoing. Diagnostic methods for detecting cancer in a patient include detection of a target gene from Table 1, as well as target products encoded by any of the foregoing, and useful portions thereof. More specifically, the targets of the present invention include genes comprising all or a portion

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of any of the nucleic acid sequences represented by SEQ ID NOs: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, or 79. These nucleic acid molecules encode the following target proteins, respectively: angio-associated, migratory cell protein (AAMP; SEQ ID NO:2), a disintegrin and metalloproteinase domain 8 (ADAM8; SEQ ID NO:4), a disintegrin-like and metalloprotease (reporlysin type) with thrombospondin type 1 motif, 17 (ADAMTS17; SEQ ID NO:6), adenylate cyclase 3 (ADCY3; SEQ ID NO:8), adrenergic beta receptor kinase 1 (ADRBK1; SEQ ID NO:10), bladder cancer associated protein (BLCAP; SEQ ID NO:12), chromosome 22 open reading frame 5 (C22orf5; SEQ ID NO:14), CD81 antigen (target of antiproliferative antibody 1 (CD81; SEQ ID NO:16), CD9 antigen (p24) (CD9; SEQ ID NO:18), claudin 4 (CLDN4; SEQ ID NO:20), chloride intracellular channel 1 (CLIC1; SEQ ID NO:22), collagen, type VI, alpha 2 (COL6A2; SEQ ID NO:24), CTL2 (CTL2; SEQ ID NO:26), endothelin converting enzyme 1 (ECE1; SEQ ID NO:28), ephrin-B1 (EFNB1; SEQ ID NO:30), flotillin 2 (FLOT2; SEQ ID NO:32), intercellular adhesion molecule 3 (ICAM3; SEQ ID NO:34), iduronate 2sulfatase (Hunter syndrome) (IDS; SEQ ID NO:36), jagged 2 (JAG2; SEQ ID NO:38), junctional adhesion molecule 1 (JAM1; SEQ ID NO:40), lectin, galactoside-binding soluble 3 binding protein (LGALS3BP; SEQ ID NO:42), similar to possible G-protein receptor (LOC146330; SEQ ID NO:44), CGI-78 protein (LOC51107; SEQ ID NO:46), lipoprotein lipase (LPL; SEQ ID NO:48), low density lipoprotein receptor-related protein 5 (LRP5; SEQ ID NO:50), Lutheran blood group (Auberger b antigen included) (LU; SEQ ID NO:52), membrane component, chromosome 11, surface marker 1 (M11S1; SEQ ID NO:54), serum constituent protein (MSE55; SEQ ID NO:56), neuropathy target esterase (NTE; SEQ ID NO:58), Homo sapiens cDNA FL31043 fis, clone HSYRA2000248 (PLEXIN A1) or Homo sapiens cDNA FLJ44113 fis, clone TESTI4046487, highly similar to Mus musculus plexin A1 (PLXNA1; SEQ ID NO:60), protein tyrosine phosphatase, receptor type, f polypeptide (PTPRF), interacting protein (liprin), alpha 3 (PPFIA3; SEQ ID NO:62), Homo sapiens peptide-histidine transporter 4 (PTR4), mRNA (PTR4; SEQ ID NO:64), solute carrier family 16 (moncarboxylic acid transporters) member 3 (SLC16A3; SEQ ID NO:66), solute carrier family 1 (neutral amino acid transporter) member 5 (SLC1A5; SEQ ID NO:68), solute carrier family 39 (zinc transporter) member 3 (SLC39A1; SEQ ID NO:70), serine protease inhibitor, Kunitz type 2 (SPINT2; SEQ ID NO:72), stanniocalcin 2 (STC2; SEQ ID NO:74), tumor necrosis receptor superfamily member 21 (TNFRSF21; SEQ ID NO:76), tumor rejection antigen (gp96) 1 (TRA1; SEQ ID NO:78), and transient receptor potential cation channel, subfamily M member 4 (TRPM4; SEQ ID NO:80). In any of the assays described herein, one can use a full-length gene, including a regulatory region of the gene, or a nucleic acid molecule encoding the gene product (protein encoded by the gene) or any fragment of such nucleic acid molecules, or any gene product or fragment thereof that is suitable for use in an assay to identify inhibitors of the target for the purpose of regulating apoptosis or inhibition of tumor growth, or to detect cancer in a patient sample.

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In one embodiment of the invention, the down-regulation of the concentration or activity of a target gene or product by an inhibitor (including a GSE) depletes a cellular component required for protecting cells from apoptosis resulting in control of cell growth. In another embodiment of the invention, the down-regulation of the concentration or activity of one target gene or product by an inhibitor (including a GSE) depletes a cellular component that interacts with another gene or gene product required for protecting cells from apoptosis resulting in control of cell growth. In a preferred embodiment of the invention, the two genes are members of the same biological pathway and one gene or gene product regulates the expression or activity of the other gene or gene product. In another preferred embodiment of the invention, the two genes are members of the same biological pathway and the substrate of a protein encoded by one gene is a product of a biochemical reaction mediated by the protein encoded by the other gene. In still another preferred embodiment of the invention, the two genes are members of the same biological pathway and the product of a protein encoded by one gene is a substrate of a biochemical reaction mediated by the protein encoded by the other gene. In another embodiment, the two genes encode proteins that are isozymes of each other. In a preferred embodiment, at least one of the genes encodes an enzyme.

Target genes or proteins identified using GSEs can be further evaluated using a variety of methods to validate their involvement in cell growth, suppression of apoptosis and tumor formation. Such methods include methods that disrupt or "knock out" the expression of a target gene in a cell capable of apoptosis. Knock-out methods include somatic cell knock-outs and inhibitory RNA molecules including anti-sense oligonucleotides, siRNA molecules, RNAi molecules and RNA decoys. Target genes or proteins can also be evaluated by methods that include nucleic acid-based experiments such as Northern Blots, Real Time polymerase chain reaction or high density microarrays.

Further evaluation can also be achieved using human/mouse xenograft models. For example, human tumor cells can be transfected with a GSE such that the GSE is expressed. Preferred tumor cells include HCT15, HT29, HCT116, SW480 and SW620 and MDA-MB-231 (e.g., see Examples). The transfected cells can then be implanted into mice, preferably nude mice. The growth of the tumor cells in the mouse can then be measured.

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Once a gene has been identified as a potential target for supporting cell growth, assays can be used for associating a potential target with different tumor types. These assays include determining gene and protein expression of potential targets in different tumor cell types at different points of differentiation. Another assay can include determining the presence of a potential marker in patient samples using standard protein detection methods known to those of skill in the art. Targets that have been associated with cancer are also referred to as biomarkers. Preferred biomarkers of the present invention are listed in Table 1 (see Examples section).

Once one or more members of a biological pathway are identified as required for cell growth, the present invention can include identifying additional members of a biological pathway that are also required for cell growth. Such subsequent identification is within the skill of one in the art. GSEs, and therefore preferred targets of the present invention, are identified by selecting cells that exhibit certain hallmarks of apoptosis upon expression of the GSEs. Isolated GSEs are further prioritized based on their specificity for a neoplastic transformation state, such as their activity in transformed and non-transformed cells, and based on the p53 pathway status in cells expressing the GSEs. For example, GSEs can be prioritized by determining if the GSEs have activity in a p53 dependent and/or independent manner. GSEs specific for the neoplastic transformation state are preferred for identifying targets for anti-cancer drugs.

It will be understood that this invention is not limited to the particular methodology, protocols, cell lines, animal species or genera, constructs, or reagents described herein, as such may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the invention that will be limited only by the appended claims. All technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this invention belongs unless clearly indicated otherwise.

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As used herein, the term "isolated nucleic acid molecule" refers to a nucleic acid molecule that has been removed from its natural milieu (i.e., a molecule that has been subject to human manipulation) and can include DNA, RNA, or derivatives of either DNA or RNA. An isolated nucleic acid molecule can be isolated from its natural source or can be produced using recombinant DNA technology (e.g., polymerase chain reaction amplification) or chemical synthesis. Isolated nucleic acid molecules include natural nucleic acid molecules and homologs thereof, including, but not limited to, natural allelic variants and modified nucleic acid molecules in which nucleotides have been inserted, deleted, substituted, or inverted in such a manner that such modifications do not substantially interfere with the nucleic acid molecule's ability to control cell growth or encode a protein that controls cell growth.

It should also be appreciated that reference to an isolated nucleic acid molecule does not necessarily reflect the extent of purity of the nucleic acid molecule. Nucleic acid molecules can be isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the nucleic acid molecule will be obtained substantially free of other nucleic acid sequences, generally being at least about 50%, and usually at least about 90% pure. Although the phrase "nucleic acid molecule" primarily refers to the physical nucleic acid molecule and the phrase "nucleic acid sequence" primarily refers to the sequence of nucleotides on the nucleic acid molecule, the two phrases can be used interchangeably.

According to the invention, reference to an "isolated nucleic acid molecule" refers to a nucleic acid molecule that is the size of or is smaller than a gene. Thus, an isolated nucleic acid molecule does not encompass isolated total genomic DNA or an isolated chromosome. As used herein, the term "gene" has the meaning that is well known in the art, that is, a nucleic acid sequence that includes the translated sequences that code for a protein ("exons") and the untranslated intervening sequences ("introns"), and any regulatory elements necessary to transcribe and/or translate the protein. Included in the invention are nucleic acid molecules that are less than a full-length gene or less than a full-length coding sequence, such as fragments of a gene or coding sequence comprising, consisting essentially of, or consisting of, for example, a fragment of any of the nucleic acid sequences for target genes described in the present invention. A coding sequence can include genomic DNA without introns, cDNA or RNA that encodes a protein. An isolated nucleic acid molecule can also include a specified nucleic acid sequence flanked

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by (i.e., at the 5' and/or the 3' end of the sequence) additional nucleic acids that do not normally flank the specified nucleic acid sequence in nature (i.e., are heterologous sequences).

In one embodiment, an isolated nucleic acid molecule useful in a method of the present invention is produced using recombinant DNA technology (e.g., polymerase chain reaction (PCR) amplification, cloning) or chemical synthesis. A nucleic acid molecule homologue can be produced using a number of methods known to those skilled in the art (see, for example, Sambrook et al., ibid.). For example, nucleic acid molecules can be modified using a variety of techniques including, but not limited to, classical mutagenesis techniques and recombinant DNA techniques, such as site-directed mutagenesis, chemical treatment of a nucleic acid molecule to induce mutations, restriction enzyme cleavage of a nucleic acid fragment, ligation of nucleic acid fragments, PCR amplification and/or mutagenesis of selected regions of a nucleic acid sequence, synthesis of oligonucleotide mixtures and ligation of mixture groups to "build" a mixture of nucleic acid molecules and combinations thereof. Nucleic acid molecule homologues can be selected from a mixture of modified nucleic acids by screening for the function of the protein encoded by the nucleic acid and/or by hybridization with a wild-type gene.

The term isolated nucleic acid molecule does not necessarily connote any specific minimum length unless set forth by reference to a minimum number of nucleotides or by a function of the nucleic acid molecule. The minimum size of a nucleic acid molecule of the present invention is generally a size sufficient to encode a protein having the desired biological activity, a size sufficient to inhibit the expression and/or activity of a target as described herein (e.g., as in a GSE), a size sufficient for use in a screening assay or diagnostic method of the invention, or a size sufficient to form a probe or oligonucleotide primer that is capable of forming a stable hybrid with the complementary sequence of a nucleic acid molecule. As such, the size of a nucleic acid molecule of the present invention can be dependent on nucleic acid composition and percent homology or identity between the nucleic acid molecule and complementary sequence as well as upon hybridization conditions per se (e.g., temperature, salt concentration, and formamide concentration) and the intended use of the nucleic acid molecule. The minimal size of a nucleic acid molecule that is used as an oligonucleotide primer or as a probe is typically at least about 12 to about 15 nucleotides in length if the nucleic acid molecules are GCrich and at least about 15 to about 18 bases in length if they are AT-rich. There is no

limit, other than a practical limit, on the maximal size of a nucleic acid molecule of the present invention, in that the nucleic acid molecule can include a fragment of a gene, a portion of a protein encoding sequence, or a nucleic acid sequence encoding a full-length protein (including a complete gene).

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Some embodiments of the present invention may include the production and/or use of a recombinant nucleic acid molecule comprising a recombinant vector and a nucleic acid molecule comprising a nucleic acid sequence encoding a gene or fragment thereof as described herein. According to the present invention, a recombinant vector is an engineered (i.e., artificially produced) nucleic acid molecule that is used as a tool for manipulating a nucleic acid sequence of choice and for introducing such a nucleic acid sequence into a host cell. The recombinant vector is therefore suitable for use in cloning, sequencing, and/or otherwise manipulating the nucleic acid sequence of choice, such as by expressing and/or delivering the nucleic acid sequence of choice into a host cell to form a recombinant cell. Such a vector typically contains heterologous nucleic acid sequences, that is nucleic acid sequences that are not naturally found adjacent to nucleic acid sequence to be cloned or delivered, although the vector can also contain regulatory nucleic acid sequences (e.g., promoters, untranslated regions) which are naturally found adjacent to nucleic acid molecules of the present invention or which are useful for expression of the nucleic acid molecules of the present invention (discussed in detail below). The vector can be either RNA or DNA, either prokaryotic or eukaryotic, and typically is a plasmid. The vector can be maintained as an extrachromosomal element (e.g., a plasmid) or it can be integrated into the chromosome of a recombinant organism (e.g., a microbe or a plant). The entire vector can remain in place within a host cell, or under certain conditions, the plasmid DNA can be deleted, leaving behind the nucleic acid molecule of the present invention. The integrated nucleic acid molecule can be under chromosomal promoter control, under native or plasmid promoter control, or under a combination of several promoter controls. Single or multiple copies of the nucleic acid molecule can be integrated into the chromosome. A recombinant vector of the present invention can contain at least one selectable marker.

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In one embodiment, a recombinant vector used in a recombinant nucleic acid molecule of the present invention is an expression vector. As used herein, the phrase "expression vector" is used to refer to a vector that is suitable for production of an encoded product (e.g., a protein of interest). In this embodiment, a nucleic acid sequence

encoding the product to be produced is inserted into the recombinant vector to produce a recombinant nucleic acid molecule. The nucleic acid sequence encoding the protein to be produced is inserted into the vector in a manner that operatively links the nucleic acid sequence to regulatory sequences in the vector that enable the transcription and translation of the nucleic acid sequence within the recombinant host cell.

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In another embodiment, a recombinant vector used in a recombinant nucleic acid molecule of the present invention is a targeting vector. As used herein, the phrase "targeting vector" is used to refer to a vector that is used to deliver a particular nucleic acid molecule into a recombinant host cell, wherein the nucleic acid molecule is used to delete or inactivate an endogenous gene within the host cell or microorganism (*i.e.*, used for targeted gene disruption or knock-out technology). Such a vector may also be known in the art as a "knock-out" vector. In one aspect of this embodiment, a portion of the vector, but more typically, the nucleic acid molecule inserted into the vector (*i.e.*, the insert), has a nucleic acid sequence that is homologous to a nucleic acid sequence of a target gene in the host cell (*i.e.*, a gene which is targeted to be deleted or inactivated). The nucleic acid sequence of the vector insert is designed to bind to the target gene such that the target gene and the insert undergo homologous recombination, whereby the endogenous target gene is deleted, inactivated or attenuated (*i.e.*, by at least a portion of the endogenous target gene being mutated or deleted).

Typically, a recombinant nucleic acid molecule includes at least one nucleic acid molecule of the present invention operatively linked to one or more expression control sequences, including transcription control sequences and translation control sequences. As used herein, the phrase "recombinant molecule" or "recombinant nucleic acid molecule" primarily refers to a nucleic acid molecule or nucleic acid sequence operatively linked to an expression control sequence, but can be used interchangeably with the phrase "nucleic acid molecule", when such nucleic acid molecule is a recombinant molecule as discussed herein. According to the present invention, the phrase "operatively linked" refers to linking a nucleic acid molecule to an expression control sequence (e.g., a transcription control sequence and/or a translation control sequence) in a manner such that the molecule is able to be expressed when transfected (i.e., transformed, transduced, transfected, conjugated or conduced) into a host cell. Transcription control sequences are sequences that control the initiation, elongation, or termination of transcription. Particularly important transcription control sequences are those that control transcription

initiation, such as promoter, enhancer, operator and repressor sequences. Suitable transcription control sequences include any transcription control sequence that can function in a host cell or organism into which the recombinant nucleic acid molecule is to be introduced.

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According to the present invention, the term "transfection" is used to refer to any method by which an exogenous nucleic acid molecule (i.e., a recombinant nucleic acid The term "transformation" can be used molecule) can be inserted into a cell. interchangeably with the term "transfection" when such term is used to refer to the introduction of nucleic acid molecules into microbial cells. In microbial systems, the term "transformation" is used to describe an inherited change due to the acquisition of exogenous nucleic acids by the microorganism and is essentially synonymous with the term "transfection." However, in animal cells, transformation has acquired a second meaning that can refer to changes in the growth properties of cells in culture (described above) after they become cancerous, for example. Therefore, to avoid confusion, the term "transfection" is preferably used with regard to the introduction of exogenous nucleic acids into animal cells, including human cells, and is used herein to generally encompass transfection of animal cells and transformation of microbial cells, to the extent that the terms pertain to the introduction of exogenous nucleic acids into a cell. Therefore, transfection techniques include, but are not limited to, transformation, chemical treatment of cells, particle bombardment, electroporation, microinjection, lipofection, adsorption, infection and protoplast fusion.

A recombinant cell is preferably produced by transforming a host cell with one or more recombinant molecules, each comprising one or more nucleic acid molecules operatively linked to an expression vector containing one or more expression control sequences.

"Hybridization" has the meaning that is well known in the art, that is, the formation of a duplex structure by two single-stranded nucleic acids due to complementary base pairing. Hybridization can occur between exactly complementary nucleic acid strands or between nucleic acid strands that contain some regions of mismatch. As used herein, reference to hybridization conditions refers to standard hybridization conditions under which nucleic acid molecules are used to identify similar nucleic acid molecules. Such standard conditions are disclosed, for example, in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Labs

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Press, 1989. Sambrook et al., *ibid.*, is incorporated by reference herein in its entirety (*see* specifically, pages 9.31-9.62). In addition, formulae to calculate the appropriate hybridization and wash conditions to achieve hybridization permitting varying degrees of mismatch of nucleotides are disclosed, for example, in Meinkoth et al., 1984, *Anal. Biochem.* 138, 267-284; Meinkoth et al., *ibid.*, is incorporated by reference herein in its entirety. "Stringent hybridization" has a meaning well-established in the art, that is, hybridization performed at a salt concentration of no more than 1M and a temperature of at least 25 degrees Celsius. For example, conditions of 5X SSPE (750 mM NaCl, 50 mM Sodium Phosphate, 5 mM EDTA, pH 7.4) and a temperature of 55 degrees to 60 degrees Celsius are suitable. For example, in one embodiment, "moderately stringent conditions" can be defined as hybridizations carried out as described above, followed by washing in 0.2X SSC and 0.1% SDS at 42 degrees Celsius (Ausubel *et al.*, 1989, *Current Protocols for Molecular Biology, ibid.*).

More particularly, moderate stringency hybridization and washing conditions, as referred to herein, refer to conditions which permit isolation of nucleic acid molecules having at least about 70% nucleic acid sequence identity with the nucleic acid molecule being used to probe in the hybridization reaction (i.e., conditions permitting about 30% or less mismatch of nucleotides). High stringency hybridization and washing conditions, as referred to herein, refer to conditions which permit isolation of nucleic acid molecules having at least about 80% nucleic acid sequence identity with the nucleic acid molecule being used to probe in the hybridization reaction (i.e., conditions permitting about 20% or Very high stringency hybridization and washing less mismatch of nucleotides). conditions, as referred to herein, refer to conditions which permit isolation of nucleic acid molecules having at least about 90% nucleic acid sequence identity with the nucleic acid molecule being used to probe in the hybridization reaction (i.e., conditions permitting about 10% or less mismatch of nucleotides). As discussed above, one of skill in the art can use the formulae in Meinkoth et al., ibid. to calculate the appropriate hybridization and wash conditions to achieve these particular levels of nucleotide mismatch. Such conditions will vary, depending on whether DNA:RNA or DNA:DNA hybrids are being formed. Calculated melting temperatures for DNA:DNA hybrids are 10°C less than for DNA:RNA hybrids. In particular embodiments, stringent hybridization conditions for DNA:DNA hybrids include hybridization at an ionic strength of 6X SSC (0.9 M Na<sup>+</sup>) at a temperature of between about 20°C and about 35°C (low stringency), more preferably,

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between about 28°C and about 42°C (more stringent), and even more preferably, between about 35°C and about 45°C (even more stringent), with appropriate wash conditions. In particular embodiments, stringent hybridization conditions for DNA:RNA hybrids include hybridization at an ionic strength of 6X SSC (0.9 M Na<sup>+</sup>) at a temperature of between about 30°C and about 45°C, more preferably, between about 38°C and about 50°C, and even more preferably, between about 45°C and about 55°C, with similarly stringent wash conditions. These values are based on calculations of a melting temperature for molecules larger than about 100 nucleotides, 0% formamide and a G + C content of about 40%. Alternatively, Tm can be calculated empirically as set forth in Sambrook et al., supra, pages 9.31 to 9.62. In general, the wash conditions should be as stringent as possible, and should be appropriate for the chosen hybridization conditions. For example, hybridization conditions can include a combination of salt and temperature conditions that are approximately 20-25°C below the calculated T<sub>m</sub> of a particular hybrid, and wash conditions typically include a combination of salt and temperature conditions that are approximately 12-20°C below the calculated T<sub>m</sub> of the particular hybrid. One example of hybridization conditions suitable for use with DNA:DNA hybrids includes a 2-24 hour hybridization in 6X SSC (50% formamide) at about 42°C, followed by washing steps that include one or more washes at room temperature in about 2X SSC, followed by additional washes at higher temperatures and lower ionic strength (e.g., at least one wash as about 37°C in about 0.1X-0.5X SSC, followed by at least one wash at about 68°C in about 0.1X-0.5X SSC).

In one embodiment of the present invention, any amino acid sequence described herein can be produced with from at least one, and up to about 20, additional heterologous amino acids flanking each of the C- and/or N-terminal ends of the specified amino acid sequence. The resulting protein or polypeptide can be referred to as "consisting essentially of" the specified amino acid sequence. According to the present invention, the heterologous amino acids are a sequence of amino acids that are not naturally found (i.e., not found in nature, in vivo) flanking the specified amino acid sequence, or that are not related to the function of the specified amino acid sequence, or that would not be encoded by the nucleotides that flank the naturally occurring nucleic acid sequence encoding the specified amino acid sequence as it occurs in the gene, if such nucleotides in the naturally occurring sequence were translated using standard codon

usage for the organism from which the given amino acid sequence is derived. Similarly, the phrase "consisting essentially of", when used with reference to a nucleic acid sequence herein, refers to a nucleic acid sequence encoding a specified amino acid sequence that can be flanked by from at least one, and up to as many as about 60, additional heterologous nucleotides at each of the 5' and/or the 3' end of the nucleic acid sequence encoding the specified amino acid sequence. The heterologous nucleotides are not naturally found (i.e., not found in nature, in vivo) flanking the nucleic acid sequence encoding the specified amino acid sequence as it occurs in the natural gene or do not encode a protein that imparts any additional function to the protein or changes the function of the protein having the specified amino acid sequence.

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As discussed above, one embodiment of the present invention relates to methods for identifying compounds that induce or increase or upregulate apoptosis in a cell by inhibiting genes or gene products involved in the control of cell growth. Once a gene has been identified as a target for supporting cell growth, an assay can be used for screening and selecting a chemical compound or a biological compound having activity as an antitumor therapeutic based on the ability of the compound to down-regulate expression of the gene or inhibit activity of its gene product. Reference herein to inhibiting a target, can refer to one or both of inhibiting expression of a target gene and inhibiting the translation and/or activity of its corresponding expression product. Such a compound can be referred to herein as therapeutic compound. For example, a cell line that naturally expresses the gene of interest or has been transfected with the gene or other recombinant nucleic acid molecule encoding the protein of interest is incubated with various compounds, also referred to as candidate compounds, test compounds, or putative regulatory compounds. A reduction of the expression of the gene of interest or an inhibition of the activities of its encoded product (e.g., biological activity, which can include the involvement of the protein in the protection of the cell from apoptotic processes) may be used to identify a therapeutic compound. Therapeutic compounds identified in this manner can then be retested, if desired, in other assays to confirm their activities against cellular apoptotic processes.

In general, the biological activity or biological action of a protein refers to any function(s) exhibited or performed by the protein that is ascribed to the naturally occurring form of the protein as measured or observed *in vivo* (*i.e.*, in the natural physiological environment of the protein) or *in vitro* (*i.e.*, under laboratory conditions).

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Modifications, activities or interactions which result in a decrease in protein expression or a decrease in the activity of the protein, can be referred to as inactivation (complete or partial), down-regulation, reduced action, or decreased action or activity of a protein. Similarly, modifications, activities or interactions which result in an increase in protein expression or an increase in the activity of the protein, can be referred to as amplification, overproduction, activation, enhancement, up-regulation or increased action of a protein. The biological activity of a protein according to the invention can be measured or evaluated using any assay for the biological activity of the protein as known in the art. Such assays can include, but are not limited to, binding assays, assays to determine internalization of the protein and/or associated proteins, enzyme assays, cell signal transduction assays (e.g., phosphorylation assays), and/or assays for determining downstream cellular events that result from activation or binding of the cell surface protein (e.g., expression of downstream genes, production of various biological mediators, etc.). The assay can also measure the ability of the protein to contribute to the regulation of apoptosis in a cell. Such assays are described in detail herein. According to the present invention, a biologically active fragment or homologue of a gene or protein maintains the ability to be useful in a method of the present invention. Therefore, the biologically active fragment or homologue maintains the ability to be used to identify regulators (e.g., inhibitors) of the native gene or protein when, for example, the biologically active fragment or homologue is expressed by a cell. Therefore, the biologically active fragment or homologue has a structure that is sufficiently similar to the structure of the native gene or protein that a regulatory compound can be identified by its ability to bind to and/or regulate the expression or activity of the fragment or homologue in a manner consistent with the regulation of the native gene or protein.

compounds such as antibodies, products of peptide libraries, and products of chemical combinatorial libraries. Compounds may also be identified using rational drug design relying on the structure of the product of a gene. Such methods are known to those of skill in the art and involve the use of three-dimensional imaging software programs. For example, various methods of drug design, useful to design or select mimetics or other therapeutic compounds useful in the present invention are disclosed in Maulik et al.,

Compounds to be screened in the methods of the invention include known organic

1997, Molecular Biotechnology: Therapeutic Applications and Strategies, Wiley-Liss,

Inc., which is incorporated herein by reference in its entirety.

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As used herein, a mimetic refers to any peptide or non-peptide compound that is able to mimic the biological action of a naturally occurring peptide, often because the mimetic has a basic structure that mimics the basic structure of the naturally occurring peptide and/or has the salient biological properties of the naturally occurring peptide. Mimetics can include, but are not limited to: peptides that have substantial modifications from the prototype such as no side chain similarity with the naturally occurring peptide (such modifications, for example, may decrease its susceptibility to degradation); anti-idiotypic and/or catalytic antibodies, or fragments thereof; non-proteinaceous portions of an isolated protein (e.g., carbohydrate structures); or synthetic or natural organic molecules, including nucleic acids and drugs identified through combinatorial chemistry, for example. Such mimetics can be designed, selected and/or otherwise identified using a variety of methods known in the art.

A mimetic can be obtained, for example, from molecular diversity strategies (a combination of related strategies allowing the rapid construction of large, chemically diverse molecule libraries), libraries of natural or synthetic compounds, in particular from chemical or combinatorial libraries (*i.e.*, libraries of compounds that differ in sequence or size but that have the similar building blocks) or by rational, directed or random drug design. See for example, Maulik et al., supra.

In a molecular diversity strategy, large compound libraries are synthesized, for example, from peptides, oligonucleotides, carbohydrates and/or synthetic organic molecules, using biological, enzymatic and/or chemical approaches. The critical parameters in developing a molecular diversity strategy include subunit diversity, molecular size, and library diversity. The general goal of screening such libraries is to utilize sequential application of combinatorial selection to obtain high-affinity ligands for a desired target, and then to optimize the lead molecules by either random or directed design strategies. Methods of molecular diversity are described in detail in Maulik, et al., *ibid*.

Maulik et al. also disclose, for example, methods of directed design, in which the user directs the process of creating novel molecules from a fragment library of appropriately selected fragments; random design, in which the user uses a genetic or other algorithm to randomly mutate fragments and their combinations while simultaneously applying a selection criterion to evaluate the fitness of candidate ligands; and a grid-based approach in which the user calculates the interaction energy between three dimensional

receptor structures and small fragment probes, followed by linking together of favorable probe sites.

As used herein, the term "test compound", "putative inhibitory compound" or "putative regulatory compound" refers to compounds having an unknown or previously unappreciated regulatory activity in a particular process. As such, the term "identify" with regard to methods to identify compounds is intended to include all compounds, the usefulness of which as a regulatory compound for the purposes of inhibiting cell growth is determined by a method of the present invention.

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In one embodiment of the invention, inhibitors of cell growth are identified by exposing a target gene to a test compound; measuring the expression of a target; and selecting a compound that down-regulates (reduces, decreases, inhibits, blocks) the expression of the target. For example, the putative inhibitor can be exposed to a cell that expresses the target gene (endogenously or recombinantly). A preferred cell to use in an assay includes a mammalian cell that either naturally expresses the target gene or has been transformed with a recombinant form of the target gene, such as a recombinant nucleic acid molecule comprising a nucleic acid sequence encoding the target protein or a useful fragment thereof. Methods to determine expression levels of a gene are well known in the art.

The conditions under which a cell, cell lysate, nucleic acid molecule or protein of the present invention is exposed to or contacted with a putative regulatory compound, such as by mixing, are any suitable culture or assay conditions. In the case of a cell-based assay, the conditions include an effective medium in which the cell can be cultured or in which the cell lysate can be evaluated in the presence and absence of a putative regulatory compound. Cells of the present invention can be cultured in a variety of containers including, but not limited to, tissue culture flasks, test tubes, microtiter dishes, and petri plates. Culturing is carried out at a temperature, pH and carbon dioxide content appropriate for the cell. Such culturing conditions are also within the skill in the art. Cells are contacted with a putative regulatory compound under conditions which take into account the number of cells per container contacted, the concentration of putative regulatory compound(s) administered to a cell, the incubation time of the putative regulatory compound with the cell, and the concentration of compound administered to a cell. Determination of effective protocols can be accomplished by those skilled in the art based on variables such as the size of the container, the volume of liquid in the container,

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conditions known to be suitable for the culture of the particular cell type used in the assay, and the chemical composition of the putative regulatory compound (i.e., size, charge etc.) being tested. A preferred amount of putative regulatory compound(s) can comprise between about 1 nM to about 10 mM of putative regulatory compound(s) per well of a 96-well plate.

As used herein, the term "expression", when used in connection with detecting the expression of a target of the present invention, can refer to detecting transcription of the target gene and/or to detecting translation of the target protein encoded by the target gene. To detect expression of a target refers to the act of actively determining whether a target is expressed or not. This can include determining whether the target expression is upregulated as compared to a control, downregulated as compared to a control, or unchanged as compared to a control. Therefore, the step of detecting expression does not require that expression of the target actually is upregulated or downregulated, but rather, can also include detecting that the expression of the target has not changed (i.e., detecting no expression of the target or no change in expression of the target). Expression of transcripts and/or proteins is measured by any of a variety of known methods in the art. For RNA expression, methods include but are not limited to: extraction of cellular mRNA and Northern blotting using labeled probes that hybridize to transcripts encoding all or part of one or more of the genes of this invention; amplification of mRNA expressed from one or more of the genes of this invention using gene-specific primers, polymerase chain reaction (PCR), and reverse transcriptase-polymerase chain reaction (RT-PCR), followed by quantitative detection of the product by any of a variety of means; extraction of total RNA from the cells, which is then labeled and used to probe cDNAs or oligonucleotides encoding all or part of the genes of this invention, arrayed on any of a variety of surfaces; in situ hybridization; and detection of a reporter gene. The term "quantifying" or "quantitating" when used in the context of quantifying transcription levels of a gene can refer to absolute or to relative quantification. Absolute quantification may be accomplished by inclusion of known concentration(s) of one or more target nucleic acids and referencing the hybridization intensity of unknowns with the known target nucleic acids (e.g. through generation of a standard curve). Alternatively, relative quantification can be accomplished by comparison of hybridization signals between two or more genes, or between two or more treatments to quantify the changes in hybridization intensity and, by implication, transcription level.

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In a preferred embodiment, the expression of the target gene is measured by the polymerase chain reaction. In another embodiment, the expression of the target gene is measured using polyacrylamide gel analysis, chromatography or spectroscopy.

In another preferred embodiment, the expression of the target gene is measured by measuring the production of the encoded protein (measuring translation of the protein). Measurement of translation of a protein includes any suitable method for detecting and/or measuring proteins from a cell or cell extract. Such methods include, but are not limited to, immunoblot (e.g., Western blot), enzyme-linked immunosorbant assay (ELISA), immunoprecipitation, immunohistochemistry, (RIA), radioimmunoassay sorting (FACS) activated cell fluorescence immunofluorescence, Particularly preferred methods for detection of immunofluorescence microscopy. proteins include any single-cell assay, including immunohistochemistry and immunofluorescence assays. For example, one can use a detection agent, such as an antibody that specifically recognizes (selectively binds to) the protein encoded by the gene. Such methods are well known in the art.

Designing a compound for testing in a method of the present invention can include creating a new chemical compound or searching databases of libraries of known compounds (e.g., a compound listed in a computational screening database containing three dimensional structures of known compounds). Designing can also be performed by simulating chemical compounds having substitute moieties at certain structural features. The step of designing can include selecting a chemical compound based on a known function of the compound. A preferred step of designing comprises computational screening of one or more databases of compounds in which the three dimensional structure of the compound is known and is interacted (e.g., docked, aligned, matched, interfaced) with the three dimensional structure of a target by computer (e.g. as described by Humblet and Dunbar, Animal Reports in Medicinal Chemistry, vol. 28, pp. 275-283, 1993, M Venuti, ed., Academic Press). Methods to synthesize suitable chemical compounds are known to those of skill in the art and depend upon the structure of the chemical being synthesized. Methods to evaluate the bioactivity of the synthesized compound depend upon the bioactivity of the compound (e.g., inhibitory or stimulatory).

Accordingly, in another embodiment of the invention, therapeutic compounds can be selected by determining the three-dimensional structure of a target; and determining or designing the three-dimensional structure of a therapeutic or regulatory compound by rational drug design or detecting a structure that interacts with the target structure from a library of known compound structures. Preferably, the structure of the therapeutic compound is determined using computer software capable of modeling the interaction of a therapeutic compound with the target. One of skill in the art can select the appropriate three-dimensional structure, therapeutic or regulatory compound, and analytical software based on the identity of the target.

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For example, suitable candidate chemical compounds can align to a subset of residues described for a target site. Preferably, a candidate chemical compound comprises a conformation that promotes the formation of covalent or noncovalent crosslinking between the target site and the candidate chemical compound. Preferably, a candidate chemical compound binds to a surface adjacent to a target site to provide an additional site of interaction in a complex. When designing an antagonist, for example, the antagonist should bind with sufficient affinity to the binding site or to substantially prohibit a ligand (i.e., a molecule that specifically binds to the target site) from binding to a target area. It will be appreciated by one of skill in the art that it is not necessary that the complementarity between a candidate chemical compound and a target site extend over all residues specified here in order to inhibit or promote binding of a ligand.

In general, the design of a chemical compound possessing stereochemical complementarity can be accomplished by techniques that optimize, chemically or geometrically, the "fit" between a chemical compound and a target site. Such techniques are disclosed by, for example, Sheridan and Venkataraghavan, *Acc. Chem Res.*, vol. 20, p. 322, 1987: Goodford, *J. Med. Chem.*, vol. 27, p. 557, 1984; Beddell, *Chem. Soc. Reviews*, vol. 279, 1985; Hol, *Angew. Chem.*, vol. 25, p. 767, 1986; and Verlinde and Hol, *Structure*, vol. 2, p. 577, 1994, each of which are incorporated by this reference herein in their entirety.

As another example, a "geometric approach" is used. In a geometric approach, the number of internal degrees of freedom (and the corresponding local minima in the molecular conformation space) is reduced by considering only the geometric (hard sphere) interactions of two rigid bodies, where one body (the active site) contains "pockets" or "grooves" that form binding sites for the second body (the complementing molecule, such as a ligand). The geometric approach is described by Kuntz et al., *J. Mol. Biol.*, vol. 161, p. 269, 1982, which is incorporated by this reference herein in its entirety. The algorithm for chemical compound design can be implemented using the software

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program DOCK Package, Version 1.0 (available from the Regents of the University of California). Pursuant to the Kuntz algorithm, the shape of the cavity or groove on the surface of a structure at a binding site or interface is defined as a series of overlapping spheres of different radii. One or more extant databases of crystallographic data (e.g., the Cambridge Structural Database System maintained by University Chemical Laboratory, Cambridge University, Lensfield Road, Cambridge CB2 IEW, U.K.) or the Protein Data Bank maintained by Brookhaven National Laboratory, is then searched for chemical compounds that approximate the shape thus defined. Chemical compounds identified by the geometric approach can be modified to satisfy criteria associated with chemical complementarity, such as hydrogen bonding, ionic interactions or Van der Waals interactions.

As yet another example, one can determine the interaction of chemical groups ("probes") with an active site at sample positions within and around a binding site or interface, resulting in an array of energy values from which three dimensional contour surfaces at selected energy levels can be generated. This method is referred to herein as a "chemical-probe approach." The chemical-probe approach to the design of a chemical compound useful of the present invention is described by, for example, Goodford, *J. Med. Chem.*, vol. 28, p. 849, 1985, which is incorporated by this reference herein in its entirety, and is implemented using an appropriate software package, including for example, GRID (available from Molecular Discovery Ltd., Oxford OX2 9LL, U.K.). The chemical prerequisites for a site-complementing molecule can be identified at the outset, by probing the active site of a protein with different chemical probes, *e.g.*, water, a methyl group, an amine nitrogen, a carboxyl oxygen and/or a hydroxyl. Preferred sites for interaction between an active site and a probe are determined. Putative complementary chemical compounds can be generated using the resulting three dimensional patterns of such sites.

Candidate compounds identified or designed by the above-described methods can be synthesized using techniques known in the art, and depending on the type of compound. Synthesis techniques for the production of non-protein compounds, including organic and inorganic compounds are well known in the art. For example, for smaller peptides, chemical synthesis methods are preferred. For example, such methods include well known chemical procedures, such as solution or solid-phase peptide synthesis, or semi-synthesis in solution beginning with protein fragments coupled through

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conventional solution methods. Such methods are well known in the art and may be found in general texts and articles in the area such as: Merrifield, 1997, *Methods Enzymol*. 289:3-13; Wade et al., 1993, *Australas Biotechnol*. 3(6):332-336; Wong et al., 1991, *Experientia* 47(11-12):1123-1129; Carey et al., 1991, *Ciba Found Symp*. 158:187-203; Plaue et al., 1990, *Biologicals* 18(3):147-157; Bodanszky, 1985, *Int. J. Pept. Protein Res*. 25(5):449-474; or H. Dugas and C. Penney, BIOORGANIC CHEMISTRY, (1981) at pages 54-92, all of which are incorporated herein by reference in their entirety. For example, peptides may be synthesized by solid-phase methodology utilizing a commercially available peptide synthesizer and synthesis cycles supplied by the manufacturer. One skilled in the art recognizes that the solid phase synthesis could also be accomplished using the FMOC strategy and a TFA/scavenger cleavage mixture. A compound that is a protein or peptide can also be produced using recombinant DNA technology and methods standard in the art, particularly if larger quantities of a protein are desired.

In still another embodiment of the invention, inhibitors of cell growth are identified by exposing a target to a candidate compound; measuring the binding of the candidate compound to the target; and selecting a compound that binds to the target at a desired concentration, affinity, or avidity. In a preferred embodiment, the assay is performed under conditions conducive to promoting the interaction or binding of the compound to the target. One of skill in the art can determine such conditions based on the target and the compound being used in the assay. In one embodiment, a BIAcore machine can be used to determine the binding constant of a complex between the target protein (a protein encoded by the target gene) and a natural ligand in the presence and absence of the candidate compound. For example, the target protein or a ligand binding fragment thereof can be immobilized on a substrate. A natural or synthetic ligand is contacted with the substrate to form a complex. The dissociation constant for the complex can be determined by monitoring changes in the refractive index with respect to time as buffer is passed over the chip (O'Shannessy et al. Anal. Biochem. 212:457-468 (1993); Schuster et al., Nature 365:343-347 (1993)). Contacting a candidate compound at various concentrations with the complex and monitoring the response function (e.g., the change in the refractive index with respect to time) allows the complex dissociation constant to be determined in the presence of the test compound and indicates whether the candidate compound is either an inhibitor or an agonist of the complex. Alternatively, the

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candidate compound can be contacted with the immobilized target protein at the same time as the ligand to see if the candidate compound inhibits or stabilizes the binding of the ligand to the target protein.

Other suitable assays for measuring the binding of a candidate compound to a target protein or for measuring the ability of a candidate compound to affect the binding of the target protein to another protein or molecule include, but are not limited to, immunosorbant assay (ELISA), immunoblot, enzyme-linked blot, plasmon (RIA), immunoprecipitation, surface resonance, radioimmunoassay chemiluminescence, fluorescent polarization, phosphorescence, immunohistochemical analysis, matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry, microcytometry, microarray, microscopy, fluorescence activated cell sorting (FACS), and flow cytometry. Other assays include those that are suitable for monitoring the effects of protein binding, including, but not limited to, cell-based assays such as: cytokine secretion assays, or intracellular signal transduction assays that determine, for example, protein or lipid phosphorylation, mediator release or intracellular Ca<sup>++</sup> mobilization.

In yet another embodiment, inhibitors of cellular growth are identified by exposing a target protein of the present invention (or a cell expressing the protein naturally or recombinantly) to a candidate compound and measuring the ability of the compound to inhibit (reduce, decrease, block) a biological activity of the protein. In one embodiment, the biological activity of a protein encoded by the target gene is measured by measuring the amount of product generated in a biochemical reaction mediated by the protein encoded by the target gene. In still another embodiment, the activity of the protein encoded by the target gene is measured by measuring the amount of substrate generated in a biochemical reaction mediated by the protein encoded by the target gene. In another embodiment, a biological activity is measured by measuring a specific event in a cell-based assay, such as release or secretion of a biological mediator or compound that is regulated by the activity of the target protein, measuring intracellular signal transduction assays that determine, for example, protein or lipid phosphorylation, mediator release or intracellular Ca<sup>++</sup> mobilization. Preferably, the activity of the protein is measured in the presence and absence of the candidate compound, or in the presence of another suitable control compound.

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In one embodiment of the invention, when the protein encoded by a target gene is an enzyme, a therapeutic compound is identified by exposing the enzyme encoded by a target gene to a test compound; measuring the activity of the enzyme encoded by the target gene in the presence and absence of the compound; and selecting a compound that down-regulates or inhibits the activity of the enzyme encoded by the target gene. Methods to measure enzymatic activity are well known to those skilled in the art and are selected based on the identity of the enzyme being tested. For example, if the enzyme is a kinase, phosphorylation assays can be used.

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In addition to methods for identifying and producing a biological compound that inhibits cell growth, the present invention includes methods known in the art that down-regulate expression or function of a target gene. For example, antisense RNA and DNA molecules may be used to directly block translation of mRNA encoded by these genes by binding to targeted mRNA and preventing protein translation. Polydeoxyribonucleotides can form sequence-specific triple helices by hydrogen bonding to specific complementary sequences in duplexed DNA to effect specific down-regulation of target gene expression. Formation of specific triple helices may selectively inhibit the replication or expression of a target gene by prohibiting the specific binding of functional trans-acting factors.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. Ribozyme action involves sequence specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. Within the scope of the invention are ribozyme embodiments including engineered hammerhead motif ribozyme molecules that specifically and efficiently catalyze endonucleolytic cleavage of RNA sequences. Antisense RNA molecules showing high-affinity binding to target sequences can also be used as ribozymes by addition of enzymatically active sequences known to those skilled in the art.

Polynucleotides to be used in triplex helix formation should be single-stranded and composed of deoxynucleotides. The base composition of these polynucleotides must be designed to promote triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of either purines or pyrimidines to be present on one strand of a duplex. Polynucleotide sequences may be pyrimidine-based, which will result in TAT and CGC triplets across the three associated strands of the resulting triple helix. The pyrimidine-rich polynucleotides provide base complementarity to a purine-rich region of a single strand of the duplex in a parallel orientation to that strand. In addition,

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polynucleotides may be chosen that are purine-rich, for example, containing a stretch of G residues. These polynucleotides will form a triple helix with a DNA duplex that is rich in GC pairs, in which the majority of the purine residues are located on a single strand of the targeted duplex, resulting in GGC triplets across the three strands in the triplex.

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Alternatively, sequences that can be targeted for triple helix formation can be increased by creating a so-called "switchback" polynucleotide. Switchback polynucleotides are synthesized in an alternating 5'-3', 3'-5' manner, so that they base pair with first one strand of a duplex and then the other, eliminating the necessity for a sizeable stretch of either purines or pyrimidines to be present on one strand of a duplex.

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Both antisense RNA and DNA molecules, and ribozymes of the invention may be prepared by any method known in the art. These include techniques for chemically synthesizing polynucleotides well known in the art such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA sequences encoding the antisense RNA molecule. Such DNA sequences may be incorporated into a wide variety of vectors that incorporate suitable RNA polymerase promoters such as the T7 or SP6 polymerase promoters. Alternatively, antisense cDNA constructs that synthesize antisense RNA constitutively or inducibly, depending on the promoter used, can be introduced stably into host cells.

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Various modifications to the nucleic acid molecules may be introduced as a means of increasing intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences of ribonucleotides or deoxyribonucleotides to the 5' or 3' ends of the molecule or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the oligodeoxyribonucleotide backbone.

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Preferably, methods used to identify therapeutic compounds are customized for each target gene or product. If the target product is an enzyme, then the enzyme will be expressed in cell culture and purified. The enzyme will then be screened *in vitro* against therapeutic compounds to look for inhibition of that enzymatic activity. If the target is a non-catalytic protein, then it will also be expressed and purified. Therapeutic compounds will then be tested for their ability to prevent, for example, the binding of a site-specific antibody or a target-specific ligand to the target product.

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In a preferred embodiment, therapeutic compounds that bind to target products are identified, then those compounds can be further tested in biological assays that test for

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characteristics such as apoptosis, tumor suppressor status (e.g., p53 status), tumor cell growth and any other customary measure of anti-cancer activity.

In one embodiment of the invention, a therapeutic compound is not toxic to a human host cell. In another embodiment, the therapeutic compound is cytostatic or cytotoxic.

In one embodiment of the invention, a pharmaceutical composition is prepared from a therapeutically-effective amount of a therapeutic compound of the invention and a pharmaceutically-acceptable carrier. Pharmaceutically-acceptable carriers are well known to those with skill in the art. The pharmaceutical compositions of the present invention can be manufactured in a manner that is itself known, e.g., by means of a conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping or lyophilizing processes. As used herein, a pharmaceutically acceptable carrier refers to any substance suitable for delivering a therapeutic composition useful in the method of the present invention to a suitable in vivo or ex vivo site. Pharmaceutical compositions for use in accordance with the present invention thus can be formulated in conventional manner using one or more physiologically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Proper formulation is dependent upon the route of administration chosen.

For injection, the compounds of the invention can be formulated in appropriate aqueous solutions, such as physiologically compatible buffers such as Hanks's solution, Ringer's solution, or physiological saline buffer. For transmucosal and transcutaneous administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art. For oral administration, the compounds can be formulated readily by combining the active compounds with pharmaceutically acceptable carriers well known in the art. Such carriers enable the compounds of the invention to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for oral ingestion by a patient to be treated. For administration by inhalation, the compounds for use according to the present invention are conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebuliser, with the use of a suitable propellant, e.g., dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. The compounds can be formulated for parenteral

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administration by injection, e.g., by bolus injection or continuous infusion. Formulations for injection can be presented in unit dosage form, e.g., in ampoules or in multi-dose containers, with an added preservative. The compounds can also be formulated in rectal compositions such as suppositories or retention enemas, e.g., containing conventional suppository bases such as cocoa butter or other glycerides.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions can be used, which can optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments can be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

In addition to the formulations described previously, the compounds can also be formulated as a depot preparation. Such long acting formulations can be administered by implantation (for example subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compounds can be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

According to the present invention, an effective administration protocol (i.e., administering a composition of the present invention in an effective manner) comprises suitable dose parameters and modes of administration that result in delivery of the compound or composition to a patient or to a target site, cell or tissue in the patient, and subsequent inhibition of the growth of the target cell, preferably so that the patient obtains some measurable, observable or perceived benefit from such administration. In some situations, where the target cell population is accessible for sampling, effective dose parameters can be determined using methods as described herein for assessment of tumor growth. Such methods include removing a sample of the target cell population from the patient prior to and after the compound or composition is administered, and measuring changes expression or biological activity of a target, as well as measuring inhibition of the growth of the cell. Alternatively, effective dose parameters can be determined by experimentation using in vitro cell cultures, in vivo animal models, and eventually, clinical trials if the patient is human. Effective dose parameters can be determined using methods standard in the art. Such methods include, for example, determination of

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survival rates, side effects (i.e., toxicity) and progression or regression of disease. Compounds which exhibit high therapeutic indices are preferred. The dosage can vary within this range depending upon the dosage form employed and the route of administration utilized. The exact formulation, route of administration and dosage can be chosen by the individual physician in view of the patient's condition. (See, e.g. Fingl et al., 1975, in "The Pharmacological Basis of Therapeutics", Ch.1, p.1).

Dosage amount and interval can be adjusted individually to provide plasma levels of the active moiety which are sufficient to maintain the inhibitory effects. Usual patient dosages for systemic administration range from 100 - 2000 mg/day. Stated in terms of patient body surface areas, usual dosages range from 50 - 910 mg/m²/day. Usual average plasma levels should be maintained within 0.1-1000  $\mu$ M. In cases of local administration or selective uptake, the effective local concentration of the compound can not be related to plasma concentration.

The amount of composition administered will, of course, be dependent on the subject being treated, on the subject's body surface area, the severity of the affliction, the manner of administration and the judgment of the prescribing physician.

Suitable routes of administration can, for example, include oral, rectal, transmucosal, transcutaneous, or intestinal administration; parenteral delivery, including intramuscular, subcutaneous, intramedullary injections, as well as intrathecal, direct intraventricular, intravenous, intraperitoneal, intranasal, or intraocular injections. Alternatively, one can administer the compound in a local rather than systemic manner, for example, *via* injection of the compound directly into a specific tissue, often in a depot or sustained release formulation. Furthermore, one can administer the compound in a targeted drug delivery system, for example, in a liposome and/or conjugated with a cell-specific antibody. The liposomes and cell-specific antibody will be targeted to and taken up selectively by tumor cells.

Accordingly, a further embodiment of the invention is a method for inducing apoptosis in a cell by inhibiting a target of the present invention, *i.e.*, a target selected from the group consisting of any of the targets listed in Table 1 and/or represented by any of SEQ ID NOs:1-80. For example, this method can be conducted *in vivo* by administering to an individual an inhibitory or therapeutic compound as generally discussed herein. In addition, the method can be conducted *in vitro* or *ex vivo*.

A further embodiment of the present invention is a method for the diagnosis of a tumor or the monitoring of a tumor growth or regression or a tumor therapy in a patient. The methods include determining the level of a marker (also referred to as a biomarker) in a patient sample, wherein the marker is selected from any of the biomarkers listed in Table 1 or represented by any of SEQ ID NOs:1-80.

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The first step of this method of the present invention includes detecting the expression or biological activity of a biomarker in a test sample from a patient (also called a patient sample). Suitable methods of obtaining a patient sample are known to a person of skill in the art. A patient sample can include any bodily fluid or tissue from a patient that may contain tumor cells or proteins of tumor cells. More specifically, according to the present invention, the term "test sample" or "patient sample" can be used generally to refer to a sample of any type which contains cells or products that have been secreted from cells to be evaluated by the present method, including but not limited to, a sample of isolated cells, a tissue sample and/or a bodily fluid sample. According to the present invention, a sample of isolated cells is a specimen of cells, typically in suspension or separated from connective tissue which may have connected the cells within a tissue in vivo, which have been collected from an organ, tissue or fluid by any suitable method which results in the collection of a suitable number of cells for evaluation by the method of the present invention. The cells in the cell sample are not necessarily of the same type, although purification methods can be used to enrich for the type of cells that are preferably evaluated. Cells can be obtained, for example, by scraping of a tissue, processing of a tissue sample to release individual cells, or isolation from a bodily fluid.

A tissue sample, although similar to a sample of isolated cells, is defined herein as a section of an organ or tissue of the body which typically includes several cell types and/or cytoskeletal structure which holds the cells together. One of skill in the art will appreciate that the term "tissue sample" may be used, in some instances, interchangeably with a "cell sample", although it is preferably used to designate a more complex structure than a cell sample. A tissue sample can be obtained by a biopsy, for example, including by cutting, slicing, or a punch. A bodily fluid sample, like the tissue sample, contains the cells to be evaluated for marker expression or biological activity and/or may contain a soluble biomarker that is secreted by cells, and is a fluid obtained by any method suitable for the particular bodily fluid to be sampled. Bodily fluids suitable for sampling include, but are not limited to, blood, mucous, seminal fluid, saliva, breast milk, bile and urine.

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In general, the sample type (i.e., cell, tissue or bodily fluid) is selected based on the accessibility and structure of the organ or tissue to be evaluated for tumor cell growth and/or on what type of cancer is to be evaluated. For example, if the organ/tissue to be evaluated is the breast, the sample can be a sample of epithelial cells from a biopsy (i.e., a cell sample) or a breast tissue sample from a biopsy (a tissue sample). The sample that is most useful in the present invention will be cells, tissues or bodily fluids isolated from a patient by a biopsy or surgery or routine laboratory fluid collection.

Once a sample is obtained from the patient, the sample is evaluated for detection of the expression or biological activity of the biomarker of the present invention in the cells of the sample. Expression and biological activity of biomarkers of the invention and methods of detecting or measuring the same have been described in detail above with regard to the description of the use of the biomarkers as targets.

For example, the level of the marker can be determined by conventional methods such as expression assays to determine the level of expression of the gene, by biochemical assays to determine the level of the gene product, or by immunoassays. If appropriate, the marker can be identified as a cell surface molecule in tissue or in a bodily fluid, such as serum. For example, a patient sample, which can be immobilized, can be contacted with an antibody, or an antibody fragment, that selectively binds to the marker and determining whether the anti-marker antibody or fragment thereof has bound to the marker. As used herein, the term "selectively binds to" refers to the specific binding of one protein to another (e.g., an antibody, fragment thereof, or binding partner to an antigen), wherein the level of binding, as measured by any standard assay (e.g., an immunoassay), is statistically significantly higher than the background control for the assay. For example, when performing an immunoassay, controls typically include a reaction well/tube that contain antibody or antigen binding fragment alone (i.e., in the absence of antigen), wherein an amount of reactivity (e.g., non-specific binding to the well) by the antibody or antigen binding fragment thereof in the absence of the antigen is considered to be background. Binding can be measured using a variety of methods standard in the art, including, but not limited to: Western blot, immunoblot, enzymelinked immunosorbant assay (ELISA), radioimmunoassay (RIA), immunoprecipitation, polarization, fluorescent resonance. chemiluminescence, surface plasmon immunohistochemical analysis, matrix-assisted laser phosphorescence, desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry, microcytometry,

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microarray, microscopy, fluorescence activated cell sorting (FACS), and flow cytometry. In a particular immunoassay, the marker level is determined using a first monoclonal antibody that binds specifically to the marker and a second antibody that binds to the first antibody.

In one embodiment, the amino acid sequence of a biomarker or the nucleic acid sequence of the corresponding gene can be used as a basis for detection. For example, detection can refer to detection of gene expression by determining the concentration of messenger RNA using common methods such as northern blot analysis, gene chip array analysis, Taqman analysis or other DNA/RNA hybridization platforms. The over or under expression of a biomarker can be an indication of the presence of a tumor or the predisposition for such tumor. Expression can be compared in patient samples versus samples isolated from healthy individuals.

In one embodiment of the method of the present invention, the level of a biomarker of the present invention is determined by determining the protein level of that biomarker in tissue. Suitable tissue tissues include tumor tissue and cell material obtained by biopsy.

In another embodiment of the method of the present invention, the level of a biomarker of the present invention is determined by determining a soluble form of a biomarker in a bodily fluid. Suitable bodily fluids include serum, ascitic or pleural fluid, serum being preferred. Levels of biomarker can be determined using various methods known in the art, including antibody binding assays, mass spectrometry analysis, 2dimensional gel analysis and other methods used to quantify the presence of protein in solution. One preferred method of the present invention is to immobilize a biomarker to a solid substrate and then incubate the biomarker with a patient's serum. Bound antibodies to the biomarker are then detected by means of an enzyme-conjugated second antibody and a color reaction. Another preferred method is to immobilize an antibody that binds to a biomarker to a solid substrate and incubate the antibody with patient serum. Biomarker in the serum binds to the immobilized antibody and is detected using a second different antibody that binds to the biomarker and a color reaction. Another preferred method of the present invention is to contact an antibody that binds to a biomarker with a patient sample and then determining whether the antibody has been bound to the biomarker. Such method can be achieved using known methods including fluorescence cell sorter (FACS) analysis.

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Suitable detection methods of a biomarker, an antibody that binds to a biomarker, or suitable nucleic acid probes, are known to those of skill in the art. The detection of biomarkers using antibodies is preferred, the same antibody being useful for both the soluble form and the form on the cell surface. Suitable antibodies for the method of the present invention include monoclonal antibodies, polyclonal antibodies, and fragments thereof. The antibody fragment refers to all parts of the antibody that bind to the biomarker including Fab, Fy or single-chain Fy fragments. Methods to produce such fragments are known to those of skill in the art. Preferred antibodies include monoclonal antibodies. Such antibodies can be produced using standard methods in the art.

Another method of the present invention can include immobilizing patient tissue in, for example, paraffin. The immobilized tissue can be sectioned and then contacted with an antibody that binds to a biomarker.

In the diagnostic/prognostic methods of the invention, if the level of the marker is greater than a normal level, the level of the marker is considered to be indicative of the presence of tumor cells. A normal level can be determined in a variety of ways. For example, if a patient history is known, a baseline level of the marker can be determined and higher levels will be indicative of tumor cells. Alternatively, a normal level can be based on the level for a healthy (i.e., without tumor) individual in a given population. That is, a normal level can be based on a population having similar characteristics (e.g., age, sex, race, medical history) as the patient in question.

More specifically, according to the present invention, a "baseline level" is a control level, and in some embodiments (but not all embodiments, depending on the method), a normal level, of biomarker expression or activity against which a test level of biomarker expression or biological activity (*i.e.*, in the test sample) can be compared. Therefore, it can be determined, based on the control or baseline level of biomarker expression or biological activity, whether a sample to be evaluated for tumor cell growth has a measurable increase, decrease, or substantially no change in biomarker expression or biological activity, as compared to the baseline level. In one aspect, the baseline level can be indicative of the cell growth expected in a normal (*i.e.*, healthy, negative control, non-tumor) cell sample. Therefore, the term "negative control" used in reference to a baseline level of biomarker expression or biological activity typically refers to a baseline level established in a sample from the patient or from a population of individuals which is believed to be normal (*i.e.*, non-tumorous, not undergoing neoplastic transformation, not

exhibiting inappropriate cell growth). It is noted that the "negative control" most typically has a lower level of biomarker expression or activity than would be detected in an experimental cell having inappropriate, increased cell growth, because the expression/biological activity of the biomarkers described herein are correlated with cell growth in most tumor cell types. In another embodiment, a baseline can be indicative of a positive diagnosis of tumor cell growth. Such a baseline level, also referred to herein as a "positive control" baseline, refers to a level of biomarker expression or biological activity established in a cell sample from the patient, another patient, or a population of individuals, wherein the sample was believed, based on data for that cell sample, to be neoplastically transformed (i.e., tumorous, exhibiting inappropriate cell growth, cancerous). It is noted that this "positive control" will most typically have a higher level of biomarker expression or activity than in a normal cell, again due to the correlative relationship between the biomarkers of the present invention and cell growth in the majority of tumor cells. In yet another embodiment, the baseline level can be established from a previous sample from the patient being tested, so that the tumor growth of a patient can be monitored over time and/or so that the efficacy of a given therapeutic protocol can be evaluated over time. Methods for detecting biomarker expression or biological activity are described in detail above.

The method for establishing a baseline level of biomarker expression or activity is selected based on the sample type, the tissue or organ from which the sample is obtained, the status of the patient to be evaluated, and, as discussed above, the focus or goal of the assay (e.g., diagnosis, staging, monitoring). Preferably, the method is the same method that will be used to evaluate the sample in the patient. In a most preferred embodiment, the baseline level is established using the same cell type as the cell to be evaluated.

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In one embodiment, the baseline level of biomarker expression or biological activity is established in an autologous control sample obtained from the patient. The autologous control sample can be a sample of isolated cells, a tissue sample or a bodily fluid sample, and is preferably a cell sample or tissue sample. According to the present invention, and as used in the art, the term "autologous" means that the sample is obtained from the same patient from which the sample to be evaluated is obtained. The control sample should be of or from the same cell type and preferably, the control sample is obtained from the same organ, tissue or bodily fluid as the sample to be evaluated, such that the control sample serves as the best possible baseline for the sample to be evaluated.

In one embodiment, when the goal of the assay is diagnosis of abnormal cell growth, it is desirable to take the control sample from a population of cells, a tissue or a bodily fluid which is believed to represent a "normal" cell, tissue, or bodily fluid, or at a minimum, a cell or tissue which is least likely to be undergoing or potentially be predisposed to develop tumor cell growth. For example, if the sample to be evaluated is an area of apparently abnormal cell growth, such as a tumorous mass, the control sample is preferably obtained from a section of apparently normal tissue (*i.e.*, an area other than and preferably a reasonable distance from the tumorous mass) in the tissue or organ where the tumorous mass is growing. In one aspect, if a tumor to be evaluated is in the colon, the test sample would be obtained from the suspected tumor mass and the control sample would be obtained from a different section of the colon, which is separate from the area where the mass is located and which does not show signs of uncontrolled cellular proliferation.

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In another embodiment, when the goal is to monitor tumor cell growth in the patient, the autologous baseline sample is typically a previous sample from the patient which was taken from an apparent or confirmed tumorous mass, and/or from apparently normal (i.e., non-tumor) tissue in the patient (or a different type of baseline for normal can be used, as discussed below).

Therefore, a second method for establishing a baseline level of biomarker expression or biological activity is to establish a baseline level of biomarker expression or biological activity from at least one measurement of biomarker expression or biological activity in a previous sample from the same patient. Such a sample is also an autologous sample, but is taken from the patient at a different time point than the sample to be tested. Preferably, the previous sample(s) were of a same cell type, tissue type or bodily fluid type as the sample to be presently evaluated. In one embodiment, the previous sample resulted in a negative diagnosis (*i.e.*, no tumor cell growth, or potential therefore, was identified). In this embodiment, a new sample is evaluated periodically (*e.g.*, at annual physicals), and as long as the patient is determined to be negative for tumor development, an average or other suitable statistically appropriate baseline of the previous samples can be used as a "negative control" for subsequent evaluations. For the first evaluation, an alternate control can be used, as described below, or additional testing may be performed to confirm an initial negative diagnosis, if desired, and the value for biomarker expression or biological activity can be used thereafter. This type of baseline control is frequently

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used in other clinical diagnosis procedures where a "normal" level may differ from patient to patient and/or where obtaining an autologous control sample at the time of diagnosis is not possible, not practical or not beneficial. For example, for a patient who has periodic mammograms, the previous mammograms serve as baseline controls for the mammary tissue of the individual patient. Similarly, for a patient who is regularly screened for prostate cancer by evaluation of levels of prostate cancer antigen (PCA), previous PCA levels are frequently used as a baseline for evaluating whether the individual patient experiences a change.

In another embodiment, the previous sample from the patient resulted in a positive diagnosis (i.e., tumor growth was positively identified). In this embodiment, the baseline provided by the previous sample is effectively a positive control for tumor growth, and the subsequent samplings of the patient are compared to this baseline to monitor the progress of the tumor growth and/or to evaluate the efficacy of a treatment which is being prescribed for the cancer. In this embodiment, it may also be beneficial to have a negative baseline level of biomarker expression or biological activity (i.e., a normal cell baseline control), so that a baseline for remission or regression of the tumor can be set. Monitoring of a patient's tumor growth can be used by the clinician to modify cancer treatment for the patient based on whether an increase or decrease in cell growth is indicated.

It will be clear to those of skill in the art that some samples to be evaluated will not readily provide an obvious autologous control sample, or it may be determined that collection of autologous control samples is too invasive and/or causes undue discomfort to the patient. In these instances, an alternate method of establishing a baseline level of biomarker expression or biological activity can be used, examples of which are described below.

Another method for establishing a baseline level of biomarker expression or biological activity is to establish a baseline level of biomarker expression or biological activity from control samples, and preferably control samples that were obtained from a population of matched individuals. It is preferred that the control samples are of the same sample type as the sample type to be evaluated for biomarker expression or biological activity (e.g., the same cell type, and preferably from the same tissue or organ). According to the present invention, the phrase "matched individuals" refers to a matching of the control individuals on the basis of one or more characteristics which are suitable

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for the type of cell or tumor growth to be evaluated. For example, control individuals can be matched with the patient to be evaluated on the basis of gender, age, race, or any relevant biological or sociological factor that may affect the baseline of the control individuals and the patient (e.g., preexisting conditions, consumption of particular substances, levels of other biological or physiological factors). To establish a control or baseline level of biomarker expression or biological activity, samples from a number of matched individuals are obtained and evaluated for biomarker expression or biological activity. The sample type is preferably of the same sample type and obtained from the same organ, tissue or bodily fluid as the sample type to be evaluated in the test patient. The number of matched individuals from whom control samples must be obtained to establish a suitable control level (e.g., a population) can be determined by those of skill in the art, but should be statistically appropriate to establish a suitable baseline for comparison with the patient to be evaluated (i.e., the test patient). The values obtained from the control samples are statistically processed using any suitable method of statistical analysis to establish a suitable baseline level using methods standard in the art for establishing such values.

A baseline such as that described above can be a negative control baseline, such as a baseline established from a population of apparently normal control individuals. Alternatively, as discussed above, such a baseline can be established from a population of individuals that have been positively diagnosed as having cancer, and particularly, cancer of a specified stage, as set forth by the medical community, so that one or more baseline levels can be established for use in staging a cancer in the patient to be evaluated. Therefore, in one embodiment, the baseline level is one or more tumor control samples that are correlated with a particular stage of tumor development for that type of tumor. For example, tumor samples from an appropriate number of individuals that have been diagnosed as having a particular stage of a given cancer (e.g., Stage I colon cancer) are tested for biomarker expression or biological activity. The values obtained from these control samples are statistically processed to establish a suitable baseline level using methods standard in the art for establishing such values, and the baseline is noted as being indicative of that particular stage of cancer. Preferably, a similar value is determined for each of the established stages of the given cancer, so that a panel of baseline values, each representing a different stage of the cancer, is formed. The level of biomarker expression or biological activity in the patient sample is then compared to each of the baseline levels

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to determine to which baseline the biomarker level of the patient is statistically closest. It will be appreciated that a given patient sample may fall between baseline levels of two different stages such that the best diagnosis is that the patient tumor is at least at the lower stage, but is perhaps in the process of advancing to the higher stage. The data provided by this method can be used in conjunction with current cancer staging methods to assist the physician in the evaluation of the patient and in prescribing suitable treatment for the cancer.

It will be appreciated by those of skill in the art that a baseline need not be established for each assay as the assay is performed but rather, a baseline can be established by referring to a form of stored information regarding a previously determined baseline level of biomarker expression for a given control sample, such as a baseline level established by any of the above-described methods. Such a form of stored information can include, for example, but is not limited to, a reference chart, listing or electronic file of population or individual data regarding "normal" (negative control) or tumor positive (including staged tumors) biomarker expression; a medical chart for the patient recording data from previous evaluations; or any other source of data regarding baseline biomarker expression that is useful for the patient to be diagnosed.

After the level of biomarker expression or biological activity is detected in the sample to be evaluated for tumor cell growth, such level is compared to the established baseline level of biomarker expression or biological activity, determined as described above. Also, as mentioned above, preferably, the method of detecting used for the sample to be evaluated is the same or qualitatively and/or quantitatively equivalent to the method of detecting used to establish the baseline level, such that the levels of the test sample and the baseline can be directly compared. In comparing the test sample to the baseline control, it is determined whether the test sample has a measurable decrease or increase in biomarker expression or biological activity over the baseline level, or whether there is no statistically significant difference between the test and baseline levels. After comparing the levels of biomarker expression or biological activity in the samples, the final step of making a diagnosis, monitoring, or staging of the patient can be performed as discussed above.

According to the present invention, detection of an increased level of biomarker expression or biological activity in the sample to be evaluated (i.e., the test sample) as compared to the baseline level indicates that, as compared to the baseline sample,

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increased cell growth or tumorigenicity or a potential therefore is indicated in the cells corresponding to the test sample. This indication of increased tumorigenicity is evaluated based on what the baseline represents, and can mean: (1) a positive diagnosis of tumorigenicity (i.e., neoplastic transformation) or potential for tumor cell growth in the patient; (2) continued or increased tumorigenicity in a patient previously diagnosed with a cancer; and/or (3) a higher stage of tumorigenicity than that represented by the baseline. More specifically, if the baseline is a normal or negative control sample (i.e., autologous or otherwise established, such as from a population control), a detection of increased biomarker expression or biological activity in the test sample as compared to the control sample indicates that the cells in the test sample are undergoing (or are at risk of undergoing) increased, and likely inappropriate (i.e., tumorous, neoplastic) cell growth. If the baseline sample is a previous sample from the patient (or a population control) and is representative of a positive diagnosis of tumor cell growth in the patient (i.e., a positive control), a detection of increased biomarker expression or biological activity in the sample as compared to the baseline may indicate that the cells in the test sample are experiencing increased tumor growth or a potential therefore, which would suggest to a clinician that a treatment currently being prescribed, for example, is not controlling the tumor growth or that tumor growth in the patient has recurred. If the baseline sample is representative of a particular stage of tumor, a detection of increased biomarker expression or biological activity in the sample as compared to the baseline may indicate that the cells in the test sample are at a higher stage of tumor growth than the stage represented by the baseline sample.

Similarly, detection of a decreased level of biomarker expression or biological activity in the sample to be evaluated (i.e., the test sample) as compared to the baseline level indicates that, as compared to the baseline sample, decreased cell growth or tumorigenicity or a potential therefore is indicated in the test cells. This indication of decreased tumorigenicity is evaluated based on what the baseline represents, and can mean: (1) a negative diagnosis of tumorigenicity (neoplastic transformation) or potential for tumor cell growth in the patient; (2) reduced tumorigenicity in a patient previously diagnosed with a cancer; and/or (3) a lower stage of tumorigenicity than that represented by the baseline. More specifically, if the baseline is a normal or negative control (autologous or otherwise established, such as from a population control), a detection of decreased biomarker expression or biological activity in the test sample as compared to

the control sample indicates that the cells in the test sample are also normal and are not predicted to be at risk of undergoing inappropriate (*i.e.*, tumorous, neoplastic) cell growth. If the baseline sample is a previous sample from the patient (or from a population control) and is representative of a positive diagnosis of tumorigenicity in the patient (*i.e.*, a positive control), a detection of decreased biomarker expression or biological activity in the sample as compared to the baseline indicates that the cells in the test sample are experiencing decreased tumorigenicity or a potential therefore, which suggests to a clinician, for a patient that has cancer, that a treatment currently being prescribed, for example, is successfully controlling the tumor growth or that a tumor in the patient is in remission or eliminated. If the baseline sample is representative of a particular stage of tumor, a detection of decreased biomarker expression or biological activity in the sample as compared to the baseline indicates that the cells in the test sample are at a lower stage of tumor growth than the stage represented by the baseline sample.

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Finally, detection of biomarker expression that is not statistically significantly different than the biomarker expression or biological activity in the baseline sample indicates that, as compared to the baseline sample, no difference in tumorigenicity or a potential therefore is indicated in the test cells. This indication of effectively a "baseline level" of cell growth in the test cell is evaluated based on what the baseline represents, and can mean: (1) a negative or positive diagnosis of tumorigenicity (neoplastic transformation) or potential therefore in the patient; (2) unchanged tumorigenicity in a patient previously diagnosed with a cancer; and/or (3) a correlation with a stage of tumor growth that is represented by the baseline. More specifically, if the baseline is a normal or negative control (autologous or otherwise established, such as from a population control), detection of biomarker expression or biological activity in the test sample that is not statistically significantly different than the baseline sample indicates that the cells in the test sample are also normal and are not predicted to be at risk of undergoing inappropriate (i.e., tumorous, neoplastic) cell growth. If the baseline sample is a previous sample from the patient (or from a population control) and is representative of a positive diagnosis of tumor cell growth in the patient (i.e., a positive control), a detection of biomarker expression or biological activity in the sample that is not statistically significantly different than the baseline indicates that the cells in the test sample are experiencing tumor cell growth or a potential therefore, and the patient should be further evaluated for cancer. In a patient who has cancer and is being monitored for tumor progression, a detection of biomarker expression or biological activity in the test sample that is not statistically significantly different than the baseline sample indicates that the tumor is neither increasing (progressing) nor decreasing (regressing). Such a diagnosis might suggest to a clinician that a treatment currently being prescribed, for example, is ineffective in controlling the tumor growth or is preventing accelerated tumor growth, but is not causing tumor growth to regress. Finally, if the baseline sample is representative of a particular stage of tumor, a detection of biomarker expression or biological activity in the test sample that is not statistically significantly different than the baseline sample indicates that the cells in the test sample are at substantially the same stage of tumor growth as the stage represented by the baseline sample.

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As discussed above, a positive diagnosis indicates that increased cell growth, and possibly tumor cell growth (neoplastic transformation), has occurred, is occurring, or is statistically likely to occur in the cells or tissue from which the sample was obtained. In order to establish a positive diagnosis, the level of biomarker activity is increased over the established baseline by an amount that is statistically significant (i.e., with at least a 95% confidence level, or p<0.05). Preferably, detection of at least about a 10% change in biomarker expression or biological activity in the sample as compared to the baseline level results in a positive diagnosis of increased cell growth for said sample, as compared to the baseline. More preferably, detection of at least about a 30% change in biomarker expression or biological activity in the sample as compared to the baseline level results in a positive diagnosis of increased cell growth for said sample, as compared to the baseline. More preferably, detection of at least about a 50% change, and more preferably at least about a 70% change, and more preferably at least about a 90% change, or any percentage change between 5% and higher in 1% increments (i.e., 5%, 6%, 7%, 8%...) in biomarker expression or biological activity in the sample as compared to the baseline level results in a positive diagnosis of increased tumorigenicity for said sample. In one embodiment, a 1.5 fold change in biomarker expression or biological activity in the sample as compared to the baseline level results in a positive diagnosis of increased tumorigenicity for said sample. More preferably, detection of at least about a 3 fold change, and more preferably at least about a 6 fold change, and even more preferably, at least about a 12 fold change, and even more preferably, at least about a 24 fold change, or any fold change from 1.5 up in increments of 0.5 fold (i.e., 1.5, 2.0, 2.5, 3.0...) in biomarker expression or biological

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activity as compared to the baseline level, results in a positive diagnosis of increased tumorigenicity for said sample.

This method of diagnosis can be used specifically to determine the prognosis for cancer in the patient or to determine the susceptibility of the patient to a therapeutic treatment. In some embodiments, the method may be useful to monitor the progress of a patient undergoing therapeutic treatment for a tumor.

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The present invention also includes a kit that utilizes the diagnostic methods of the present invention. The kit preferably contains any means of detecting the expression or activity of a biomarker of the present invention in a test sample, and preferably includes a probe, PCR primers, or an antibody, antigen binding peptide, or fragment thereof, that binds to a biomarker. The kit can include any reagent needed to perform a diagnostic method envisioned herein. The antibody, or fragment thereof, can be conjugated to another unit, for example a marker or immobilized to a solid carrier (substrate). The kit can also contain a second antibody for the detection of biomarker:antibody complexes. In one embodiment, the kit can contain a means for detecting a control marker characteristic of a cell type in the test sample. The antibody, or fragment thereof, may be present in free form or immobilized to a substrate such as a plastic dish, a test tube, a test rod and so on. The kit can also include suitable reagents for the detection of and/or for the labeling of positive or negative controls, wash solutions, dilution buffers and the like.

More specifically, according to the present invention, a means for detecting biomarker expression or biological activity can be any suitable reagent that can be used in a method for detection of biomarker expression or biological activity as described previously herein. Such reagents include, but are not limited to: a probe that hybridizes under stringent hybridization conditions to a nucleic acid molecule encoding the biomarker or a fragment thereof (including to a biomarker-specific regulatory region in the biomarker-encoding gene); RT-PCR primers for amplification of mRNA encoding the biomarker or a fragment thereof; and/or an antibody, antigen-binding fragment thereof or other antigen-binding peptide that selectively binds to the biomarker.

According to the present invention, a probe is a nucleic acid molecule which typically ranges in size from about 8 nucleotides to several hundred nucleotides in length. Such a molecule is typically used to identify a target nucleic acid sequence in a sample by hybridizing to such target nucleic acid sequence under stringent hybridization conditions. Hybridization conditions have been described in detail above.

PCR primers are also nucleic acid sequences, although PCR primers are typically oligonucleotides of fairly short length which are used in polymerase chain reactions. PCR primers and hybridization probes can readily be developed and produced by those of skill in the art, using sequence information from the target sequence. (See, for example, Sambrook et al., *supra* or Glick et al., *supra*).

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Antibodies that selectively bind to a biomarker in the sample can be produced using information available in the art. Antibodies useful in the assay kit and methods of the present invention can include polyclonal and monoclonal antibodies, divalent and monovalent antibodies, bi- or multi-specific antibodies, serum containing such antibodies, antibodies that have been purified to varying degrees, and any functional equivalents of whole antibodies. Isolated antibodies of the present invention can include serum containing such antibodies, or antibodies that have been purified to varying degrees. Whole antibodies of the present invention can be polyclonal or monoclonal. Alternatively, functional equivalents of whole antibodies, such as antigen binding fragments in which one or more antibody domains are truncated or absent (e.g., Fv, Fab, Fab', or F(ab)<sub>2</sub> fragments), as well as genetically-engineered antibodies or antigen binding fragments thereof, including single chain antibodies or antibodies that can bind to more than one epitope (e.g., bi-specific antibodies), or antibodies that can bind to one or more different antigens (e.g., bi- or multi-specific antibodies), may also be employed in the invention.

Genetically engineered antibodies include those produced by standard recombinant DNA techniques involving the manipulation and re-expression of DNA encoding antibody variable and/or constant regions. Particular examples include, chimeric antibodies, where the V<sub>H</sub> and/or V<sub>L</sub> domains of the antibody come from a different source to the remainder of the antibody, and CDR grafted antibodies (and antigen binding fragments thereof), in which at least one CDR sequence and optionally at least one variable region framework amino acid is (are) derived from one source and the remaining portions of the variable and the constant regions (as appropriate) are derived from a different source. Construction of chimeric and CDR-grafted antibodies is described, for example, in European Patent Applications: EP-A 0194276, EP-A 0239400, EP-A 0451216 and EP-A 0460617.

Generally, in the production of an antibody, a suitable experimental animal, such as, for example, but not limited to, a rabbit, a sheep, a hamster, a guinea pig, a mouse, a

rat, or a chicken, is exposed to an antigen against which an antibody is desired. Typically, an animal is immunized with an effective amount of antigen that is injected into the animal. An effective amount of antigen refers to an amount needed to induce antibody production by the animal. The animal's immune system is then allowed to respond over a pre-determined period of time. The immunization process can be repeated until the immune system is found to be producing antibodies to the antigen. In order to obtain polyclonal antibodies specific for the antigen, serum is collected from the animal that contains the desired antibodies (or in the case of a chicken, antibody can be collected from the eggs). Such serum is useful as a reagent. Polyclonal antibodies can be further purified from the serum (or eggs) by, for example, treating the serum with ammonium sulfate.

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Monoclonal antibodies may be produced according to the methodology of Kohler and Milstein (*Nature* 256:495-497, 1975). For example, B lymphocytes are recovered from the spleen (or any suitable tissue) of an immunized animal and then fused with myeloma cells to obtain a population of hybridoma cells capable of continual growth in suitable culture medium. Hybridomas producing the desired antibody are selected by testing the ability of the antibody produced by the hybridoma to bind to the desired antigen.

The invention also extends to non-antibody polypeptides, sometimes referred to as antigen binding partners or antigen binding peptides, which have been designed to bind selectively to the protein of interest (a biomarker). Examples of the design of such polypeptides, which possess a prescribed ligand specificity, are given in Beste et al. (*Proc. Natl. Acad. Sci.* 96:1898-1903, 1999), incorporated herein by reference in its entirety.

In one embodiment, a means for detecting a control marker that is characteristic of the cell type being sampled can generally be any type of reagent that can be used in a method of detecting the presence of a known marker in a sample, such as by a method for detecting the presence of a biomarker described previously herein. Specifically, the means is characterized in that it identifies a specific marker of the cell type being analyzed that positively identifies the cell type. For example, in a breast tumor assay, it is desirable to screen breast epithelial cells for the level of the biomarker expression and/or biological activity. Therefore, the means for detecting a control marker identifies a marker that is characteristic of an epithelial cell and preferably, a breast epithelial cell, so

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that the cell is distinguished from other cell types, such as a fibroblast. Such a means increases the accuracy and specificity of the assay of the present invention. Such a means for detecting a control marker include, but are not limited to: a probe that hybridizes under stringent hybridization conditions to a nucleic acid molecule encoding a protein marker; PCR primers which amplify such a nucleic acid molecule; and/or an antibody, antigen binding fragment thereof, or antigen binding peptide that selectively binds to the control marker in the sample. Nucleic acid and amino acid sequences for many cell markers are known in the art and can be used to produce such reagents for detection.

The means for detecting a biomarker and/or a control marker of the assay kit of the present invention can be conjugated to a detectable tag or detectable label. Such a tag can be any suitable tag which allows for detection of the reagents used to detect the biomarker or control marker and includes, but is not limited to, any composition or label detectable by spectroscopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Useful labels in the present invention include biotin for staining with labeled streptavidin conjugate, magnetic beads (e.g., Dynabeads<sup>TM</sup>), fluorescent dyes (e.g., fluorescein, texas red, rhodamine, green fluorescent protein, and the like), radiolabels (e.g., <sup>3</sup>H, <sup>125</sup>I, <sup>35</sup>S, <sup>14</sup>C, or <sup>32</sup>P), enzymes (e.g., horse radish peroxidase, alkaline phosphatase and others commonly used in an ELISA), and colorimetric labels such as colloidal gold or colored glass or plastic (e.g., polystyrene, polypropylene, latex, etc.) beads.

In addition, the means for detecting of the assay kit of the present invention can be immobilized on a substrate. Such a substrate can include any suitable substrate for immobilization of a detection reagent such as would be used in any of the previously described methods of detection. Briefly, a substrate suitable for immobilization of a means for detecting includes any solid support, such as any solid organic, biopolymer or inorganic support that can form a bond with the means for detecting without significantly effecting the activity and/or ability of the detection means to detect the desired target molecule. Exemplary organic solid supports include polymers such as polystyrene, nylon, phenol-formaldehyde resins, acrylic copolymers (e.g., polyacrylamide), stabilized intact whole cells, and stabilized crude whole cell/membrane homogenates. Exemplary biopolymer supports include cellulose, polydextrans (e.g., Sephadex®), agarose, collagen and chitin. Exemplary inorganic supports include glass beads (porous and nonporous),

stainless steel, metal oxides (e.g., porous ceramics such as ZrO<sub>2</sub>, TiO<sub>2</sub>, Al<sub>2</sub>O<sub>3</sub>, and NiO) and sand.

According to the present invention, the method and assay for assessing the tumorigenicity of cells in a patient, as well as other methods disclosed herein, are suitable for use in a patient or cells from a patient or host that is a member of the Kingdom, Animalia, and particularly of the Vertebrate class, Mammalia, including, without limitation, primates, livestock and domestic pets (e.g., a companion animal). Most typically, a patient will be a human patient or host cells will be derived from human patients, although the use of the methods of the invention in any suitable non-human animal model or host cell is also encompassed.

All publications cited herein are incorporated by reference in their entirety.

The Examples, which follow, are illustrative of specific embodiments of the invention, and various uses thereof. They are set forth for explanatory purposes only, and are not to be taken as limiting the invention.

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### **EXAMPLES**

### Example 1

The purpose of this experiment was to perform a nearly saturated genome wide GSE screen in a tumor cell line model for GSEs that protect cells against apoptosis.

## 1. V93 Vector Design and Construction

Vector V98 was created through modification of p610SL, a derivative of pLNCO<sub>3</sub> (B-D Chang and I.B. Roninson, Gene 183 (1996) 137-142.) A schematic of V98 is shown in Figure 1. The region flanking the multiple cloning site (MCS) downstream of the inducible CMV promoter was re-engineered (1) to introduce restriction endonuclease sites for enzymes expected to occur with low frequency in the human genome [e.g., Fse I (1 per 170 kBp), Mlu I (1 per 300 kBp), and Rsr II (1 per 260 kBp)], (2) to introduce a short sequence of nucleic acid containing stop codons in all three DNA reading frames downstream of the MCS, (3) to introduce between the Fse I and Mlu I sites on the reengineered vector backbone a Kozak sequence for efficient translation initiation of peptides encoded by random fragments cloned into the MCS (4) to introduce sequences complementary to well established DNA primers used for DNA sequencing (e.g., M13F-20 and M13R), to permit rapid and efficient sequencing of inserts cloned into the MCS, (5) to introduce sequences flanking the MCS, derived from the genome of Zea mays, and

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(6) to introduce into the MCS a "stuffer" fragment of about 2.2 kBp, which permits easy assessment of the completeness of vector digestion and selection of the appropriate backbone fragment during vector preparation.

A cDNA encoding the open reading frame of the murine Lyt-2-alpha' gene was recovered from Marathon ready mouse spleen cDNA (Clontech) using PCR with the following conditions: 5 µL of marathon ready cDNA, 5 µL 10X cDNA PCR buffer, 1 µL 10 mM dNTP mix, 1 μL Advantage 2 polymerase mix (Clontech, #8430-1), 1 μL of 10 uM upstream primer 5'- ACC ATG GCC TCA CCG TTG ACC CGC TTT -3' (SEQ ID NO:81), 1 μL or 10 μM downstream primer 5'- CTA GCG GCT GTG GTA GCA GAT GAG A -3' (SEQ ID NO:82), and 36 µL of water. Cycling parameters were: 94°C for 3 min.: followed by 30 cycles of 94°C for 30 sec., 55°C for 30 sec., and 72°C for 2 minutes; followed by 72°C for 10 minutes; followed by a 4°C soak. The resultant PCR product of 669 nucleotides was subcloned into a pCRII TOPO vector (InVitrogen). independent clones were sequenced to confirm no mutations were introduced in the Lyt-2-alpha' ORF by the PCR. One pCRII-TOPO-Lyt-2-alpha' clone was shown to be free of mutations, clone #2. DNA from clone #2 was subjected to a second round of PCR (Vt = 50 μL) using the following conditions: 1 ng plasmid DNA, 5 μL 10X cDNA PCR buffer, 0.8 μL of 10 mM dNTP mix, 1 μL of Advantage 2 polymerase mix (Clontech, #8430-1), 2.5 µL of 10 µM upstream primer 5'- CTA CGG ATC CAC CAT GGC CTC ACC GTT GA -3' (SEQ ID NO:83) and 2.5 µL of 10 µM downstream primer 5'- GTA CAT CGA TCT AGC GGC TGT GGT AGC AGA TGA GA -3' (SEQ ID NO:84). These primers permitted recovery the ORF of the Lyt-2-alpha' gene flanked by BamH I (upstream) and Cla I restriction endonuclease sites. Cycling parameters were: 94°C for 3 min.; followed by 30 cycles of 94°C for 30 sec., 55°C for 30 sec., and 72°C for 2 minutes; followed by 72°C for 10 minutes; followed by a 4°C soak. The resulting 689-bp PCR product was purified from surrounding proteins and salts using a Qiagen PCR clean up kit following The purified Clone #2 DNA digested with Bam HI manufacturer's instructions. restriction endonuclease (NEB, #R0136S). The digested product was purified using a Qiagen PCR clean up kit and the buffer was changed. The digested DNA was then further digested with Cla I restriction endonuclease (NEB, #R0197S). The doubly restricted Clone #2 DNA was then subcloned into the backbone fragment of the 610SL retroviral vector produced by double digestion of 610SL with Bcl I (NEB, #R0160S) and

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Sfu I (Roche, #1243497) restriction endonucleases. Sequencing of DNA harvested from several independent bacterial colonies that were produced from this subcloning step yielded a clone that showed no mutations in the Lyt-2-alpha' ORF. This clone was named V97.

The modifications to the MCS regions of vector 610SL were created by sequential cloning of various double stranded oligonucleotides containing the desired sequences into several precursor plasmids. Sequences designed to be located 5' to the Fse I GSE cloning site in V98, e.g., M13F-20 primer site, primer site for P1X, were created by subcloning annealed oligonucleotides 5'- AGC TGT AAA ACG ACG GCC AGT GAG CGT TTA AAC GAA TTC CAG ACT AGT GGC CGG CCG TGC A -3' (SEQ ID NO:85) and 5'- CGG CCG GCC ACT AGT CTG GAA TTC GTT TAA ACG CTC ACT GGC CGT CGT TTT AC -3' (SEQ ID NO:86) into the vector pEFGP-1 (ClonTech) between the HinD III and Pst I sites, to create pEGFP5'. The duplex produced by annealing primers 5'- AAT TCT GCA GCC CAG GTA AAA TTC GCT AGC CT -3' (SEQ ID NO:87) and 5'- CTA GAG GCT AGC GAA TTT TAC CTG GGC TGC AG -3' (SEQ ID NO:88), which contains the priming site for P1X sequence, was subcloned between the Eco RI and Spe I sites of pEGFP5' to yield pEGFP54. The modified 5' region of the MCS was recovered from plasmid pEFGP54 as a Bgl II - Not I flanked fragment, and subcloned between the Bgl II and Not I sites of p610SL, to yield p610-E54P1. Sequences designed to be located 3' to the Rsr II GSE cloning site in V98, e.g., 3 frame stop cassette, primer P2X, M13R sequencing primer, were created by subcloning of annealed oligonucleotides 5'- CGG TCC GTG AGT GAG TGA GGC GCG CC G GAT CCT AAC CTA GGT AAT CAT GGT CAT AGC TGT TTC CTG CAG GGC -3' (SEQ ID NO:89) and 5'- GGC CGC CCT GCA GGA AAC AGC TAT GAC CAT GAT TAC CTA GGT TAG GAT CCG GCG CGC CTC ACT CAC TCA CGG ACC GTG CA -3' (SEQ ID NO:90) into the vector pBlueScript II (Stratagene) between the Pst I and Not I sites, to create plasmid pBS3.3'. The duplex produced by annealing primers 5'- GAT CCC GGG TCG TGT ATT CAG CTT TCC TTG TTC CT -3' (SEQ ID NO:91) and 5'- CTA GAG GAA CAA GGA AAG CTG AAT ACA CGA CCC GG-3' (SEO ID NO:92), which contains the priming site for P2X sequence, was subcloned between the BamH I and Avr II sites of pBS3.3' to yield pBS3.3'P12.

The stuffer fragment for V98 was designed to contain a luciferase ORF joined to a prokaryotic blasticidin S transferase (bsd) expression cassette, in order to yield a 2.2 kBp

DNA fragment. The luciferase ORF and was created by PCR using the following primers 5'- CAT CAA GCT TGG CCG GCC ACC ATG GAC GCG TCC GAA GAC GCC AAA AAC ATA AAG -3' (SEQ ID NO:93) and 5'- CAC GTG GAT ATC TTA CAA TTT GGA CTT TCC GCC CT -3' (SEQ ID NO:94) to amplify the luciferase ORF from the 5 plasmid pNFkB-luc (Strategene, #219078), while the bsd expression cassette was created by PCR using the primers 5'-TTG TAA GAT ATC CAC GTG TTG ACA ATT AAT C -3' (SEQ ID NO:95) and 5'- CAT CAG ATC TGT CGA CCG GAC CGA CGC GTC CAC GAA GTG CTT AGC -3' (SEQ ID NO:96) to amplify the E7-blasticindin S transferase open reading frame cassette from plasmid EM7-bsd. (InVitrogen, #V511-20). Both reactions were performed using the following cycling parameters: 95°C for 3 min; 10 followed by 30 cycles of 94°C for 30 sec., 60°C for 30 sec., 72°C for 2 min.; followed by 72°C for 10 min.; followed by a soak at 4°C. PCR products of the desired size were purified by agarose gel electrophoresis followed by recovery of the DNA from the gel using the Qiagen Gel Extraction kit according to manufacturer's instructions. The 15 luciferase ORF and the bsd expression cassette were spliced together to generate a 2.2 kBp stuffer fragment using splice overlap extension PCR (Horton, R.M., Hunt, H.D., Ho, S.N., Pullen, J.K. and Pease, L.R. (1989)). Engineering hybrid genes without the use of restriction enzymes: gene splicing by overlap extension. Gene 77, 61-68) and the primers 5'- CAT CAA GCT TGG CCG GCC ACC ATG GAC GCG TCC GAA GAC GCC AAA AAC ATA AAG -3' (SEO ID NO:97) and 5'- CAT CAG ATC TGT CGA CCG GAC 20 CGA CGC GTC CAC GAA GTG CTT AGC -3' (SEQ ID NO:98). The resultant SOE PCR product was purified away from the proteins, primers, and salts using a Oiagen PCR clean up kit and following manufacturer's instructions. The product was digested with HinD III and Sal I restriction endonucleases, the restricted product was purified by agarose gel electrophoresis followed by recovery of the DNA from the gel using the 25 Oiagen Gel Extraction kit according to manufacturer's instructions, and the purified DNA was subcloned between the HinD III (NEB, #R0104S) and Xho I (NEB, #R0146S) sites of plasmid pBluescript to yield pBSlucSOEK. Plasmid pBSlucSOEK was sequenced to confirm it was free of unwanted mutations, and the stuffer fragment recovered from the 30 pBSlucSOEK as a HinD III and Rsr II fragment, which was purified by agarose gel electrophoresis, followed by recovery of the DNA from the gel using the Qiagen Gel Extraction kit according to manufacturer's instructions and subcloning of the fragment into the HinD III and Rsr II sites of plasmid pBS3.3'P12 to yield plasmid

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pBS33P2lucSOEK. Plasmid V87 was then constructed by recovering from plasmid pBS33P2lucSOEK the luciferase-E7-bsd stuffer fragment along with the 3' flanking sequences as an Fse I – Not I flanked 2.2 kBp DNA product, which was purified by agarose gel electrophoresis followed by recovery of the DNA from the gel using the Qiagen Gel Extraction kit according to manufacturer's instructions. This 2.2 kBp DNA was subcloned between the Fse I and Not I site of plasmid p610-E54P1, to yield vector V87. A schematic drawing of the construction of V87 is shown in Figure 2.

The downstream Mlu I site was removed from V87 by PCR amplification of the stuffer fragment of V90,a derivative of V87 containing the same stuffer as V87, using 1 ng of V90 template DNA and 2.5 µL of primer 5'- CAT CAA GCT TGG CCG GCC ACG CGT GTT GGT AAA ATG GAA GAC G-3' (SEQ ID NO:99) and 2.5 μL of primer 5'- CAT GAG ATC TGT CGA CCG GAC CGC CAC GAA GTG CTT AAG C -3' (SEQ ID NO:100) in a standard 50 µL PCR reaction using Taq DNA polymerase (Roche, 1146165). Cycling parameters were: 94°C for 3 min.; followed by 30 cycles of 94°C for 20 sec., 55°C for 20 sec., and 72°C for 3 minutes; followed by 72°C for 10 minutes; followed by a 4°C soak. The resultant 2.2 kBp PCR product was purified from the proteins and salt using a Qiagen PCR clean up kit following manufacturer's instructions. The PCR product was digested with Fse I and Rsr II endonucleases, purified by agarose gel electrophoresis, and recovered from the gel using a Qiagen Gel Extraction kit according to manufacturer's instructions. The restricted and purified fragment was then subcloned into the Fse I and Rsr II sites of V86, a vector related to V87 but containing a stuffer fragment containing the luciferase ORF but not the bsd ORF, to yield V94. The MCS GSE cassette was recovered from V94 as a 2.2 kBp DNA fragment by digestion of V94 with Bgl II (NEB, #R0144S) and Not I (NEB, #R0189S) restriction endonucleases. Vector V98 was created by subcloning this 2.2 kBp DNA fragment from V94 into the Bgl II and Not I sites on the V97 backbone. A schematic of the construction of V98 is shown in Figure 3.

### 2. Random Fragment Library Construction

For construction of the starting AOLC1U library, V98 vector described above was restricted at 37°C for 3 hours, using Mlu I (NEB, #R0198S) and Rsr II restriction endonucleases. For construction of all other selected libraries, e.g., AOLC1A, AOLC1B, AOLC1C, V98 vector DNA was restricted at 37°C for 3 hours, using Fse I and Rsr II restriction endonucleases. The vector DNA was purified from the digest using a Qiagen

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PCR clean up spin column according to manufacturer's instructions, and the vector backbone DNA was purified by subjecting the eluate from the column to agarose gel electrophoresis to resolve the various DNA digestion products according to mass. A gel slice containing the 7.7 kBp backbone fragment was excised, and the DNA recovered from the agarose slice using the Qiagen Gel Extraction kit according to manufacturer's The concentration of DNA present in the vector preparations was instructions. determined by ethidium bromide staining in an 0.8% agarose gel following electrophoresis, by comparison to a DNA sample composed of various bands of known size and mass (High DNA Mass Ladder, Life Technologies, 10406-016). Vector preparations were quality controlled in series of test ligations as follows: vector alone control reaction, composed of x µL vector DNA (30 fmol), z µL water, 4 µL 5X ligase buffer, 1  $\mu$ L T4 DNA ligase (BRL, 5 U/ $\mu$ L, #15224-041), where x + z = 15  $\mu$ L; and a vector + insert reaction, composed of x μL vector DNA (30 fmol), y μL insert DNA (90 fmol), z  $\mu$ L water, 4  $\mu$ L 5X ligase buffer, 1  $\mu$ L T4 DNA ligase, where x +y + z = 15  $\mu$ L. Ligation reactions were incubated at 16°C for at least 16 hours. At the end of the incubation period, ligation products were precipitated under ethanol, the ethanol decanted and the precipitate washed three times with 70% EtOH, and the pellet dried and resuspended in 20 µL of water. One microliter of resuspended DNA solution was electrotransformed into DH10B electrocompetent cells (Life Technologies, 18290-015) according to manufacturers instructions. Following transformation, bacteria was recovered in 960 µL of room temperature SOC media, and recovery mixtures incubated at 37°C in a rotary shaker, 250-300 rpm, for at least 40 minutes. After the recovery period, 4 ten-fold serial dilutions of each transformation culture were created, i.e., 1:10, 1:100, 1:1000, and 1:10000, and 50 µL of each bacterial dilution mixture was plated on LB-agar plates containing carbenicillin. Plates were incubated at 37°C overnight, and scored the following morning. Stock solutions of the double-restricted vector were aliquoted and stored frozen at -20°C, preferably in 30 fmol / tube amounts.

# 3. Preparation of Randomly fragmented cDNAs from Cell Line mRNA

Total RNA was harvested from five colon cancer cell lines: HCT15, HT29, HCT116, SW480 and SW620, using a Qiagen RNeasy kit, according to manufacturer's instructions. Poly A+ mRNA was purified from the total RNA using an Oligotex kit (Qiagen) following manufacturer's instructions. The purified mRNA pools were fragmented by boiling the sample at 100° C for 8 minutes, a time empirically determined

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to give a good distribution of cDNA fragments as demonstrated using a published fragmentation protocol (Gudkov and Roninson, "Isolation of Genetic Suppressor Elements (GSEs) from Random Fragment cDNA libraries in Retroviral Vectors." Chapter 18, in Methods in Molecular Biology, Vol. 69: cDNA Library Protocols, p. 228, I.G. Cowell and C. A. Austin, eds. Humana Press Inc., Totowa NJ, 1997). Two parallel first strand cDNA synthesis reactions were performed using the fragmented mRNAs as template, with either an Asc I-No random primer 5'- GTA ATA CGA CTC ACT ATA GGC GCG CCN<sub>9</sub>-3' (SEQ ID NO:101) or an Rsr II-N9 random primer 5'- GTA ATA CGA CTC ACT ATA GGC GGA CCG No -3' (SEQ ID NO:102) and the SuperScript Choice Systems for cDNA synthesis (Gibco BRL) following manufacturer's instructions. Second strand synthesis was performed using the method of Gubler and Hoffman Gene 25:263-9, 1989) again using the SuperScript kit. The resultant double strand cDNAs were blunted using T4 DNA polymerase (NEB, #M0203S), then ligated to double stranded adapters, produced by annealing the oligonucleotides 5'- ATG ATT ACG CCA CGG ACC GTC -3' (SEQ ID NO:103) and 5'- GAC GGT CCG TGG CGT AAT CAT GGT CAT AGC -3' (SEQ ID NO:104) to yield adapters containing an Rsr II restriction site, or the oligonucleotides 5'- ATG ATT ACG CCA GGC GCG CCA C-3' (SEQ ID NO:105) and 5'- GTG GCG CGC CTG GCG TAA TCA TGG TCA TAG C -3' (SEQ ID NO:106) to yield adapters containing an Asc I restriction site. cDNA samples prepared using the Asc-N<sub>9</sub> primer were ligated to the adapters containing the Rsr II restriction site, while cDNA samples prepared using the Rsr II-No primer were ligated to adapters containing the Asc I restriction site. After ligation of the adapters to the cDNA fragments, excess adapters were removed by spun column chromatography.

## 4. Preparation of Normalized Inserts for Starting AOLC1U Library

Eluted cDNAs ligated to appropriate adapters were subjected to 22 cycles of PCR to amplify the inserts and to generate large quantities of insert for self-normalization: those inserts ligated to Rsr II adapters were amplified using the primers 5'- GCT ATG ACC ATG ATT ACG CCA CGG ACC GTC -3' (SEQ ID NO:107) and 5'- GTA ATA CGA CTC ACT ATA GGC -3' (SEQ ID NO:108), while inserts ligated to the Asc I adapters were amplified using the primers 5'- GCT ATG ACC ATG ATT ACG CCA GGC GCG CCA C -3' (SEQ ID NO:109) and 5'- GTA ATA CGA CTC ACT ATA GGC GGA C -3' (SEQ ID NO:110). PCR products were pooled, purified using a Qiagen PCR

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kit following manufacturer's instructions, evaporated to dryness using a rotary evaporator, and then resuspended in 25 μL of 10 mM Tris-HCl (pH 8.5). The cDNA fragments were normalized by self-hybridization and batch binding hydroxyapatite (HAP) chromatography, essentially as described by Gudkov and Roninson (*op.cit*)., except that samples were collected at 24, 48, 72, 96 hours. The extent of normalization was evaluated using real-time PCR at five loci: ACTB, TP53, CASP3, 18S and a mitochondrial locus.

Purified, normalized ssDNA fractions from the HAP columns were reconverted to dsDNA and amplified using PCR: again, those inserts ligated to Rsr II adapters were amplified using the primers 5'- GCT ATG ACC ATG ATT ACG CCA CGG ACC GTC -3' (SEO ID NO:111) and 5'- GTA ATA CGA CTC ACT ATA GGC -3' (SEQ ID NO:112), while inserts ligated to the Asc I adapters were amplified using the primers 5'- GCT ATG ACC ATG ATT ACG CCA GGC GCG CCA C -3' (SEQ ID NO:113) and 5'- GTA ATA CGA CTC ACT ATA GGC GGA C -3' (SEQ ID NO:114). PCR products were purified using Qiagen PCR clean up columns following manufacturer's instructions, and the PCR products from the two types of inserts (e.g, those with Rsr II adapters and those with Asc adapters) were mixed one-to-one molar ratio. Approximately 100 ng of mixed PCR product was digested with Asc I (NEB, #R0558S) and Rsr II restriction endonucleases for 2 hours at 37°C in multiple parallel reactions. DNA was recovered from the pooled digestions using a Qiagen PCR clean up kit following manufacturer's instructions. The concentration of restricted PCR products in the eluate was determined by resolving the DNA present in an aliquot of the eluate by 2% agarose gel electrophoresis. The fluorescent intensity of the PCR product band was compared to the intensity of bands in a DNA sample composed of a mixture of DNA fragments of known size and mass (Low DNA Mass Ladder, Life Technologies, 10068-013).

### 5. Isolation of GSEs for AOLC1A, AOLC1B, or AOLC1C Libraries.

Colon adenocarcinoma SW480 cells were engineered to stably express ecotropic retroviral receptor (EcoR), and the resulting cell line was termed SW480 E. Phoenix Eco retrovirus packaging cells were transfected with library plasmid DNA and SW480 E cells were transduced with viral supernatant harvested from the packaging cells. Floating SW480 E cells were collected at times 24, 48, 72 and 96 hours post-transduction and fixed with 100% methanol. Apoptotic cells were collected from all time points by

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staining the fixed cells with a monoclonal antibody against caspase-cleaved cytokeratin 18 (M30 CytoDeath Antibody, Roche Diagnostics), and selecting stained cells by fluorescence activated cell sorting (FACS). Genomic DNA was isolated from the collected cells (typically between 1 x 10<sup>5</sup> and 2 x 10<sup>6</sup> cells, depending upon the selection round) using the Qiagen DNeasy kit (Qiagen). Recovered genomic DNA was quantitated using the PicoGreen DNA quantitation kit (Molecular Probes) in a fluorometric assay performed according to manufacturer's instructions.

GSEs were recovered from the integrated proviruses contained in the harvested genomic DNA using PCR and the following reaction recipe: 10 μL genomic DNA solution, about 1 μg DNA, 5 μL of 3.3 μM p5x primer 5'- TCT GCA GCC CAG GTA AAA TTC GCT AGC CTC TAG T -3' (SEQ ID NO:115), 5 μL of 3.3 μM p6x primer 5'- GAG GAA CAA GGA AAG CTG AAT ACA CGA CCC GTG AT -3' (SEQ ID NO:116), 2 μL of 10 mM dNTP mix, 17 μL of H<sub>2</sub>O, 10 μL 5X PCR buffer, 1 μL of Thermozyme (InVitrogen E120-01). Cycling conditions for the PCR were: 95°C for 3 min.; followed by 30 cycles of 95°C for 30 sec., 68°C for 30 sec., 72°C for 1 min.; followed by 72°C for 10 min., followed by a soak at 4°C. At least 10, and typically 96 reactions were performed in parallel.

Two hundred µL of pooled PCR product from the genomic PCR samples was purified from proteins and salts using a Qiagen PCR clean up kit following manufacturer's instructions. The concentration of PCR product in the eluate was determined by resolving the DNA present in an aliquot of the eluate by 2% agarose gel electrophoresis and then comparing the fluorescent intensity of the PCR product band to the intensity of bands in a DNA sample composed of a mixture of DNA fragments of known size and mass (Low DNA Mass Ladder, Life Technologies, 10068-013). Multiple parallel restriction digests were then set up using samples of the purified PCR product present in the eluate using the following recipe: 10 µL 10X NEB buffer #4 (final 1X concentration: 20 mM Tris-acetate, 10 mM magnesium acetate, 50 mM potassium acetate, 1 mM dithiothreitol), 1 µL 100X BSA (NEB), 7 µL Fse I restriction endonuclase (2 U/μL, NEB #R0588S), 3 μL Rsr II restriction endonuclease (4 U/μL, NEB #R0501S), X μL aliquot of PCR product, about 100 ng, 79 μL water, to bring total digestion volume up to 100 µL. Restriction digests were incubated at 37°C for 3 hours. DNA products from the digest were separated from proteins and salts and concentrated using a Zymo DNA Clean & Concentrator-5 concentrator kit (#D4004), following the manufacturer's

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instructions with the following modifications: after addition of DNA binding buffer to each of the digestion reactions, all of the reactions were spun through the same column to concentrate 600 to 800 ng of digested insert onto a single column. Columns were washed according to the manufacturer's protocols, and the DNA eluted from the column by two sequential additions of 8 µL of 50 mM Tris-HCl, pH 8.5. DNA of desired sizes (100-500 bp) was recovered from the concentrated eluate by purification using gel electrophoresis on 1% low melting point agarose (NuSieve GTG agarose, FMC bioproducts) gels. DNA bands in the gel were visualized following ethidium bromide staining of the gel, using a hand-held shortwave ultraviolet light source. Gel slices containing the desired DNA were excised using clean razor blades, and DNA extracted from the gel slice using the Qiagen gel purification kit, following manufacturer's instructions. The concentration of restricted and purified PCR product was determined by ethidium bromide staining of an agarose gel containing an aliquot of the purified PCR product, and a DNA sample of known composition and mass, as described above.

### 5. cDNA Library Preparation

Ligation reactions for each batch of insert prepared were set up as follows: (reaction 1) Vector control reaction: x μL vector DNA (150 ng), z μL water, 4 μL 5X ligase buffer, 1  $\mu$ L T4 DNA ligase (BRL, 5 U/ $\mu$ L, #15224-041), where x + z = 15  $\mu$ L; (reaction 2) Vector + insert: x µL vector DNA (150 ng), y µL insert DNA (12 ng); z µL water; 4  $\mu$ L 5X ligase buffer; 1  $\mu$ L T4 DNA ligase, where x + y + z = 15  $\mu$ L. Ligation mixtures were incubated at 4°C for at least 16 hours. At the end of the ligation period, ligation products were precipitated under ethanol, the ethanol decanted and the precipitate washed three times with 70% EtOH, and the pellet dried and resuspended in 20 µL of water. One µL of resuspended ligation product was used to electrotransform DH10B electrocompetent cells (Life Technologies, 18290-015) according to manufacturers instructions; the balance of the ligation mixture was stored at -20°C. Following transformation, bacteria was recovered in 960 µL of room temperature SOC media, and recovery mixtures incubated at 37°C in a rotary shaker, 250-300 rpm, for at least 40 minutes. After the recovery period, 4 ten-fold serially diluted samples (i.e., 1:10, 1:100, 1:1000, and 1:10000) of each transformation culture were set up, and 50 µL from each dilution was plated on LB-agar plates containing carbenicillin. Plates were incubated at 37°C overnight, and colony counts for each plate scored the following morning.

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Insert sizes in a subset of clones were determined by performing PCR directly on bacterial colonies as follows. A disposable pipette tip was used to harvest a single bacterial colony from the LB plate of interest. The colony was transferred into 25 µL of water, carefully swishing the tip to dislodge the bacterial colony. Five microliters of bacterial solution was spotted to an LB plate and allowed to incubate overnight. PCR was performed on the bacterial solution using the following recipe: 2 µL 10 mM primer M13F(17) 5'- GTA AAA CGA CGG CCA GT-3' (SEQ ID NO:117), 2 μL 10 mM primer p6X 5'- TCT GCA GCC CAG GTA AAA TTC GCT AGC CTC TAG T -3' (SEQ ID NO:118), 4 µL 10X PCR buffer, 1 µL 25 mM dNTP mix, 0.5 µL Taq DNA polymerase (Roche, 1146165), 10.5 μL PCR grade water, 20 μL of bacterial solution. The cycling parameters were 95°C for 3 min., then 25 cycles of 95°C for 30 sec., 60°C for 30 sec., 72°C for 1 min., followed by 72°C for 5 min., and a 4°C soak. At the completion of the PCR, 10 µL of each PCR product was resolved on a 2% agarose gel containing ethidium bromide. DNA mobility for each of the samples was evaluated. The balance of the PCR product was submitted for DNA sequencing to determine the sequence content of the inserts for these clones.

Ligations described above were used for further electro-transformations. The calculated cfu / µg for each of the QC controlled ligations was used to compute the total number of electrotransformations required to achieve the required complexity for the library being constructed. Multiple electrotransformations were performed in parallel, using 1 µL of ligation mix per transformation as described above. At the end of the 40minute recovery period following the electrotransformation, up to 10 independent transformations were pooled, and 50 µL from these pooled samples used to establish 4 ten-fold serially diluted samples (i.e., 1:10, 1:100, 1:1000, and 1:10000). Fifty μL of each serial dilution (i.e., 1:10, 1:100, 1:1000, and 1:10000) was plated on LB-agar plates containing carbenicillin. The remaining volumes of undiluted and diluted transformation solutions were used to seed a bacterial culture flask containing 0.5L of LB broth, after which the seeded flask was incubated at 30°C overnight, about 14 - 16 hours, in a rotary shaker at 300 rpm. Plates from the serially diluted samples were incubated at 37°C overnight, and colony counts for each plate scored the following morning to determine the total number of colonies seeded into the 0.5L culture. Library plasmid DNA was recovered from the 0.5L cultures using a Qiagen Maxiprep plasmid kit, according to manufacturer's instructions.

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The AOLC1U library was constructed using the normalized inserts prepared as described in section 4 above; the library was composed of greater than 80 million transformants. The AOLC1A library was constructed from GSEs recovered from apoptotic HCT116 cells collected 24, 48, 72 and 96 hours after transduction. The AOLC1B library was constructed from GSEs recovered from apoptotic HCT116 cells collected 24, 48, and 72 hours after transduction. The AOLC1C library was constructed from GSEs recovered from apoptotic HCT116 cells collected 48 hours after transduction. It was found that the AOLC1C library was highly enriched for RPX and E. coli sequences; with the RPL5, RPL36, RPL8, Fau, RPL13a species being the majority species. To subtract these sequences from AOLC1C library, the following procedure was performed: (1) library DNA was linearized using FseI restriction endonuclease, (2) primers specific to selected RPX and E. coli species were annealed to linearized DNA and (3) DNA synthesis extended from the primer using Bst DNA polymerase, a polymerase that lacks 3' exonuclease activity. Upon primer extension, the overhang of the FseI half-site adjacent to the insert will be lost, since extension products will yield blunt dsDNA. This blunted Fse I half-site will be incapable of adhering to the cohesive Fse I half-site present at the other end of the plasmid. Therefore, the DNA molecules to which primers have bound (e.g. the RPX and E. coli species) should have their Fse I sites blunted, and therefore be incapable of resealing by T4 DNA ligase. Hence, all linearized library DNA is treated with T4 DNA ligase, and the ligation products are transformed into electocompetent DH10B E.coli to generate a library enriched in sequences that do not contain RPX or E coli species. The enriched or subtracted library so created from the AOLC1C library was termed AOLC1CS. Sequencing of the AOLC1CS library showed that the targeted plasmids were substantially reduced in number, but they were still predominant species in the AOLC1CS library. Thus, the AOLC1CS library was subjected to another round of subtraction using the same method, with the resulting library termed AOLC1CS2 (AOLC1C library after 2 rounds of subtraction). The primers used in this method to make library AOLC1CS were: RPS5: 5'- TCG TTC GAG GAG CCC TTG GCA GCA T -3' (SEQ ID NO:119); RPL36A, 5'- CGC CCT TCC GCC ACG GCC GTC TCT -3' (SEQ ID NO:120); RPL18 5'- GAA AGG ACC CGT CGC CAT GGG CCG T-3' (SEO ID NO:121); Fau, 5'- CAG TCG CCA ATA TGC AGC TCT TTG T-3' (SEQ ID NO:122); RPL13A, 5'-CGA GGT ATG CTG CCC CAC AA-3' (SEQ ID NO:123). For library AOLC1CS2, the above primers were used and these primers were

added as well: RPS5, 5'- CGA GCG CCT GTG CAC AGC AGC CAG A -3' (SEQ ID NO:124); RPL36A, 5'- GCG GGA CAT GAT TCG GGA GGT GTG T -3' (SEQ ID NO:125); RPL8, 5'- CTG CGC GCC TGC GCG CCG TGG ATT T -3' (SEQ ID NO:126); Fau, 5'- CTT CGA GGT GAC CGG CCA GGA AAC G -3' (SEQ ID NO:127); RPL13A, 5'- CAG GCC GCT CTG GAC CGT CTC AAG G -3' (SEQ ID NO:128); *E coli*, 5'- AAC GGT GGG CTT GTT GCT GCT CTG G -3' (SEQ ID NO:129), 5'- ATT GGT ATT GGT AAC GGG CGT CAG G -3' (SEQ ID NO:130), 5'- ACC ATC TTC CAG GCG CAG TTG AGT T -3' (SEQ ID NO:131).

The target genes and encoded proteins identified by the present invention are explicitly disclosed in Table 1, which contains a common name for the gene and the GENBANK accession number, which can be retrieved from public sequence databases, as well as a sequence identifier for the nucleic acid sequence (first number) and encoded amino acid sequence (second number).

### 15 **Table 1**

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Accession	Common	Sequence Identifier	Description
Number	Name	(nucleic acid & protein)	Description
NM_001087	AAMP	SEQ ID NO:1 & 2	angio-associated, migratory cell protein
NM_001109	ADAM8	SEQ ID NO:3 & 4	a disintegrin and metalloproteinase domain 8
			a disintegrin-like and metalloprotease (reprolysin
NM_139057	ADAMTS17	SEQ ID NO:5 & 6	type) with thrombospondin type 1 motif, 17
NM_004036	ADCY3	SEQ ID NO:7 & 8	adenylate cyclase 3
NM_001619	ADRBK1	SEQ ID NO:9 & 10	adrenergic, beta, receptor kinase 1
NM_006698	BLCAP	SEQ ID NO:11 & 12	bladder cancer associated protein
NM_012264	C22orf5	SEQ ID NO:13 & 14	chromosome 22 open reading frame 5
NM_004356	CD81	SEQ ID NO:15 & 16	CD81 antigen (target of antiproliferative antibody 1)
NM_001769	CD9	SEQ ID NO:17 & 18	CD9 antigen (p24)
NM_001305	CLDN4	SEQ ID NO:19 & 20	claudin 4
NM_001288	CLIC1	SEQ ID NO:21 & 22	chloride intracellular channel 1
NM_058175	COL6A2	SEQ ID NO:23 & 24	collagen, type VI, alpha 2
AF070636 or			
NM_020428	CTL2	SEQ ID NO:25 & 26	CTL2 gene
NM_001397	ECE1	SEQ ID NO:27 & 28	endothelin converting enzyme 1
NM_004429	EFNB1	SEQ ID NO:29 & 30	ephrin-B1
NM_004475	FLOT2	SEQ ID NO:31 & 32	flotillin 2
AC011511 or			
BC058903	ICAM3	SEQ ID NO:33 & 34	intercellular adhesion molecule 3

Accession	Common	Sequence Identifier	
Number	Name	(nucleic acid & protein)	Description
NM_006123	IDS	SEQ ID NO:35 & 36	iduronate 2-sulfatase (Hunter syndrome)
NM_002226	JAG2	SEQ ID NO:37 & 38	jagged 2
BC001699	JAM1	SEQ ID NO:39 & 40	junctional adhesion molecule 1
NM_005567	LGALS3BP	SEQ ID NO:41 & 42	lectin, galactoside-binding, soluble, 3 binding protein
XM_085426	LOC146330	SEQ ID NO:43 & 44	similar to possible G-protein receptor
BC020590	LOC51107	SEQ ID NO:45 & 46	CGI-78 protein
NM_000237	LPL.	SEQ ID NO:47 & 48	lipoprotein lipase
NM_002335	LRP5	SEQ ID NO:49 & 50	low density lipoprotein receptor-related protein 5
NM_005581	LU	SEQ ID NO:51 & 52	Lutheran blood group (Auberger b antigen included)
NIM DOEBDO	M44C4	SEQ ID NO:53 & 54	membrane component, chromosome 11, surface marker 1
NM_005898 NM_007061	M11S1 MSE55		
L <u> </u>		SEQ ID NO:55 & 56	serum constituent protein
NM_006702	NTE	SEQ ID NO:57 & 58	neuropathy target esterase
			Homo sapiens cDNA FLJ31043 fis, clone
AK055605 or			HSYRA2000248 (PLEXIN A1) or Homo sapiens cDNA FLJ44113 fis, clone TESTI4046487, highly
AK126101	PLXNA1	SEQ ID NO:59 & 60	similar to Mus musculus plexin A1
			protein tyrosine phosphatase, receptor type, f
			polypeptide (PTPRF), interacting protein (liprin),
AF034800	PPFIA3		alpha 3
			Homo sapiens peptide-histidine transporter 4 (PTR4),
NM_145648	PTR4	SEQ ID NO:63 & 64	mRNA
			solute carrier family 16 (monocarboxylic acid
NM_004207	SLC16A3	SEQ ID NO:65 & 66	transporters), member 3
			solute carrier family 1 (neutral amino acid
NM_005628	SLC1A5	SEQ ID NO:67 & 68	transporter), member 5
NM_014437	SLC39A1	SEQ ID NO:69 & 70	solute carrier family 39 (zinc transporter), member 3
NM_021102	SPINT2	SEQ ID NO:71 & 72	serine protease inhibitor, Kunitz type, 2
NM_003714	STC2	SEQ ID NO:73 & 74	stanniocalcin 2
			tumor necrosis factor receptor superfamily, member
	TNFRSF21	SEQ ID NO:75 & 76	21
NM_003299	TRA1	SEQ ID NO:77 & 78	tumor rejection antigen (gp96) 1
			transient receptor potential cation channel, subfamily
NM_017636	TRPM4	SEQ ID NO:79 & 80	M, member 4

It should be understood that the foregoing disclosure emphasizes certain specific embodiments of the invention and that all modifications or alternatives equivalent thereto are within the spirit and scope of the invention as set forth in the appended claims.

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### WHAT WE CLAIM IS:

A method for identifying a compound for inducing apoptosis, comprising 1. identifying an inhibitor of a target selected from the group consisting of: angioassociated, migratory cell protein (AAMP, comprising SEQ ID NO:2), a disintegrin and metalloproteinase domain 8 (ADAM8, comprising SEQ ID NO:4), a disintegrin-like and metalloprotease (reporlysin type) with thrombospondin type 1 motif, 17 (ADAMTS17, comprising SEQ ID NO:6), adenylate cyclase 3 (ADCY3, comprising SEQ ID NO:8), adrenergic beta receptor kinase 1 (ADRBK1, comprising SEQ ID NO:10), bladder cancer associated protein (BLCAP, comprising SEQ ID NO:12), chromosome 22 open reading frame 5 (C22orf5, comprising SEQ ID NO:14), CD81 antigen (target of antiproliferative antibody 1 (CD81, comprising SEQ ID NO:16), CD9 antigen (p24) (CD9, comprising SEO ID NO:18), claudin 4 (CLDN4, comprising SEQ ID NO:20), chloride intracellular channel 1 (CLIC1, comprising SEQ ID NO:22), collagen, type VI, alpha 2 (COL6A2, comprising SEQ ID NO:24), CTL2 (CTL2, comprising SEQ ID NO:26), endothelin converting enzyme 1 (ECE1, comprising SEQ ID NO:28), ephrin-B1 (EFNB1, comprising SEQ ID NO:30), flotillin 2 (FLOT2, comprising SEQ ID NO:32), intercellular adhesion molecule 3 (ICAM3, comprising SEQ ID NO:34), iduronate 2sulfatase (Hunter syndrome) (IDS, comprising SEQ ID NO:36), jagged 2 (JAG2, comprising SEQ ID NO:38), junctional adhesion molecule 1 (JAM1, comprising SEQ ID NO:40), lectin, galactoside-binding soluble 3 binding protein (LGALS3BP, comprising SEO ID NO:42), similar to possible G-protein receptor (LOC146330, comprising SEQ ID NO:44), CGI-78 protein (LOC51107, comprising SEQ ID NO:46), lipoprotein lipase (LPL, comprising SEQ ID NO:48), low density lipoprotein receptor-related protein 5 (LRP5, comprising SEQ ID NO:50), Lutheran blood group (Auberger b antigen included) (LU, comprising SEO ID NO:52), membrane component, chromosome 11, surface marker 1 (M11S1, comprising SEQ ID NO:54), serum constituent protein (MSE55, comprising SEQ ID NO:56), neuropathy target esterase (NTE, comprising SEQ ID NO:58), Homo sapiens cDNA FL31043 fis, clone HSYRA2000248 (PLEXIN A1) or Homo sapiens cDNA FLJ44113 fis, clone TESTI4046487, highly similar to Mus musculus plexin A1 (PLXNA1, comprising SEQ ID NO:60), protein tyrosine phosphatase, receptor type, f polypeptide (PTPRF), interacting protein )(liprin), alpha 3 (PPFIA3, comprising SEQ ID NO:62), Homo sapiens peptide-histidine transporter 4 (PTR4), mRNA (PTR4, comprising SEQ ID NO:64), solute carrier family 16

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(moncarboxylic acid transporters) member 3 (SLC16A3, comprising SEQ ID NO:66), solute carrier family 1 (neutral amino acid transporter) member 5 (SLC1A5, comprising SEQ ID NO:68), solute carrier family 39 (zinc transporter) member 3 (SLC39A1, comprising SEQ ID NO:70), serine protease inhibitor, Kunitz type 2 (SPINT2, comprising SEQ ID NO:72), stanniocalcin 2 (STC2, comprising SEQ ID NO:74), tumor necrosis receptor superfamily member 21 (TNFRSF21, comprising SEQ ID NO:76), tumor rejection antigen (gp96) 1 (TRA1, comprising SEQ ID NO:78), and transient receptor potential cation channel, subfamily M member 4 (TRPM4, comprising SEQ ID NO:80).

- 10 2. The method of Claim 1, further comprising assessing the ability of an identified inhibitor to induce apoptosis in a cell.
  - 3. The method of Claim 2, further comprising detecting whether a compound identified as inducing apoptosis inhibits growth of tumor cells.
- 4. The method of Claim 1, wherein the step of identifying comprises identifying an inhibitor of expression or activity of the target.
  - 5. The method of Claim 1, comprising the steps of:
  - a) contacting a host cell with a putative regulatory compound, wherein the host cell expresses the target or a biologically active fragment thereof; and
  - b) detecting whether the putative regulatory compound inhibits the target or biologically active fragment thereof, wherein a putative regulatory compound that inhibits the target as compared to in the absence of the compound is indicated to be a candidate compound for the induction of apoptosis in a host cell.
    - 6. The method of Claim 5, wherein the host cell is a tumor cell line.
  - 7. The method of Claim 5, wherein the step of detecting is selected from the group consisting of:
    - a) detecting expression of the target in the presence of the putative regulatory compound; and
    - b) detecting activity of the target in the presence of the putative regulatory compound.
  - 8. The method of Claim 7, wherein the expression of the target is measured by polymerase chain reaction.
    - 9. The method of Claim 7, wherein the expression of the target is measured using an antibody or antigen binding partner that selectively binds to the target.

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- 10. The method of Claim 7, wherein the activity of the target is measured by measuring the amount of a product generated in a biochemical reaction mediated by the target.
- 11. The method of Claim 7, wherein the activity of the target is measured by measuring the amount of a substrate consumed in a biochemical reaction mediated by the target.
  - 12. The method of Claim 1, comprising the steps of:
  - a) determining the three-dimensional structure of the target;
- b) identifying the three-dimensional structure of a putative inhibitor by using
   computer software to model an interaction between the target structure and a structure of a test compound; and
  - c) synthesizing compounds identified in (b) and assaying the compounds in an *in vitro* assay to determine whether the compound inhibits the expression or activity of the target.
- 15 13. The method of Claim 1, wherein the target has been validated as being involved in tumor cell growth.
  - 14. The method of Claim 14, wherein the target has been validated as being involved in tumor cell growth by a process comprising:
- a) inhibiting the target in a cell by a method selected from the group
   20 consisting of gene knock-out, anti-sense oligonucleotide expression, use of RNAi molecules and GSE expression; and
  - b) assaying the cell for the ability of the cell to grow.
  - 15. A method for inducing apoptosis, comprising inhibiting the expression or activity of a target or a gene encoding the target, wherein the target is selected from the group consisting of: angio-associated, migratory cell protein (AAMP, comprising SEQ ID NO:2), a disintegrin and metalloproteinase domain 8 (ADAM8, comprising SEQ ID NO:4), a disintegrin-like and metalloprotease (reporlysin type) with thrombospondin type 1 motif, 17 (ADAMTS17, comprising SEQ ID NO:6), adenylate cyclase 3 (ADCY3, comprising SEQ ID NO:8), adrenergic beta receptor kinase 1 (ADRBK1, comprising SEQ ID NO:10), bladder cancer associated protein (BLCAP, comprising SEQ ID NO:12), chromosome 22 open reading frame 5 (C22orf5, comprising SEQ ID NO:14), CD81 antigen (target of antiproliferative antibody 1 (CD81, comprising SEQ ID NO:16), CD9 antigen (p24) (CD9, comprising SEQ ID NO:18), claudin 4 (CLDN4, comprising SEQ ID

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NO:20), chloride intracellular channel 1 (CLIC1, comprising SEQ ID NO:22), collagen. type VI, alpha 2 (COL6A2, comprising SEQ ID NO:24), CTL2 (CTL2, comprising SEQ ID NO:26), endothelin converting enzyme 1 (ECE1, comprising SEQ ID NO:28), ephrin-B1 (EFNB1, comprising SEQ ID NO:30), flotillin 2 (FLOT2, comprising SEQ ID NO:32), intercellular adhesion molecule 3 (ICAM3, comprising SEQ ID NO:34), iduronate 2-sulfatase (Hunter syndrome) (IDS, comprising SEQ ID NO:36), jagged 2 (JAG2, comprising SEQ ID NO:38), junctional adhesion molecule 1 (JAM1, comprising SEO ID NO:40), lectin, galactoside-binding soluble 3 binding protein (LGALS3BP, comprising SEQ ID NO:42), similar to possible G-protein receptor (LOC146330, comprising SEQ ID NO:44), CGI-78 protein (LOC51107, comprising SEQ ID NO:46), lipoprotein lipase (LPL, comprising SEQ ID NO:48), low density lipoprotein receptorrelated protein 5 (LRP5, comprising SEQ ID NO:50), Lutheran blood group (Auberger b antigen included) (LU, comprising SEQ ID NO:52), membrane component, chromosome 11, surface marker 1 (M11S1, comprising SEQ ID NO:54), serum constituent protein (MSE55, comprising SEQ ID NO:56), neuropathy target esterase (NTE, comprising SEQ ID NO:58), Homo sapiens cDNA FL31043 fis, clone HSYRA2000248 (PLEXIN A1) or Homo sapiens cDNA FLJ44113 fis, clone TESTI4046487, highly similar to Mus musculus plexin A1 (PLXNA1, comprising SEQ ID NO:60), protein tyrosine phosphatase, receptor type, f polypeptide (PTPRF), interacting protein )(liprin), alpha 3 (PPFIA3, comprising SEQ ID NO:62), Homo sapiens peptide-histidine transporter 4 (PTR4), mRNA (PTR4, comprising SEQ ID NO:64), solute carrier family 16 (moncarboxylic acid transporters) member 3 (SLC16A3, comprising SEQ ID NO:66), solute carrier family 1 (neutral amino acid transporter) member 5 (SLC1A5, comprising SEQ ID NO:68), solute carrier family 39 (zinc transporter) member 3 (SLC39A1, comprising SEQ ID NO:70), serine protease inhibitor, Kunitz type 2 (SPINT2, comprising SEQ ID NO:72), stanniocalcin 2 (STC2, comprising SEQ ID NO:74), tumor necrosis receptor superfamily member 21 (TNFRSF21, comprising SEQ ID NO:76), tumor rejection antigen (gp96) 1 (TRA1, comprising SEQ ID NO:78), and transient receptor potential cation channel, subfamily M member 4 (TRPM4, comprising SEO ID NO:80).

16. The method of Claim 15, wherein the step of inhibiting is conducted by contacting a cell with an inhibitor of the target, wherein the inhibitor induces apoptosis in the cell.

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- 17. A method for the diagnosis of a tumor comprising:
- detecting a level of expression or activity of at least one biomarker in a test a) sample from a patient to be diagnosed, wherein the biomarker is selected from the group consisting of: angio-associated, migratory cell protein (AAMP, comprising SEO ID NO:2), a disintegrin and metalloproteinase domain 8 (ADAM8, comprising SEO ID NO:4), a disintegrin-like and metalloprotease (reporlysin type) with thrombospondin type 1 motif, 17 (ADAMTS17, comprising SEQ ID NO:6), adenylate cyclase 3 (ADCY3, comprising SEQ ID NO:8), adrenergic beta receptor kinase 1 (ADRBK1, comprising SEQ ID NO:10), bladder cancer associated protein (BLCAP, comprising SEQ ID NO:12), chromosome 22 open reading frame 5 (C22orf5, comprising SEQ ID NO:14), CD81 antigen (target of antiproliferative antibody 1 (CD81, comprising SEQ ID NO:16), CD9 antigen (p24) (CD9, comprising SEQ ID NO:18), claudin 4 (CLDN4, comprising SEQ ID NO:20), chloride intracellular channel 1 (CLIC1, comprising SEQ ID NO:22), collagen, type VI, alpha 2 (COL6A2, comprising SEQ ID NO:24), CTL2 (CTL2, comprising SEQ ID NO:26), endothelin converting enzyme 1 (ECE1, comprising SEQ ID NO:28), ephrin-B1 (EFNB1, comprising SEQ ID NO:30), flotillin 2 (FLOT2, comprising SEQ ID NO:32), intercellular adhesion molecule 3 (ICAM3, comprising SEQ ID NO:34), iduronate 2-sulfatase (Hunter syndrome) (IDS, comprising SEQ ID NO:36), jagged 2 (JAG2, comprising SEQ ID NO:38), junctional adhesion molecule 1 (JAM1, comprising SEQ ID NO:40), lectin, galactoside-binding soluble 3 binding protein (LGALS3BP, comprising SEO ID NO:42), similar to possible G-protein receptor (LOC146330, comprising SEQ ID NO:44), CGI-78 protein (LOC51107, comprising SEQ ID NO:46), lipoprotein lipase (LPL, comprising SEQ ID NO:48), low density lipoprotein receptorrelated protein 5 (LRP5, comprising SEQ ID NO:50), Lutheran blood group (Auberger b antigen included) (LU, comprising SEQ ID NO:52), membrane component, chromosome 11, surface marker 1 (M11S1, comprising SEQ ID NO:54), serum constituent protein (MSE55, comprising SEQ ID NO:56), neuropathy target esterase (NTE, comprising SEQ ID NO:58), Homo sapiens cDNA FL31043 fis, clone HSYRA2000248 (PLEXIN A1) or Homo sapiens cDNA FLJ44113 fis, clone TESTI4046487, highly similar to Mus musculus plexin A1 (PLXNA1, comprising SEQ ID NO:60), protein tyrosine phosphatase, receptor type, f polypeptide (PTPRF), interacting protein )(liprin), alpha 3 (PPFIA3, comprising SEQ ID NO:62), Homo sapiens peptide-histidine transporter 4 (PTR4), mRNA (PTR4, comprising SEQ ID NO:64), solute carrier family 16

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(moncarboxylic acid transporters) member 3 (SLC16A3, comprising SEQ ID NO:66), solute carrier family 1 (neutral amino acid transporter) member 5 (SLC1A5, comprising SEQ ID NO:68), solute carrier family 39 (zinc transporter) member 3 (SLC39A1, comprising SEQ ID NO:70), serine protease inhibitor, Kunitz type 2 (SPINT2, comprising SEQ ID NO:72), stanniocalcin 2 (STC2, comprising SEQ ID NO:74), tumor necrosis receptor superfamily member 21 (TNFRSF21, comprising SEQ ID NO:76), tumor rejection antigen (gp96) 1 (TRA1, comprising SEQ ID NO:78), and transient receptor potential cation channel, subfamily M member 4 (TRPM4, comprising SEQ ID NO:80);

- b) comparing the level of expression or activity of the biomarker in the test sample to a baseline level of biomarker expression or activity established from a control sample; wherein detection of a statistically significant difference in the expression or activity of the biomarker in the test sample, as compared to the baseline level of the expression or biological activity of the biomarker, is an indicator of a difference in the tumorigenicity or potential therefore of cells in the patient.
  - 18. The method of Claim 17, wherein the step of detecting comprises detecting biomarker mRNA transcription in the test sample.
  - 19. The method of Claim 18, wherein the step of detecting is by a method selected from the group consisting of polymerase chain reaction (PCR), reverse transcriptase-PCR (RT-PCR), *in situ* hybridization, Northern blot, sequence analysis, gene microarray analysis, and detection of a reporter gene.
  - 20. The method of Claim 17, wherein the step of detecting comprises detecting the biomarker protein in the test sample.
  - 21. The method of Claim 20, wherein the step of detecting is by a method selected from the group consisting of immunoblot, enzyme-linked immunosorbant assay (ELISA), radioimmunoassay (RIA), immunoprecipitation, immunohistochemistry and immunofluorescence.
    - 22. The method of Claim 17, wherein the step of detecting comprises detecting biomarker biological activity in the test sample.
- 30 23. The method of Claim 17, wherein detection of a statistically significant difference in the level of biomarker expression or activity in the test sample as compared to the baseline level, with a confidence of p<0.05, indicates that the cells in the test

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sample have a difference in tumorigenicity or potential therefore as compared to the control sample.

- 24. The method of Claim 17, wherein the test sample is from a patient being diagnosed for cancer and wherein the baseline level is established from a control sample that is established as non-tumorigenic.
- 25. The method of Claim 24, wherein an increase in the level of biomarker expression or activity of the test sample as compared to the baseline level of expression or activity indicates that cells from which the test sample was derived are predicted to be tumorigenic or predisposed to becoming tumorigenic.
- 26. The method of Claim 17, wherein the test sample is from a patient who is known to have cancer, and wherein the baseline level comprises a level of biomarker expression or activity from a previous tumor cell sample from the patient;

wherein a statistically significant decrease in the level of biomarker expression or activity in the test sample as compared to the first baseline level of expression or activity from the previous tumor cell sample, indicates that the test sample is less tumorigenic than the previous tumor cell sample;

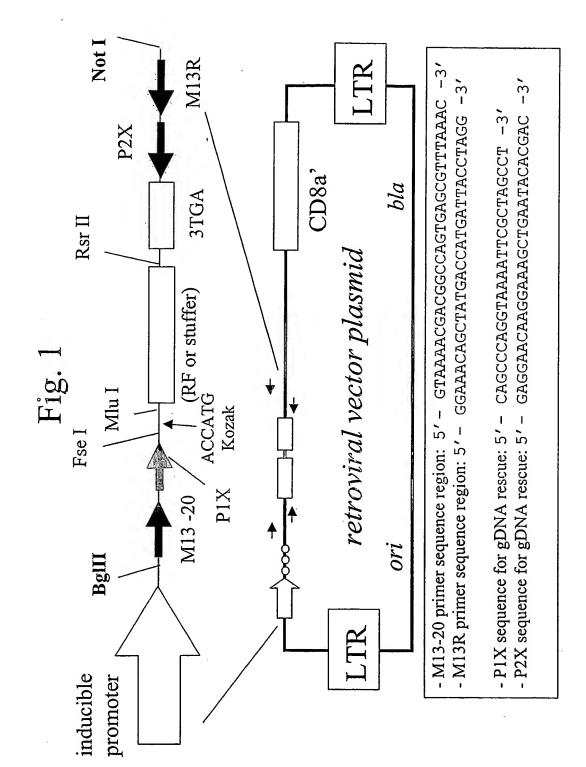
and wherein a statistically significant increase in the level of biomarker expression or activity in the test sample as compared to the first baseline level of expression or activity, indicates that the test sample is more tumorigenic than the previous tumor cell sample.

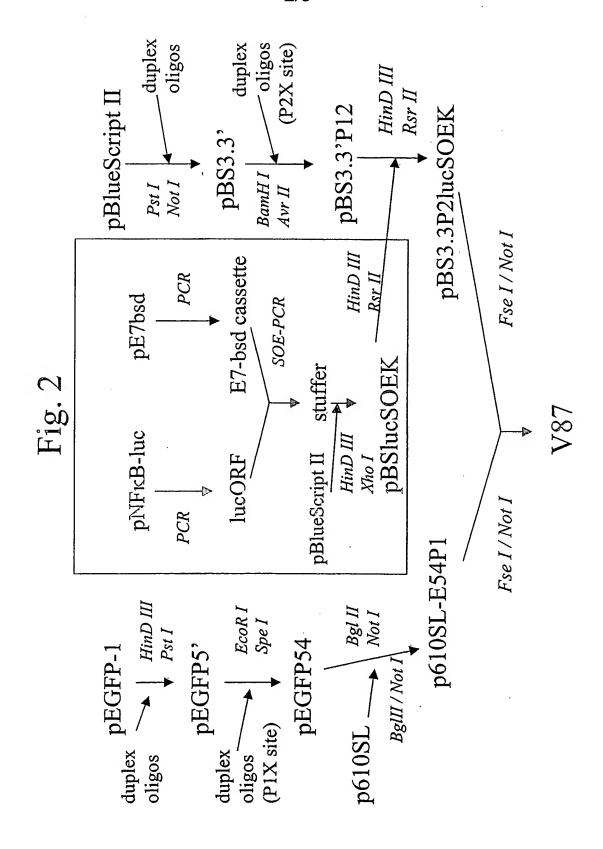
- 27. The method of Claim 26, wherein the method further comprises a step (c) of modifying cancer treatment for the patient based on whether an increase or decrease in tumorigenicity is indicated in step (b).
- 28. The method of Claim 17, wherein the baseline level is established by a method selected from the group consisting of:
  - (1) establishing a baseline level of biomarker expression or activity in an autologous control sample from the patient, wherein the autologous sample is from a same cell type, tissue type or bodily fluid type as the test sample of step (a);
- (2) establishing a baseline level of biomarker expression or activity from at least one previous detection of biomarker expression or activity in a previous test sample from the patient, wherein the previous test sample was of a same cell type, tissue type or bodily fluid type as the test sample of step (a); and

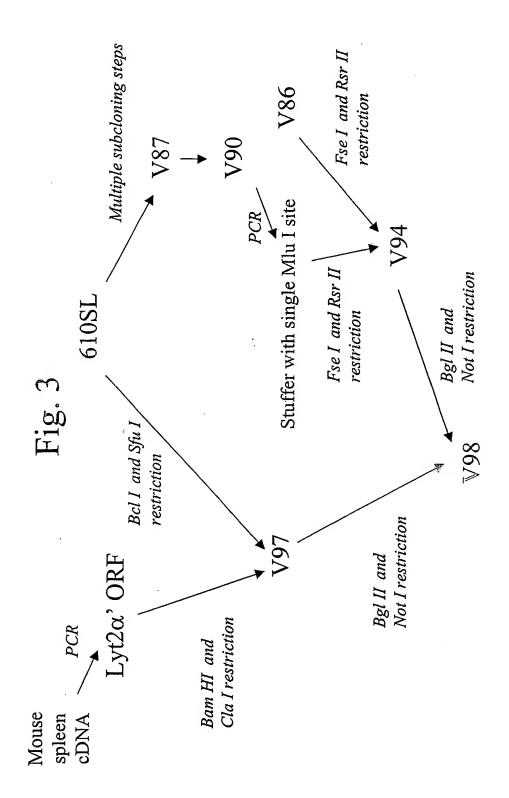
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- (3) establishing a baseline level of biomarker expression or activity from an average of control samples of a same cell type, tissue type or bodily fluid type as the test sample of step (a), the control samples having been obtained from a population of matched individuals.
- 29. The method of Claim 17, wherein the patient test sample is immobilized on a substrate.
  - 30. The method of Claim 17, wherein the test sample is a bodily fluid sample.
- 31. The method of Claim 17, wherein the biomarker level is determined by contacting the patient test sample with an antibody or a fragment thereof that selectively binds specifically to the biomarker, and determining whether the antibody or fragment thereof has bound to the marker.
- 32. The method of Claim 17, wherein the method is used to determine the prognosis for cancer in the patient.
- 33. The method of Claim 17, wherein the method is used to determine the susceptibility of the patient to a therapeutic treatment.







## SEQUENCE LISTING

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<sup>824</sup> PRT Homo sapiens

Ser Gly Tyr Thr Glu Thr Tyr Thr Ala Ala Asn Gly Ser Glu Val Thr Glu Gln Pro Arg Gly Gln Asp His Cys Leu Tyr Gln Gly His Val Glu 100 100 110 Gly Tyr Pro Asp Ser Ala Ala Ser Leu Ser Thr Cys Ala Gly Leu Arg Gly Phe Phe Gln Val Gly Ser Asp Leu His Leu Ile Glu Pro Leu Asp 130 Glu Gly Gly Gly Gly Arg His Ala Val Tyr Gln Ala Glu His Leu 145 150 160 Leu Gln Thr Ala Gly Thr Cys Gly Val Ser Asp Asp Ser Leu Gly Ser Leu Leu Gly Pro Arg Thr Ala Ala Val Phe Arg Pro Arg Pro Gly Asp Ser Leu Pro Ser Arg Glu Thr Arg Tyr Val Glu Leu Tyr Val Val Val 195 Asp Asn Ala Glu Phe Gln Met Leu Gly Ser Glu Ala Ala Val Arg His Arg Val Leu Glu Val Val Asn His Val Asp Lys Leu Tyr Gln Lys Leu 225 235 240 Asn Phe Arg Val Val Leu Val Gly Leu Glu Ile Trp Asn Ser Gln Asp 245 Arg Phe His Val Ser Pro Asp Pro Ser Val Thr Leu Glu Asn Leu Leu 265 270 Thr Trp Gln Ala Arg Gln Arg Thr Arg Arg His Leu His Asp Asn Val Gln Leu Ile Thr Gly Val Asp Phe Thr Gly Thr Thr Val Gly Phe Ala 290 300 Arg Val Ser Ala Met Cys Ser His Ser Ser Gly Ala Val Asn Gln Asp 305 His Ser Lys Asn Pro Val Gly Val Ala Cys Thr Met Ala His Glu Met 325 Gly His Asn Leu Gly Met Asp His Asp Glu Asn Val Gln Gly Cys Arg Cys Gln Glu Arg Phe Glu Ala Gly Arg Cys Ile Met Ala Gly Ser Ile Gly Ser Ser Phe Pro Arg Met Phe Ser Asp Cys Ser Gln Ala Tyr Leu Glu Ser Phe Leu Glu Arg Pro Gln Ser Val Cys Leu Ala Asn Ala Pro

Asp Leu Ser His Leu Val Gly Gly Pro Val Cys Gly Asn Leu Phe Val Glu Arg Gly Glu Gln Cys Asp. Cys Gly Pro Pro Glu Asp Cys Arg Asn 420 425 Arg Cys Cys Asn Ser Thr Thr Cys Gln Leu Ala Glu Gly Ala Gln Cys Ala His Gly Thr Cys Cys Gln Glu Cys Lys Val Lys Pro Ala Gly Glu Leu Cys Arg Pro Lys Lys Asp Met Cys Asp Leu Glu Glu Phe Cys Asp 465 Gly Arg His Pro Glu Cys Pro Glu Asp Ala Phe Gln Glu Asn Gly Thr Pro Cys Ser Gly Gly Tyr Cys Tyr Asn Gly Ala Cys Pro Thr Leu Ala 500 505 Gln Gln Cys Gln Ala Phe Trp Gly Pro Gly Gln Ala Ala Glu Glu 515 Ser Cys Phe Ser Tyr Asp Ile Leu Pro Gly Cys Lys Ala Ser Arg Tyr 530 540 Arg Ala Asp Met Cys Gly Val Leu Gln Cys Lys Gly Gly Gln Gln Pro Leu Gly Arg Ala Ile Cys Ile Val Asp Val Cys His Ala Leu Thr Thr Glu Asp Gly Thr Ala Tyr Glu Pro Val Pro Glu Gly Thr Arg Cys Gly Pro Glu Lys Val Cys Trp Lys Gly Arg Cys Gln Asp Leu His Val Tyr Arg Ser Ser Asn Cys Ser Ala Gln Cys His Asn His Gly Val Cys Asn 610 620 His Lys Gln Glu Cys His Cys His Ala Gly Trp Ala Pro Pro His Cys 625 635 640 Ala Lys Leu Leu Thr Glu Val His Ala Ala Ser Gly Ser Leu Pro Val 645 655 Leu Val Val Val Leu Val Leu Leu Ala Val Leu Val Thr Leu 660 665 670 Ala Gly Ile Ile Val Tyr Arg Lys Ala Arg Ser Arg Ile Leu Ser Arg Asn Val Ala Pro Lys Thr Thr Met Gly Arg Ser Asn Pro Leu Phe His 690 700 Gln Ala Ala Ser Arg Val Pro Ala Lys Gly Gly Ala Pro Ala Pro Ser

Arg Gly Pro Gln Glu Leu Val Pro Thr His Pro Gly Gln Pro Ala 735 Arg His Pro Ala Ser Ser Val Ala Leu Lys Arg Pro Pro Pro Ala Pro 740 745 Pro Val Thr Val Ser Ser Pro Pro Phe Pro Val Pro Val Tyr Thr Arg 755 760 765 Gln Ala Pro Lys Gln Val Ile Lys Pro Thr Phe Ala Pro Pro Val Pro 770 780 Pro Val Lys Pro Gly Ala Gly Ala Ala Asn Pro Gly Pro Ala Glu Gly 785 795 800 Ala Val Gly Pro Lys Val Ala Leu Lys Pro Pro Ile Gln Arg Lys Gln 815 Gly Ala Gly Ala Pro Thr Ala Pro

<210> <211> <212> <213> 5 3470 DNA

Homo sapiens

CDS (24)..(3311)

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V	VO 20	04/07	6682											P	CT/U	<b>S2004/00602</b> 0
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acc Thr	ccc Pro	agc Ser	cct Pro 190	tct Ser	gct Ala	gag Glu	gcc Ala	cag Gln 195	aga Arg	cct Pro	gag Glu	cag Gln	ctc Leu 200	tgc Cys	aag Lys	629
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tgg Trp	cgg Arg 220	gag Glu	cgg Arg	agg Arg	aac Asn	gct Ala 225	atc Ile	cgg Arg	ctc Leu	acc Thr	agc Ser 230	gag Glu	cac His	acg Thr	gtg Val	725
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atg Met	ttt Phe	cag Gln	cac His 270	cag Gln	agc ser	ctg Leu	ggg GTy	att Ile 275	aaa Lys	att Ile	aac Asn	att Ile	caa Gln 280	gtg Val	acc Thr	869
aag Lys	ctt Leu	gtc Val 285	ctg Leu	cta Leu	cga Arg	caa Gln	cgt Arg 290	ccc Pro	gct Ala	aag Lys	ttg Leu	tcc Ser 295	att Ile	ggg GTy	cac His	917
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tat Tyr 315	gga Gly	gga Gly	gcg Ala	cga Arg	tac Tyr 320	ctc Leu	ggc Gly	aat Asn	aac Asn	cag Gln 325	gtt Val	CCC Pro	ggc Gly	ggg Gly	aag Lys 330	1013
gac Asp	gac Asp	ccg Pro	ccc Pro	ctg Leu 335	gtg Val	gat Asp	gct Ala	gct Ala	gtg Val 340	ttt Phe	gtg Val	acc Thr	agg Arg	aca Thr 345	gat Asp	1061
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cac His	atc Ile	atg Met	tca Ser	gga Gly 415	gag Glu	tgg Trp	gtg Val	aaa Lys	ggc Gly 420	cgg Arg	aac Asn	cca Pro	agt Ser	gac Asp 425	ctc Leu	1301
tct Ser	tgg Trp	tcc Ser	tcc Ser 430	tgc Cys	agc Ser	cga Arg	gat Asp	gac Asp 435	ctt Leu	gaa Glu	aac Asn	ttc Phe	ctc Leu 440	aag Lys	tca Ser	1349
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gta Val	cgc Arg 460	ctc Leu	ccg Pro	cac His	aag Lys	ctg Leu 465	ccg Pro	ggc Gly	atg Met	cac His	tac Tyr 470	agt Ser	gcc Ala	aac Asn	gag Glu	1445
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( Jan

W	O 200	4/076	682											PC	CT/US2	004/006020
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tcc ser	tgc Cys	aag Lys	acc Thr 510	aag Lys	ctg Leu	gac Asp	cct Pro	CCC Pro 515	ctg Leu	gat Asp	ggc Gly	acc Thr	gag Glu 520	tgt Cys	ggg Gly	1589
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ccg Pro	gag Glu 540	cat His	gtg Val	gac Asp	gga Gly	gac Asp 545	tgg Trp	agc ser	ccg Pro	tgg Trp	ggc Gly 550	gcc Ala	tgg Trp	agc Ser	atg Met	1685
tgc Cys 555	agc Ser	cga Arg	aca Thr	tgt Cys	ggg GTy 560	acg Thr	gga Gly	gcc Ala	cgc Arg	ttc Phe 565	agg Arg	cag Gln	agg Arg	aaa Lys	tgt Cys 570	1733
gac Asp	aac Asn	ccc Pro	ccc Pro	cct Pro 575	ggg GTy	cct Pro	gga Gly	ggc GTy	aca Thr 580	cac His	tgc Cys	ccg Pro	ggt Gly	gcc Ala 585	agt Ser	1781
gta Val	gaa Glu	cat His	gcg Ala 590	gtc Val	tgc Cys	gag Glu	aac Asn	ctg Leu 595	ccc Pro	tgc Cys	ccc Pro	aag Lys	ggt Gly 600	ctg Leu	ccc Pro	1829
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ctc Leu 635	tac Tyr	tgc Cys	tcg Ser	ccc Pro	ctc Leu 640	ggg Gly	aag Lys	gag Glu	tcc Ser	cca Pro 645	ctg Leu	ctg Leu	gtg Val	gcc Ala	gac Asp 650	1973
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gtg Val	cac His	ggc Gly	aag Lys 670	tgc Cys	cag Gln	aaa Lys	atc Ile	ggc Gly 675	tgt Cys	gac Asp	ggc Gly	atc Ile	atc Ile 680	ggg GTy	tct Ser	2069
gca Ala	gcc Ala	aaa Lys 685	gag Glu	gac Asp	aga Arg	tgc Cys	ggg G1y 690	gtc Val	tgc Cys	agc Ser	ggg Gly	gac Asp 695	ggc Gly	aag Lys	acc Thr	2117
tgc Cys	cac His 700	ttg Leu	gtg Val	aag Lys	ggc Gly	gac Asp 705	ttc Phe	agc Ser	cac His	gcc Ala	cgg Arg 710	ggg GTy	aca Thr	gct Ala	ctc Leu	2165
aaa Lys 715	gac Asp	tcg Ser	ggt Gly	aag Lys	999 G1y 720	tcc Ser	atc Ile	aac Asn	agt Ser	gac Asp 725	tgg Trp	aag Lys	ata Ile	gag Glu	ctc Leu 730	2213
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gaa Glu	tac Tyr 780	act Thr	gtt Val	cct Pro	gta Val	aac Asn 785	cgc Arg	act Thr	gcg Ala	gaa Glu	aat Asn 790	caa Gln	agc Ser	gaa Glu	cca Pro	2405
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3470

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<212> PRT <213> Homo sapiens

<400> 6

Met Cys Asp Gly Ala Leu Leu Pro Pro Leu Val Leu Pro Val Leu Leu Leu Leu Val Trp Gly Leu Asp Pro Gly Thr Ala Val Gly Asp Ala Ala  $\begin{array}{c} 20 \\ 20 \end{array}$ Ala Asp Val Glu Val Val Leu Pro Trp Arg Val Arg Pro Asp Asp Val His Leu Pro Pro Leu Pro Ala Ala Pro Gly Pro Arg Arg Arg Arg 50 60 60 Pro Arg Thr Pro Pro Ala Ala Pro Arg Ala Arg Pro Gly Glu Arg Ala 65 75 80 Leu Leu His Leu Pro Ala Phe Gly Arg Asp Leu Tyr Leu Gln Leu 85 90 95 Arg Arg Asp Leu Arg Phe Leu Ser Arg Gly Phe Glu Val Glu Glu Ala Gly Ala Ala Arg Arg Gly Arg Pro Ala Glu Leu Cys Phe Tyr Ser 115 125 Gly Arg Val Leu Gly His Pro Gly Ser Leu Val Ser Leu Ser Ala Cys Gly Ala Ala Gly Gly Leu Val Gly Leu Ile Gln Leu Gly Gln Glu Gln 145 150 160 Val Leu Ile Gln Pro Leu Asn Asn Ser Gln Gly Pro Phe Ser Gly Arg Glu His Leu Ile Arg Arg Lys Trp Ser Leu Thr Pro Ser Pro Ser Ala Glu Ala Gln Arg Pro Glu Gln Leu Cys Lys Val Leu Thr Glu Lys Lys Lys Pro Thr Trp Gly Arg Pro Ser Arg Asp Trp Arg Glu Arg Arg Asn 210 215Ala Ile Arg Leu Thr Ser Glu His Thr Val Glu Thr Leu Val Val Ala 225 230 235 Ile Leu Thr Val Met Asn Met Val Tyr Asn Met Phe Gln His Gln Ser Leu Gly Ile Lys Ile Asn Ile Gln Val Thr Lys Leu Val Leu Leu Arg 280 285

Gln Arg Pro Ala Lys Leu Ser Ile Gly His His Gly Glu Arg Ser Leu 290 300 Glu Ser Phe Cys His Trp Gln Asn Glu Glu Tyr Gly Gly Ala Arg Tyr 305 315 320 Leu Gly Asn Asn Gln Val Pro Gly Gly Lys Asp Asp Pro Pro Leu Val Asp Ala Ala Val Phe Val Thr Arg Thr Asp Phe Cys Val His Lys Asp 340 350 Glu Pro Cys Asp Thr Val Gly Ile Ala Tyr Leu Gly Gly Val Cys Ser 355 Ala Lys Arg Lys Cys Val Leu Ala Glu Asp Asn Gly Leu Asn Leu Ala 370 380 Phe Thr Ile Ala His Glu Leu Gly His Asn Leu Gly Met Asn His Asp 395 400 Asp Asp His Ser Ser Cys Ala Gly Arg Ser His Ile Met Ser Gly Glu
415 Trp Val Lys Gly Arg Asn Pro Ser Asp Leu Ser Trp Ser Ser Cys Ser Arg Asp Asp Leu Glu Asn Phe Leu Lys Ser Lys Val Ser Thr Cys Leu 435 Leu Val Thr Asp Pro Arg Ser Gln His Thr Val Arg Leu Pro His Lys Leu Pro Gly Met His Tyr Ser Ala Asn Glu Gln Cys Gln Ile Leu Phe 465 470 480 Gly Met Asn Ala Thr Phe Cys Arg Asn Met Glu His Leu Met Cys Ala 485 490 491 Gly Leu Trp Cys Leu Val Glu Gly Asp Thr Ser Cys Lys Thr Lys Leu 500 505 Asp Pro Pro Leu Asp Gly Thr Glu Cys Gly Ala Asp Lys Trp Cys Arg 515 Ala Gly Glu Cys Val Ser Lys Thr Pro Ile Pro Glu His Val Asp Gly 530 Asp Trp Ser Pro Trp Gly Ala Trp Ser Met Cys Ser Arg Thr Cys Gly 545 Thr Gly Ala Arg Phe Arg Gln Arg Lys Cys Asp Asn Pro Pro Gly Pro Gly Gly Thr His Cys Pro Gly Ala Ser Val Glu His Ala Val Cys 580 585 Glu Asn Leu Pro Cys Pro Lys Gly Leu Pro Ser Phe Arg Asp Gln Gln
595
600 WO 2004/076682 PCT/US2004/006020

Cys Gln Ala His Asp Arg Leu Ser Pro Lys Lys Gly Leu Leu Thr 610 620 Ala Val Val Asp Asp Lys Pro Cys Glu Leu Tyr Cys Ser Pro Leu 625 635 Gly Lys Glu Ser Pro Leu Leu Val Ala Asp Arg Val Leu Asp Gly Thr 645 655 Pro Cys Gly Pro Tyr Glu Thr Asp Leu Cys Val His Gly Lys Cys Gln 660 670 Lys Ile Gly Cys Asp Gly Ile Ile Gly Ser Ala Ala Lys Glu Asp Arg Cys Gly Val Cys Ser Gly Asp Gly Lys Thr Cys His Leu Val Lys Gly Asp Phe Ser His Ala Arg Gly Thr Ala Leu Lys Asp Ser Gly Lys Gly Ser Ile Asn Ser Asp Trp Lys Ile Glu Leu Pro Gly Glu Phe Gln Ile Ala Gly Thr Thr Val Arg Tyr Val Arg Arg Gly Leu Trp Glu Lys Ile
740
750 Ser Ala Lys Gly Pro Thr Lys Leu Pro Leu His Leu Met Val Leu Leu
755 760 765 Phe His Asp Gln Asp Tyr Gly Ile His Tyr Glu Tyr Thr Val Pro Val Asn Arg Thr Ala Glu Asn Gln Ser Glu Pro Glu Lys Pro Gln Asp Ser Leu Phe Ile Trp Thr His Ser Gly Trp Glu Gly Cys Ser Val Gln Cys 805 810 810 Gly Gly Glu Arg Arg Thr Ile Val Ser Cys Thr Arg Ile Val Asn 820 825 830 Lys Thr Thr Leu Val Asn Asp Ser Asp Cys Pro Gln Ala Ser Arg 835 840 Pro Glu Pro Gln Val Arg Arg Cys Asn Leu His Pro Cys Gln Ser Arg 850 860 Trp Val Ala Gly Pro Trp Ser Pro Cys Ser Ala Thr Cys Glu Lys Gly 865 875 880 Phe Gln His Arg Glu Val Thr Cys Val Tyr Gln Leu Gln Asn Gly Thr 885 890 His Val Ala Thr Arg Pro Leu Tyr Cys Pro Gly Pro Arg Pro Ala Ala Val Gln Ser Cys Glu Gly Gln Asp Cys Leu Ser Ile Trp Glu Ala Ser

Glu Trp Ser Gln Cys Ser Ala Ser Cys Gly Lys Gly Val Trp Lys Arg

Thr Val Ala Cys Thr Asn Ser Gln Gly Lys Cys Asp Ala Ser Thr Arg

945 Val Ala Cys Thr Asn Ser Gln Gly Lys Cys Asp Ala Ser Thr Arg

960

Pro Arg Ala Glu Glu Ala Cys Glu Asp Tyr Ser Gly Cys Tyr Glu Trp

Lys Thr Gly Asp Trp Ser Thr Cys Ser Ser Thr Cys Gly Lys Gly Leu

Gln Ser Arg Val Val Gln Cys Met His Lys Val Thr Gly Arg His Gly

Ser Glu Cys Pro Ala Leu Ser Lys Pro Ala Pro Tyr Arg Gln Cys

Tyr Gln Glu Val Cys Asn Asp Arg Ile Asn Ala Asn Thr Ile Thr

Ser Pro Arg Leu Ala Ala Leu Thr Tyr Lys Cys Thr Arg Asp Gln

Trp Thr Val Tyr Cys Arg Val Ile Arg Glu Lys Asn Leu Cys Gln

Asp Met Arg Trp Tyr Gln Arg Cys Cys Gln Thr Cys Arg Asp Phe

Tyr Ala Asn Lys Met Arg Gln Pro Pro Pro Ser Ser 1095

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<sup>&</sup>lt;210> 7 <211> 4342

<sup>&</sup>lt;213> Homo sapiens

<sup>&</sup>lt;220> <221> CDS <222> (148)..(3582)

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gct Ala	CCC Pro	ctc Leu	gcc Ala	gtg Val 110	gct Ala	gga Gly	att Ile	gga Gly	ctg Leu 115	gtg Val	ttg Leu	gac Asp	atc Ile	atc Ile 120	ctc Leu	510
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agc ser	ctc Leu	agc ser	CCC Pro	atc Ile 190	gtg Val	atc Ile	atc Ile	tcc Ser	gtg Val 195	gtc Val	tcc Ser	tgt Cys	gtg Val	gtg Val 200		750
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										ctt Leu 340						1182
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cgg Arg	att Ile	aag Lys	atc Ile 365	ctg Leu	ggc GTy	gac Asp	tgc Cys	tac Tyr 370	tac Tyr	tgc Cys	atc Ile	tgc Cys	ggc Gly 375	ttg Leu		1278
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atg	gtg	gag	gcc	atc	tcg	tat	gtg	cgg	gag	aag	acc	aag	act	<b>ggg</b>	gtg	1374

Met"	"Va7" 395	Glü	Ala	Ile	Ser	Туг 400	val	Arg	Glu	Lys	Thr 405	Lys	Thr	Gly	۷a۱	•	
gac Asp 410	atg Met	cgt Arg	gtg Val	ggg GTy	gtg Val 415	cac His	acg Thr	ggc Gly	acc Thr	gtg Val 420	ctg Leu	999 GTy	ggc Gly	gtc val	ctg Leu 425		1422
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<sup>&</sup>lt;213> Homo sapiens

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446 Glu Ser Pro Phe Phe Arg Ser Leu Asp Trp Gln Met Val Phe Leu Gln Arg Tyr Pro Pro Pro Leu Ile Pro Pro Arg Gly Glu Val Asn Ala Ala 465 470 480

Asp Ala Phe Asp Ile Gly Ser Phe Asp Glu Glu Asp Thr Lys Gly Ile 485 490 Lys Leu Leu Asp Ser Asp Gln Glu Leu Tyr Arg Asn Phe Pro Leu Thr 500 505 Ile Ser Glu Arg Trp Gln Gln Glu Val Ala Glu Thr Val Phe Asp Thr 515 525 Ile Asn Ala Glu Thr Asp Arg Leu Glu Ala Arg Lys Lys Ala Lys Asn 530 540 Lys Gln Leu Gly His Glu Glu Asp Tyr Ala Leu Gly Lys Asp Cys Ile 545 550 560 Met His Gly Tyr Met Ser Lys Met Gly Asn Pro Phe Leu Thr Gln Trp Gln Arg Arg Tyr Phe Tyr Leu Phe Pro Asn Arg Leu Glu Trp Arg Gly 580 585 Glu Gly Glu Ala Pro Gln Ser Leu Leu Thr Met Glu Glu Ile Gln Ser Val Glu Glu Thr Gln Ile Lys Glu Arg Lys Cys Leu Leu Leu Lys Ile 610 610Arg Gly Gly Lys Gln Phe Ile Leu Gln Cys Asp Ser Asp Pro Glu Leu 625 635 640 Val Gln Trp Lys Lys Glu Leu Arg Asp Ala Tyr Arg Glu Ala Gln Gln 655 Leu Val Gln Arg Val Pro Lys Met Lys Asn Lys Pro Arg Ser Pro Val Val Glu Leu Ser Lys Val Pro Leu Val Gln Arg Gly Ser Ala Asn Gly 675

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WO 2004/076682 PCT/US200	04/006020
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att tgt gcc ttg gtt ttc ctg gca gcc ctg ttc ctt atc tgc tat agc Ile Cys Ala Leu Val Phe Leu Ala Ala Leu Phe Leu Ile Cys Tyr Ser 45 50 55 60	434
tgc tgg gga aac tgt ttc ctg tac cac tgc tcc gat tcc ccg ctt cca Cys Trp Gly Asn Cys Phe Leu Tyr His Cys Ser Asp Ser Pro Leu Pro 65 70 75	482
gaa tcg gcg cat gat ccc ggc gtt gtg ggc acc taa cggcctgccc Glu Ser Ala His Asp Pro Gly Val Val Gly Thr 80 85	528
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<sup>12</sup> 87 PRT Homo sapiens

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Asp Pro Gly Val Val Gly Thr

13 3578 DNA Homo sapiens

CDS (268)..(1389)

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3578

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<sup>14</sup> 373 PRT

Homo sapiens

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90
95
100 343 ttt gcc tgt gag gtg gcc gcc ggc atc tgg ggc ttt gtc aac aag gac Phe Ala Cys Glu Val Ala Ala Gly Ile Trp Gly Phe Val Asn Lys Asp 105 110 391 cag atc gcc aag gat gtg aag cag ttc tat gac cag gcc cta cag cag Gln Ile Ala Lys Asp Val Lys Gln Phe Tyr Asp Gln Ala Leu Gln Gln 120 125 439 gcc gtg gtg gat gat gac gcc aac aac gcc aag gct gtg gtg aag acc Ala Val Val Asp Asp Asp Ala Asn Asn Ala Lys Ala Val Val Lys Thr 135 140 487 ttc cac gag acg ctt gac tgc tgt ggc tcc agc aca ctg act gct ttg Phe His Glu Thr Leu Asp Cys Cys Gly Ser Ser Thr Leu Thr Ala Leu 150 160 165 535 acc acc tca gtg ctc aag aac aat ttg tgt ccc tcg ggc agc aac atc Thr Thr Ser Val Leu Lys Asn Asn Leu Cys Pro Ser Gly Ser Asn Ile 170 180 583 atc agc aac ctc ttc aag gag gac tgc cac cag aag atc gat gac ctc Ile Ser Asn Leu Phe Lys Glu Asp Cys His Gln Lys Ile Asp Asp Leu 185 190 195 631 ttc tcc ggg aag ctg tac ctc atc ggc att gct gcc atc gtg gtc gct Phe Ser Gly Lys Leu Tyr Leu Ile Gly Ile Ala Ala Ile Val Val Ala 200 210 679 gtg atc atg atc ttc gag atg atc ctg agc atg gtg ctg tgc tgt ggc val Ile Met Ile Phe Glu Met Ile Leu Ser Met Val Leu Cys Cys Gly 220 727 atc cgg aac agc tcc gtg tac tga ggccccgcag ctctggccac agggacctct Ile Arg Asn Ser Ser Val Tyr 230 781 jcagtgcccc ctaagtgacc cggacacttc cgagggggcc atcaccgcct gtgtatataa 841 egtttccggt attactctgc tacacgtagc ctttttactt, ttggggtttt gtttttgttc 901 tgaactttcc tgttaccttt tcagggctga cgtcacatgt aggtggcgtg tatgagtgga 961 jacgggcctg ggtcttgggg actggagggc aggggtcctt ctgccctggg gtcccagggt 1021 active ctcagccagg cctctcctgg gagccactcg cccagagact cagcttggcc 1081

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16 236 PRT Homo sapiens

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<213> Homo sapiens

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85
90
95 Leu Val Ile Phe Ala Ile Glu Ile Ala Ala Ile Trp Gly Tyr Ser 100 105His Lys Asp Glu Val Ile Lys Glu Val Gln Glu Phe Tyr Lys Asp Thr 115 120 125 Tyr Asn Lys Leu Lys Thr Lys Asp Glu Pro Gln Arg Glu Thr Leu Lys 130 140 Ala Ile His Tyr Ala Leu Asn Cys Cys Gly Leu Ala Gly Gly Val Glu 145 150 160 Gln Phe Ile Ser Asp Ile Cys Pro Lys Lys Asp Val Leu Glu Thr Phe Thr Val Lys Ser Cys Pro Asp Ala Ile Lys Glu Val Phe Asp Asn Lys Phe His Ile Ile Gly Ala Val Gly Ile Gly Ile Ala Val Val Met Ile Phe Gly Met Ile Phe Ser Met Ile Leu Cys Cys Ala Ile Arg Arg Asn 210 215

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<213> Homo sapiens

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<212> PRT <213> Homo sapiens

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va1

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Homo sapiens

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gac Asp	ccc Pro 510	ggc GTy	agg Arg	cct Pro	gga Gly	ttc Phe 515	agc Ser	tac Tyr	cca Pro	gga Gly	ccc Pro 520	cga Arg	gga Gly	gca Ala	ccc Pro	1645
gga Gly 525	gaa Glu	aaa Lys	ggc GTy	gag Glu	ccc Pro 530	ggc GTy	cca Pro	cgc Arg	ggc GTy	ccc Pro 535	gag Glu	gga Gly	ggc Gly	cga Arg	ggc Gly 540	1693
gac Asp	ttt Phe	ggc GTy	ttg Leu	aaa Lys 545	gga Gly	gaa Glu	cct Pro	ggg GTy	agg Arg 550	aaa Lys	gga Gly		aaa Lys	gga GTy 555	gag Glu	1741

2850 2910

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<sup>&</sup>lt;213> PKI <213> Homo sapiens

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Lys Gly Glu Pro Gly Arg Lys Gly Glu Lys Gly Glu Pro Ala Asp Pro Gly Pro Pro Gly Glu Pro Gly Pro Arg Gly Pro Arg Gly Val Pro Gly 575 Pro Glu Gly Pro Gly Pro Pro Gly Asp Pro Gly Leu Thr Glu Cys 580 585 Asp Val Met Thr Tyr Val Arg Glu Thr Cys Gly Cys Cys Asp Cys Glu 595 Lys Arg Cys Gly Ala Leu Asp Val Val Phe Val Ile Asp Ser Ser Glu Ser Ile Gly Tyr Thr Asn Phe Thr Leu Glu Lys Asn Phe Val Ile Asn 625 635 Val Val Asn Arg Leu Gly Ala Ile Ala Lys Asp Pro Lys Ser Glu Thr Gly Thr Arg Val Gly Val Val Gln Tyr Ser His Glu Gly Thr Phe Glu 660 665 Ala Ile Gln Leu Asp Asp Glu His Ile Asp Ser Leu Ser Ser Phe Lys 675Glu Ala Val Lys Asn Leu Glu Trp Ile Ala Gly Gly Thr Trp Thr Pro Ser Ala Leu Lys Phe Ala Tyr Asp Arg Leu Ile Lys Glu Ser Arg Arg 705 Gln Lys Thr Arg Val Phe Ala Val Val Ile Thr Asp Gly Arg His Asp 725 730 Pro Arg Asp Asp Leu Asn Leu Arg Ala Leu Cys Asp Arg Asp Val Thr Val Thr Ala Ile Gly Ile Gly Asp Met Phe His Glu Lys His Glu 755 Ser Glu Asn Leu Tyr Ser Ile Ala Cys Asp Lys Pro Gln Gln Val Arg 770 780 Asn Met Thr Leu Phe Ser Asp Leu Val Ala Glu Lys Phe Ile Asp Asp 785 Met Glu Asp Val Leu Cys Pro Asp Pro Gln Ile Val Cys Pro Asp Leu 805 810 815 Pro Cys Gln Thr Gly Leu Asp Gly Ala Val Leu Cys

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Tyr Asn Arg Gly Cys Thr Asp Ile Ile Cys Cys Val Phe Leu Leu Leu
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Trp Leu Ala Phe Met Ile Ile Leu Ser Ile Leu Glu Val Ile Ile Ile
315 320 325 1071 ttg ctg ctc atc ttt ctc cgg aag aga att ctc atc gcg att gca ctc Leu Leu Leu Ile Phe Leu Arg Lys Arg Ile Leu Ile Ala Ile Ala Leu 335 340 345 1119 atc aaa gaa gcc agc agg gct gtg gga tac gtc atg tgc tcc ttg ctc Ile Lys Glu Ala Ser Arg Ala Val Gly Tyr Val Met Cys Ser Leu Leu 350 355 360 1167 tac cca ctg gtc acc ttc ttc ttg ctg tgc ctc tgc atc gcc tac tgg
Tyr Pro Leu Val Thr Phe Phe Leu Leu Cys Leu Cys Ile Ala Tyr Trp
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Thr Asn Phe Cys Thr Ser Ala Arg Asn Ala Phe Phe Leu Leu Met Arg
575 580 585 1839 aac atc atc aga gtg gct gtc ctg gat aaa gtt act gac ttc ctc ttc Asn Ile Ile Arg Val Ala Val Leu Asp Lys Val Thr Asp Phe Leu Phe 590 600 1887 ctg ttg ggc aaa ctt ctg atc gtt ggt agt gtg ggg atc ctg gct ttc Leu Leu Gly Lys Leu Leu Ile Val Gly Ser Val Gly Ile Leu Ala Phe 605 610 1935

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Lys Tyr Asp Pro Thr Phe Lys Gly Pro Ile Tyr Asn Arg Gly Cys Thr  $20 \ \ \,$ 

<sup>26</sup> 706 PRT Homo sapiens

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Glu Asp Leu Glu Arg Asn Asp Gly Ser Ala Glu Arg Pro Tyr Phe Met 675 680

Ser Ser Thr Leu Lys Lys Pro Leu Asn Lys Thr Asn Lys Lys Ala Ala 690 700

Glu Ser 705

27 2409 DNA Homo sapiens

CDS (1)..(2313)

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Ğ]n 225	٧a٦	va1	Thr	Аla	His 230	Tyr	Arg	Thr	Ser	Pro 235	Phe	Phe	Ser	val	Tyr 240	
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Homo sapiens

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264

His His Asp Tyr Tyr Ile Thr Ser Thr Ser Asn Gly Ser Leu Glu Gly 130 Leu Glu Asn Arg Glu Gly Gly Val Cys Arg Thr Arg Thr Met Lys Ile 145 150 160 Ile Met Lys Val Gly Gln Asp Pro Asn Ala Val Thr Pro Glu Gln Leu 165 170 175 Thr Thr Ser Arg Pro Ser Lys Glu Ala Asp Asn Thr Val Lys Met Ala 180 180 190 Thr Gln Ala Pro Gly Ser Arg Gly Ser Leu Gly Asp Ser Asp Gly Lys His Glu Thr Val Asn Gln Glu Glu Lys Ser Gly Pro Gly Ala Ser Gly 210 220 Gly Ser Ser Gly Asp Pro Asp Gly Phe Phe Asn Ser Lys Val Ala Leu 225 230 236 Phe Ala Ala Val Gly Ala Gly Cys Val Ile Phe Leu Leu Ile Ile Ile 255 Phe Leu Thr Val Leu Leu Leu Lys Leu Arg Lys Arg His Arg Lys His 260 270 Thr Gln Gln Arg Ala Ala Ala Leu Ser Leu Ser Thr Leu Ala Ser Pro 280 285 Lys Gly Gly Ser Gly Thr Ala Gly Thr Glu Pro Ser Asp Ile Ile Ile 290 295 Pro Leu Arg Thr Thr Glu Asn Asn Tyr Cys Pro His Tyr Glu Lys Val 305 310 315 Ser Gly Asp Tyr Gly His Pro Val Tyr Ile Val Gln Glu Met Pro Pro Gln Ser Pro Ala Asn Ile Tyr Tyr Lys Val

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216

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<sup>32</sup> 379 PRT Homo sapiens

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<sup>34</sup> 547 PRT Homo sapiens

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Phe Leu Met Arg Thr Asn Thr

37 5077 DNA Homo sapiens

CDS (405)..(4121)

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848

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cga Arg 165	gtg Val	tcg Ser	cat His	gcc Ala	ggc Gly 170	atg Met	atc Ile	aac Asn	ccg Pro	gag Glu 175	gac Asp	cgc Arg	tgg Trp	aag Lys	agc Ser 180	944
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gac Asp 405	ggc Gly	ttt Phe	gag Glu	tgc Cys	atc Ile 410	tgc Cys	ccc Pro	gag Glu	cag Gln	tgg Trp 415	gtg Val	ggg GTy	gcc Ala	acc Thr	tgc Cys 420	1664
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WO 2004/076682 PCT/US2004/00	06020
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aac ccc atc cgc aac ccc atc gag cgg ccg ggg ggc cac aag gac Asn Pro Ile Arg Asn Pro Ile Glu Arg Pro Gly Gly His Lys Asp 1135 1140 1145	3842
gtg ctc tac cag tgc aag aac ttc acg ccg ccg cgc agg gcg Val Leu Tyr Gln Cys Lys Asn Phe Thr Pro Pro Pro Arg Ala 1150 1155 1160	3887
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<sup>38</sup> 1238 PRT Homo sapiens

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Leu Gly Gln Pro Cys Arg Asn Gly Gly Thr Cys Ile Asp Glu Val Asp 645 655 Ala Phe Arg Cys Phe Cys Pro Ser Gly Trp Glu Gly Glu Leu Cys Asp 660 670 . Thr Asn Pro Asn Asp Cys Leu Pro Asp Pro Cys His Ser Arg Gly Arg 675 685 Cys Tyr Asp Leu Val Asn Asp Phe Tyr Cys Ala Cys Asp Asp Gly Trp 690 700 Lys Gly Lys Thr Cys His Ser Arg Glu Phe Gln Cys Asp Ala Tyr Thr 705 710 720 Cys Ser Asn Gly Gly Thr Cys Tyr Asp Ser Gly Asp Thr Phe Arg Cys 735 Ala Cys Pro Pro Gly Trp Lys Gly Ser Thr Cys Ala Val Ala Lys Asn 740 750 Ser Ser Cys Leu Pro Asn Pro Cys Val Asn Gly Gly Thr Cys Val Gly 755 Ser Gly Ala Ser Phe Ser Cys Ile Cys Arg Asp Gly Trp Glu Gly Arg 770 780 Thr Cys Thr His Asn Thr Asn Asp Cys Asn Pro Leu Pro Cys Tyr Asn 785 Gly Gly Ile Cys Val Asp Gly Val Asn Trp Phe Arg Cys Glu Cys Ala Pro Gly Phe Ala Gly Pro Asp Cys Arg Ile Asn Ile Asp Glu Cys Gln 820 825 Ser Ser Pro Cys Ala Tyr Gly Ala Thr Cys Val Asp Glu Ile Asn Gly 835 Tyr Arg Cys Ser Cys Pro Pro Gly Arg Ala Gly Pro Arg Cys Gln Glu 850 860 Val Ile Gly Phe Gly Arg Ser Cys Trp Ser Arg Gly Thr Pro Phe Pro 865 His Gly Ser Ser Trp Val Glu Asp Cys Asn Ser Cys Arg Cys Leu Asp Gly Arg Arg Asp Cys Ser Lys Val Trp Cys Gly Trp Lys Pro Cys Leu 900 910 Leu Ala Gly Gln Pro Glu Ala Leu Ser Ala Gln Cys Pro Leu Gly Gln 915 Arg Cys Leu Glu Lys Ala Pro Gly Gln Cys Leu Arg Pro Pro Cys Glu 930 940 Ala Trp Gly Glu Cys Gly Ala Glu Glu Pro Pro Ser Thr Pro Cys Leu 950 955 960

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Pro Arg Ser Gly His Leu Asp Asn Asn Cys Ala Arg Leu Thr Leu His Phe Asn Arg Asp His Val Pro Gln Gly Thr Thr Val Gly Ala Ile Cys Ser Gly Ile Arg Ser Leu Pro Ala Thr Arg Ala Val Ala Arg Asp Arg Leu Leu Val Leu Leu Cys Asp Arg Ala Ser Ser Gly Ala Ser Ala 1010 1015 Val Glu Val Ala Val Ser Phe Ser Pro Ala Arg Asp Leu Pro Asp 1025 1030 1035 Ser Ser Leu Ile Gln Gly Ala Ala His Ala Ile Val Ala Ala Ile 1040 1045 Thr Gln Arg Gly Asn Ser Ser Leu Leu Leu Ala Val Thr Glu Val Lys Val Glu Thr Val Val Thr Gly Gly Ser Ser Thr Gly Leu Leu Val Pro Val Leu Cys Gly Ala Phe Ser Val Leu Trp Leu Ala Cys val Val Leu Cys Val Trp Trp Thr Arg Lys Arg Arg Lys Glu Arg Glu Arg Ser Arg Leu Pro Arg Glu Glu Ser Ala Asn Asn Gln Trp Ala Pro Leu Asn Pro Ile Arg Asn Pro Ile Glu Arg Pro Gly Gly His Lys Asp Val Leu Tyr Gln Cys Lys Asn Phe Thr Pro Pro Pro 1145 Arg Arg Ala Asp Glu Ala Leu Pro Gly Pro Ala Gly His Ala Ala 1160 1165 Val Arg Glu Asp Glu Glu Asp Glu Asp Leu Gly Arg Gly Glu Glu Asp Ser Leu Glu Ala Glu Lys Phe Leu Ser His Lys Phe Thr Lys Asp Pro Gly Arg Ser Pro Gly Arg Pro Ala His Trp Ala Ser Gly 1210 1215 Pro Lys Val Asp Asn Arg Ala Val Arg Ser Ile Asn Glu Ala Arg Tyr Ala Gly Lys Glu 1235

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<sup>&</sup>lt;212> UNA <213> Homo sapiens

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gga Gly	cgg Arg	gcc Ala	cgg Arg 35	ctc Leu	ttc Phe	gac Asp	gtg Val	cgc Arg 40	tct Ser	cgc Arg	gag Glu	gag Glu	gcg ATa 45	gca Ala	gct Ala	143
ggg Gly	acc Thr	atc Ile 50	cca Pro	ggg GTy	gcg Ala	ctc Leu	aac Asn 55	atc Ile	ccg Pro	gtg Val	tcc Ser	gag Glu 60	ttg Leu	gag Glu	agt Ser	191
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ggg Gly	tac Tyr	ggg Gly	gag Glu 115	gtg Val	tgg Trp	ctg Leu	cta Leu	gct Ala 120	ggg Gly	agg Arg	tga	tggg	ggact	gc		381
ctġt	cati	tcc t	gtca	ıgtct	c to	acgo	ttct	ttg	tcto	cac	aggg	ctc	gca a	ectac	gctgg	441
agco	tata	aga g	gaatg	gttg	g ag	jaaag	jagag	tta	ggca	ıgga	ggca	igctt	ac t	gatt	gccac	501
cccc	tgg	ccc c	ttaa	tggc	c ac	ctta	ıacta	agg	gtgt	gaa	cggg	ıctga	ict t	ggtg	aattg	561
ggca	acto	ct t	atag	ıtgti	g tg	caca	caaa	ago	atca	aat	aaag	aaca	itt t	aato	aaaaa	621
aaaa	aaaa	aaa a	iaa													634
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PRT Homo sapiens

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Arg Gly Leu Gln Ala Thr Gln Leu Ala Arg Ser Leu Gly Tyr Thr Gly 100 105

Tyr Gly Glu Val Trp Leu Leu Ala Gly Arg 115 120

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aad Ly 22	g tgo S Cys	c tt s Ph	c ca e Hi:	c aag s Ly	g cto S Lei 230	g gco i Ala )	tct Ser	gç	c tat a Tyr	ggg GT 23	g gco / Ala	c ago a Aro	ca Gli	g ct 1 Le	g cag u Gln 240	899
gg( GT)	tao Tyr	tg Cy:	c gca s Ala	a age a Sei 24:	cto Leu	ttt Phe	gcc	ate Ile	ctc Leu 250	cto Leu	cco Pro	cac o Glr	gaq Asp	C CC Pro 25	tcg Ser	947
tto Phe	caç Gli	ate Me	g cco E Pro 260	cto Lei	g gad I Asp	ctg Leu	tat Tyr	gco Ala 265	tat Tyr	gca Ala	gto Val	gco I Ala	aca Thr 270	gg Gl	g gac / Asp	995
gço Ala	cto Lei	Lei 27:	4 711	aag Lys	cto Leu	tgc Cys	cta Leu 280	GH	ttc Phe	ctg Leu	gço Ala	tgg Trp 285	AST	tto Pho	gag Glu	1043
gcc Ala	ttg Leu 290	aco Thi	cac Glr	g gco i Ala	gag Glu	gcc Ala 295	tgg Trp	CCC Pro	agt Ser	gtc Val	Pro 300	aca Thr	ga o Asp	cto Lei	ctc Leu	1091
caa Gln 305	ctg Leu	cto Lei	ctg Leu	CCC Pro	agg Arg 310	agc Ser	gac Asp	ctg Leu	gcg Ala	gtg Val 315	ccc Pro	agc Ser	gag Glu	cto Lei	gcc Ala 320	1139
cta Leu	ctg Leu	Lys	gcc Ala	gtg Val 325	gac Asp	acc Thr	tgg Trp	agc Ser	tgg Trp 330	ggg Gly	gag Glu	cgt Arg	gcc Ala	tco Ser 335	cat	1187
	gag Glu		340					343					350		ctc	1235
cct Pro	gag Glu	gag Glu 355	ctc Leu	ttt Phe	gag Glu	ctg Leu	cag Gln 360	ttc Phe	aac Asn	ctg Leu	tcc Ser	ctg Leu 365	tac Tyr	tgg Trp	agc Ser	1283
cac His	gag Glu 370	gcc Ala	ctg Leu	ttc Phe	cag Gln	aag Lys 375	aag Lys	act Thr	ctg Leu	cag Gln	gcc Ala 380	ctg Leu	gaa Glu	ttc Phe	cac His	1331
act Thr 385	gtg Val	CCC Pro	ttc Phe	cag Gln	ttg Leu 390	ctg Leu	gcc Ala	cgg Arg	tac Tyr	aaa Lys 395	ggc Gly	ctg Leu	aac Asn	ctc Leu	acc Thr 400	1379
gag Glu	gat Asp	acc Thr	tac Tyr	aag Lys 405	ccc Pro	cgg Arg	att Ile	tac Tyr	acc Thr 410	tcg Ser	ccc Pro	acc Thr	tgg Trp	agt Ser 415	gcc Ala	1427
Phe	gtg Val	aca Thr	gac Asp 420	agt Ser	tcc Ser	tgg Trp	agt Ser	gca Ala 425	cgg Arg	aag Lys	tca Ser	caa Gln	ctg Leu 430	gtc Val	Tyr	1475
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gcc Ala	CCC Pro 450	tct Ser	gac Asp	tac Tyr	aga Arg	tac Tyr 455	tac Tyr	ccc Pro	tac Tyr	cag Gln	tcc Ser 460	ttc Phe	cag Gln	act Thr	cca Pro	1571
465	1113	710	361	FIIC	470	riie	GIII	ASP	Lys	475	vaı	tcc Ser	Trp	Ser	Leu 480	1619
				103					450			ggc Gly		495		1667
tcc Ser	tcg Ser	gac Asp	gag Glu 500	ctc Leu	cct Pro	gtc Val	ctg Leu	ggc Gly 505	ctc Leu	acc Thr	aag Lys	tct Ser	ggc Gly 510	ggc Gly	tca Ser	1715
gat Asp	cgc Arg	acc Thr 515	att Ile	gcc Ala	tac Tyr	Giu i	aac Asn 520	aaa Lys	gcc Ala	ctg Leu	atg Met	ctc Leu 525	tgc Cys	gaa Glu	ggg GTy	1763
ctc Leu	ttc Phe 530	gtg Val	gca Ala	gac Asp	* W I	acc Thr 535	gat i	ttc Phe	gag Glu	o i y	tgg Trp 540	aag Lys	gct Ala	gcg Ala	att Ile	1811

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ccc agt gcc Pro Ser Ala 545	ctg gac acc aac Leu Asp Thr Asn 550	agc tcg aag Ser Ser Lys	agc acc tcc tcc ttc ccc Ser Thr Ser Ser Phe Pro 555	1859
tgc ccg gca Cys Pro Ala	ggg cac ttc aac Gly His Phe Asn 565	ggc ttc cgc Gly Phe Arg 570	acg gtc atc cgc ccc ttc Thr Val Ile Arg Pro Phe 575	1907
tac ctg acc Tyr Leu Thr	aac tcc tca ggt Asn Ser Ser Gly 580	gtg gac tag Val Asp 585	acgcgtggcc aagggtggtg	1957
agaaccggag a	aaccccagga cgccc	tcact gcaggct	ccc ctcctcggct tccttcctc	t 2017
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gtttccaggt g	ggtaagcacc aggag	gccct cgaggtt	gct ctggatcccc ccacagccc	c 2197
tggtcagtct g	gcccttgtca ctggt	ctgag gtcatta	aaa ttacattgag gttccta	2254

Met Thr Pro Pro Arg Leu Phe Trp Val Trp Leu Leu Val Ala Gly Thr 10 15 Gln Gly Val Asn Asp Gly Asp Met Arg Leu Ala Asp Gly Gly Ala Thr Asn Gln Gly Arg Val Glu Ile Phe Tyr Arg Gly Gln Trp Gly Thr Val 45 Cys Asp Asn Leu Trp Asp Leu Thr Asp Ala Ser Val Val Cys Arg Ala 50Leu Gly Phe Glu Asn Ala Thr Gln Ala Leu Gly Arg Ala Ala Phe Gly Gln Gly Ser Gly Pro Ile Met Leu Asp Glu Val Gln Cys Thr Gly Thr Cys Arg His Glu Arg Asp Ala Gly Val Val Cys Thr Asp Glu Thr Arg Ser Thr His Thr Leu Asp Leu Ser Arg Glu Leu Ser Glu Ala Leu Gly 130 140 Gln Ile Phe Asp Ser Gln Arg Gly Cys Asp Leu Ser Ile Ser Val Asn 150 155 160 Val Gln Gly Glu Asp Ala Leu Gly Phe Cys Gly His Thr Val Ile Leu 165 170 175 Thr Ala Asn Leu Glu Ala Gln Ala Leu Trp Lys Glu Pro Gly Ser Asn 180 Val Thr Met Ser Val Asp Ala Glu Cys Val Pro Met Val Arg Asp Leu 195 200

<sup>42</sup> 585 PRT Homo sapiens

Leu Arg Tyr Phe Tyr Ser Arg Arg Ile Asp Ile Thr Leu Ser Ser Val Lys Cys Phe His Lys Leu Ala Ser Ala Tyr Gly Ala Arg Gln Leu Gln 225 235 Gly Tyr Cys Ala Ser Leu Phe Ala Ile Leu Leu Pro Gln Asp Pro Ser 245 250 255 Phe Gln Met Pro Leu Asp Leu Tyr Ala Tyr Ala Val Ala Thr Gly Asp Ala Leu Leu Glu Lys Leu Cys Leu Gln Phe Leu Ala Trp Asn Phe Glu 275 . 280 . 285 Ala Leu Thr Gln Ala Glu Ala Trp Pro Ser Val Pro Thr Asp Leu Leu 290 300 Gln Leu Leu Pro Arg Ser Asp Leu Ala Val Pro Ser Glu Leu Ala 305 310 320 Leu Leu Lys Ala Val Asp Thr Trp Ser Trp Gly Glu Arg Ala Ser His Glu Glu Val Glu Gly Leu Val Glu Lys Ile Arg Phe Pro Met Met Leu 340 Pro Glu Glu Leu Phe Glu Leu Gln Phe Asn Leu Ser Leu Tyr Trp Ser 365 His Glu Ala Leu Phe Gln Lys Lys Thr Leu Gln Ala Leu Glu Phe His 370 380 Thr Val Pro Phe Gln Leu Leu Ala Arg Tyr Lys Gly Leu Asn Leu Thr 385 395 400 Glu Asp Thr Tyr Lys Pro Arg Ile Tyr Thr Ser Pro Thr Trp Ser Ala 405 410 415 Phe Val Thr Asp Ser Ser Trp Ser Ala Arg Lys Ser Gln Leu Val Tyr 420 430 Gln Ser Arg Arg Gly Pro Leu Val Lys Tyr Ser Ser Asp Tyr Phe Gln Ala Pro Ser Asp Tyr Arg Tyr Tyr Pro Tyr Gln Ser Phe Gln Thr Pro
450
450 Gln His Pro Ser Phe Leu Phe Gln Asp Lys Arg Val Ser Trp Ser Leu 465 470 475 Val Tyr Leu Pro Thr Ile Gln Ser Cys Trp Asn Tyr Gly Phe Ser Cys Ser Ser Asp Glu Leu Pro Val Leu Gly Leu Thr Lys Ser Gly Gly Ser Asp Arg Thr Ile Ala Tyr Glu Asn Lys Ala Leu Met Leu Cys Glu Gly 515

Leu Phe Val Ala Asp Val Thr Asp Phe Glu Gly Trp Lys Ala Ala Ile
Pro Ser Ala Leu Asp Thr Asn Ser Ser Lys Ser Thr Ser Ser Phe Pro
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Cys Pro Ala Gly His Phe Asn Gly Phe Arg Thr Val Ile Arg Pro Phe
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Met 165	Аla	Leu	Glu	Tyr	Val 170	Ala	Cys	Asp	Leu	His 175	Arg	Leu	Glu	Glu	Leu 180	•
gtg Val	ctc Leu	gac Asp	agg Arg	tgt Cys 185	gta Val	cgc Arg	atc Ile	acg Thr	gac Asp 190		ggc GTy	ctc Leu	agc Ser	tat Tyr 195	ctg Leu	870
tcc Ser	acc Thr	atg Met	tcg Ser 200	tcc Ser	ctc Leu	cgc Arg	agc Ser	ctc Leu 205	tac Tyr	ctg Leu	cga Arg	tgg Trp	tgc Cys 210	tgc Cys	cag Gln	918
gtg Va I	caa Gln	gac Asp 215	ttc Phe	ggg Gly	ctg Leu	aag Lys	cac His 220	ctc Leu	ctg Leu	gcc Ala	ctg Leu	ggg G I y 225		ttg Leu		966
ctc Leu	ctg Leu 230	tct Ser	ctg Leu	gca Ala	ggc Gly	tgc Cys 235	ccg Pro	ctg Leu	ctc Leu	acc Thr	acc Thr 240	acc Thr	ggg Gly	ctg Leu	tcg Ser	1014
ggc GTy 245	ctg Leu	gtg Val	cag Gln	Ctg Leu	cag Gln 250	gag Glu	ctg Leu	gag Glu	gag Glu	ctg Leu 255	gag Glu	ctg Leu	acc Thr	aac Asn	tgc Cys 260	1062
ccc Pro	ggg Gly	gcc Ala	1111	ccc Pro 265	gag Glu	ctc Leu	ttc Phe	aag Lys	tat Tyr 270	ttc Phe	tcg Ser	cag Gln	cac His	ctg Leu 275	ccc Pro	1110
cgc Arg	tgc Cys	ctc Leu	gtc Val 280	att Ile	gag Glu	tag	cgcg	aggc	cc c	cgcc	ccgg	rt cg	cggg	aacc	:	1161
cggc	catg	ac c	tggg	cggg	g gc	gc										1185

PCT/US2004/006020

<400> 44

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<sup>&</sup>lt;210> 44 <211> 282

<sup>&</sup>lt;213> Homo sapiens

Arg Ile Thr Asp Met Ala Leu Glu Tyr Yal Ala Cys Asp Leu His Arg Leu Glu Glu Leu Val Leu Asp Arg Cys Val Arg Ile Thr Asp Thr Gly Leu Ser Tyr Leu Ser Thr Met Ser Ser Leu Arg Ser Leu Tyr Leu Arg Trp Cys Cys Gln Val Gln Asp Phe Gly Leu Lys His Leu Leu Ala Leu Gly Ser Leu Arg Leu Leu Ala Leu Gly Ser Leu Arg Leu Leu Ala Cys Pro Leu Leu Thr Thr Gly Leu Ser Gly Leu Val Gln Leu Glu Glu Leu Glu Glu Leu Glu Glu Leu Thr Asp Cys Pro Gly Ala Thr Pro Glu Leu Phe Lys Tyr Phe Ser Gln His Leu Pro Arg Cys Leu Yal Ile Glu

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<210> 46 <211> 265 <212> PRT

<213> Homo sapiens

<400> 46

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Ile Ile Leu Val Ala Gly Ala Phe Phe Trp Leu Val Ser Leu Leu Leu 35 Ala Ser Val Val Trp Phe Ile Leu Val His Val Thr Asp Arg Ser Asp Ala Arg Leu Gln Tyr Gly Leu Leu Ile Phe Gly Ala Ala Val Ser Val Leu Leu Gln Glu Val Phe Arg Phe Ala Tyr Tyr Lys Leu Leu Lys Lys Ala Asp Glu Gly Leu Ala Ser Leu Ser Glu Asp Gly Arg Ser Pro Ile 100 100 Ser Ile Arg Gln Met Ala Tyr Val Ser Gly Leu Ser Phe Gly Ile Ile 115 125 Ser Gly Val Phe Ser Val Ile Asn Ile Leu Ala Asp Ala Leu Gly Pro 130 140 Gly Val Val Gly Ile His Gly Asp Ser Pro Tyr Tyr Phe Leu Thr Ser Ala Phe Leu Thr Ala Ala Ile Ile Leu Leu His Thr Phe Trp Gly Val 165 170 175 Val Phe Phe Asp Ala Cys Glu Arg Arg Arg Tyr Trp Ala Leu Gly Leu 180 180 Val Val Gly Ser His Leu Leu Thr Ser Gly Leu Thr Phe Leu Asn Pro 200 205 Trp Tyr Glu Ala Ser Leu Leu Pro Ile Tyr Ala Val Thr Val Ser Met 210 220 Gly Leu Trp Ala Phe Ile Thr Ala Gly Gly Ser Leu Arg Ser Ile Gln 230 235 Arg Ser Leu Leu Cys Arg Arg Gln Glu Asp Ser Arg Val Met Val Tyr Ser Ala Leu Arg Ile Pro Pro Glu Asp 260

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gag agc aaa gcc ctg ctc gtg ctg act ctg gcc gtg tgg ctc cag agt

<sup>47</sup> 3549 DNA Homo sapiens

CDS (175)..(1602)

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Ala Lys Arg Ser Ser Lys Met Tyr Leu Lys Thr Arg Ser Gln Met Pro	
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agt gaa acc cat acc aat cag gcc ttt gag att tct ctg tat ggc acc Ser Glu Thr His Thr Asn Gln Ala Phe Glu Ile Ser Leu Tyr Gly Thr 355 360 365	1281
gtg gcc gag agt gag aac atc cca ttc act ctg cct gaa gtt tcc aca Val Ala Glu Ser Glu Asn Ile Pro Phe Thr Leu Pro Glu Val Ser Thr 370 385	1329
aat aag acc tac tcc ttc cta att tac aca gag gta gat att gga gaa Asn Lys Thr Tyr Ser Phe Leu Ile Tyr Thr Glu Val Asp Ile Gly Glu 390 400	1377
cta ctc atg ttg aag ctc aaa tgg aag agt gat tca tac ttt agc tgg Leu Leu Met Leu Lys Leu Lys Trp Lys Ser Asp Ser Tyr Phe Ser Trp 405 410 415	1425
tca gac tgg tgg agc agt ccc ggc ttc gcc att cag aag atc aga gta Ser Asp Trp Trp Ser Ser Pro Gly Phe Ala Ile Gln Lys Ile Arg Val 425 430	1473
aaa gca gga gag act cag aaa aag gtg atc ttc tgt tct agg gag aaa Lys Ala Gly Glu Thr Gln Lys Lys Val Ile Phe Cys Ser Arg Glu Lys 435 440 445	1521
gtg tct cat ttg cag aaa gga aag gca cct gcg gta ttt gtg aaa tgc Val Ser His Leu Gln Lys Gly Lys Ala Pro Ala Val Phe Val Lys Cys 450 460 465	1569
cat gac aag tct ctg aat aag aag tca ggc tga aactgggcga atctacagaa His Asp Lys Ser Leu Asn Lys Lys Ser Gly 470 475	1622
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2942 cccgactgtg aaagtatgtg atatctgaac acatactaga aagctctgca tgtgtgttgt ccttcagcat aattcggaag ggaaaacagt cgatcaaggg atgtattgga acatgtcgga 3002 gtagaaattg ttcctgatgt gccagaactt cgaccctttc tctgagagag atgatcgtgc 3062 ctataaatag taggaccaat gttgtgatta acatcatcag gcttggaatg aattctctct 3122 aaaaataaaa tgatgtatga tttgttgttg gcatcccctt tattaattca ttaaatttct 3182 ggatttgggt tgtgacccag ggtgcattaa cttaaaagat tcactaaagc agcacatagc 3242 actgggaact ctggctccga aaaactttgt tatatatatc aaggatgttc tggctttaca 3302 3362 ttttatttat tagctgtaaa tacatgtgtg gatgtgtaaa tggagcttgt acatattgga 3422 aaggtcattg tggctatctg catttataaa tgtgtggtgc taactgtatg tgtctttatc agtgatggtc tcacagagcc aactcactct tatgaaatgg gctttaacaa aacaagaaag 3482 3542 aaacgtactt aactgtgtga agaaatggaa tcagctttta ataaaattga caacatttta 3549 ttaccac

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<400> 48

 Met
 Glu
 Ser
 Lys
 Åla
 Leu
 Leu
 Val
 Leu
 Thr
 Leu
 Ala
 Val
 Leu
 Ala
 th

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Glu Ala Pro Ser Arg Leu Ser Pro Asp Asp Ala Asp Phe Val Asp Val Leu His Thr Phe Thr Arg Gly Ser Pro Gly Arg Ser Ile Gly Ile Gln 210 220 Lys Pro Val Gly His Val Asp Ile Tyr Pro Asp Gly Gly Thr Phe Gln 225 235 240 Pro Gly Cys Asn Ile Gly Glu Ala Ile Arg Val Ile Ala Glu Arg Gly Leu Gly Asp Val Asp Gln Leu Val Lys Cys Ser His Glu Arg Ser Ile 260 270 His Leu Phe Ile Asp Ser Leu Leu Asn Glu Glu Asn Pro Ser Lys Ala 275 285 1 Tyr Arg Cys Ser Ser Lys Glu Ala Phe Glu Lys Gly Leu Cys Leu Ser Cys Arg Lys Asn Arg Cys Asn Asn Leu Gly Tyr Glu Ile Asn Lys Val Arg Ala Lys Arg Ser Ser Lys Met Tyr Leu Lys Thr Arg Ser Gln Met Pro Tyr Lys Val Phe His Tyr Gln Val Lys Ile His Phe Ser Gly Thr Glu Ser Glu Thr His Thr Asn Gln Ala Phe Glu Ile Ser Leu Tyr Gly Thr Val Ala Glu Ser Glu Asn Ile Pro Phe Thr Leu Pro Glu Val Ser Thr Asn Lys Thr Tyr Ser Phe Leu Ile Tyr Thr Glu Val Asp Ile Gly 385 395 400 Glu Leu Leu Met Leu Lys Leu Lys Trp Lys Ser Asp Ser Tyr Phe Ser 405 410 415 Trp Ser Asp Trp Trp Ser Ser Pro Gly Phe Ala Ile Gln Lys Ile Arg Val Lys Ala Gly Glu Thr Gln Lys Lys Val Ile Phe Cys Ser Arg Glu Lys Val Ser His Leu Gln Lys Gly Lys Ala Pro Ala Val Phe Val Lys Cys His Asp Lys Ser Leu Asn Lys Lys Ser Gly 465 475

<sup>49</sup> 5100 DNA Homo sapiens

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aag ctg gag tcc acc atc gtg gtc agc ggc ctg gag gat gcg gcc gca Lys Leu Glu Ser Thr Ile Val Val Ser Gly Leu Glu Asp Ala Ala Ala 55 60 65	49
gtg gac ttc cag ttt tcc aag gga gcc gtg tac tgg aca gac gtg agc val Asp Phe Gln Phe Ser Lys Gly Ala Val Tyr Trp Thr Asp Val Ser 70 75 80	97
gag gag gcc atc aag cag acc tac ctg aac cag acg ggg gcc gcc gtg Glu Glu Ala Ile Lys Gln Thr Tyr Leu Asn Gln Thr Gly Ala Ala Val 85 90 95	45
cag aac gtg gtc atc tcc ggc ctg gtc tct ccc gac ggc ctc gcc tgc Gln Asn Val Val Ile Ser Gly Leu Val Ser Pro Asp Gly Leu Ala Cys 100 115	93
ASP Trp val Gly Lys Leu Tyr Trp Thr Asp Ser Glu Thr Asn Arg 120 125 130	41
atc gag gtg gcc aac ctc aat ggc aca tcc cgg aag gtg ctc ttc tgg Ile Glu Val Ala Asn Leu Asn Gly Thr Ser Arg Lys Val Leu Phe Trp 135 140 48	89
cag gac ctt gac cag ccg agg gcc atc gcc ttg gac ccc gct cac ggg Gln Asp Leu Asp Gln Pro Arg Ala Ile Ala Leu Asp Pro Ala His Gly 150 155 160	37
Tŷr Met Tŷr Trp Thr Asp Trp Gly Glu Thr Pro Arg Ile Glu Arg Ala 165 170	85
ggg atg gat ggc agc acc cgg aag atc att gtg gac tcg gac att tac Gly Met Asp Gly Ser Thr Arg Lys Ile Ile Val Asp Ser Asp Ile Tyr 180 195	33
tgg ccc aat gga ctg acc atc gac ctg gag gag cag aag ctc tac tgg Trp Pro Asn Gly Leu Thr Ile Asp Leu Glu Glu Gln Lys Leu Tyr Trp 200 205 210	81
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Gln	Pro	Phe	Phe 295	His	Thr	Arg	Cys	Glu 300	Glu	Asp	Asn	Gly	G]y 305	Cys	Ser	
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ccc Pro	acg Thr 325	ggt Gly	gtg Val	cag Gln	ctg Leu	cag Gln 330	gac Asp	aac Asn	ggc GTy	agg Arg	acg Thr 335	tgt Cys	aag Lys	gca Ala	gga Gly	1065
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tgc	agc	cac	ctg	tgc	ttc	ttc	aca	ccc	cac	gca	acc	cgg	tgt	ggc	tgc	1929

Cys Ser His Leu Cys Phe Phe Thr Pro His Ala Thr Arg Cys Gly Cys 615 620 6251977 ccc atc ggc ctg gag ctg ctg agt gac atg aag acc tgc atc gtg cct Pro Ile Gly Leu Glu Leu Leu Ser Asp Met Lys Thr Cys Ile Val Pro 630 640 gag gcc ttc ttg gtc ttc acc agc aga gcc gcc atc cac agg atc tcc Glu Ala Phe Leu Val Phe Thr Ser Arg Ala Ala Ile His Arg Ile Ser 645 655 2025 ctc gag acc aat aac aac gac gtg gcc atc ccg ctc acg ggc gtc aag Leu Glu Thr Asn Asn Asn Asp Val Ala Ile Pro Leu Thr Gly Val Lys 660 675 2073 gag gcc tca gcc ctg gac ttt gat gtg tcc aac aac cac atc tac tgg Glu Ala Ser Ala Leu Asp Phe Asp Val Ser Asn Asn His Ile Tyr Trp 680 685 690 2121 2169 aca gac gtc agc ctg aag acc atc agc cgc gcc ttc atg aac ggg agc Thr Asp Val Ser Leu Lys Thr Ile Ser Arg Ala Phe Met Asn Gly Ser 695 700 705 2217 tcg gtg gag cac gtg gtg gag ttt ggc ctt gac tac ccc gag ggc atg Ser Val Glu His Val Val Glu Phe Gly Leu Asp Tyr Pro Glu Gly Met 710 720 2265 gcc gtt gac tgg atg ggc aag aac ctc tac tgg gcc gac act ggg acc Ala Val Asp Trp Met Gly Lys Asn Leu Tyr Trp Ala Asp Thr Gly Thr 725 730 735 2313 aac aga atc gaa gtg gcg cgg ctg gac ggg cag ttc cgg caa gtc ctc Asn Arg Ile Glu Val Ala Arg Leu Asp Gly Gln Phe Arg Gln Val Leu 740 755 2361 gtg tgg agg gac ttg gac aac ccg agg tcg ctg gcc ctg gat ccc acc Val Trp Arg Asp Leu Asp Asn Pro Arg Ser Leu Ala Leu Asp Pro Thr 760 765 770 aag ggc tac atc tac tgg acc gag tgg ggc ggc aag ccg agg atc gtg Lys Gly Tyr Ile Tyr Trp Thr Glu Trp Gly Gly Lys Pro Arg Ile Val 775 780 785 2409 cgg gcc ttc atg gac ggg acc aac tgc atg acg ctg gtg gac aag gtg Arg Ala Phe Met Asp Gly Thr Asn Cys Met Thr Leu Val Asp Lys Val 790 800 2457 2505 ggc cgg gcc aac gac ctc acc att gac tac gct gac cag cgc ctc tac Gly Arg Ala Asn Asp Leu Thr Ile Asp Tyr Ala Asp Gln Arg Leu Tyr 805 815 2553 tgg acc gac ctg gac acc aac atg atc gag tcg tcc aac atg ctg ggt
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820 830 835 2601 cag gag cgg gtc gtg att gcc gac gat ctc ccg cac ccg ttc ggt ctg Gln Glu Arg Val Val Ile Ala Asp Asp Leu Pro His Pro Phe Gly Leu 840 845 2649 acg cag tac agc gat tat atc tac tgg aca gac tgg aat ctg cac agc Thr Gln Tyr Ser Asp Tyr Ile Tyr Trp Thr Asp Trp Asn Leu His Ser 855 860 865 att gag cgg gcc gac aag act agc ggc cgg aac cgc acc ctc atc cag Ile Glu Arg Ala Asp Lys Thr Ser Gly Arg Asn Arg Thr Leu Ile Gln 870 875 2697 2745 ggc cac ctg gac ttc gtg atg gac atc ctg gtg ttc cac tcc tcc cgc Gly His Leu Asp Phe Val Met Asp Ile Leu Val Phe His Ser Ser Arg 885 2793 cag gat ggc ctc aat gac tgt atg cac aac aac ggg cag tgt ggg Cag Gln Asp Gly Leu Asn Asp Cys Met His Asn Asn Gly Gln Cys Gly Gln 900 910 915 ctg tgc ctt gcc atc ccc ggc ggc cac cgc tgc ggc tgc gcc tca cac Leu Cys Leu Ala Ile Pro Gly Gly His Arg Cys Gly Cys Ala Ser His 920 925 930 2841 2889 tac acc ctg gac ccc agc agc cgc aac tgc agc ccg ccc acc acc ttc

Tyr	Thr	Leu	Asp 935	Pro	Ser	Ser	Arg	Asn 940	Cys	Ser	Pr	^O P	ro i	Thr 945	Tŀ	ır	Phe	
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cag Gln	cac His 965	agc Ser	ccg Pro	gat Asp	ctc Leu	atc Ile 970	ctg Leu	ccc Pro	ctg Leu	cat His	gg G] 97	ga c Iy L 75	tg i	agg Arg	aa As	iC sn	gtc Val	2985
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Leu Val Asn Ala Ser Leu Gly Trp Pro Asn Gly Leu Ala Leu Asp Leu
500
510 Gln Glu Gly Lys Leu Tyr Trp Gly Asp Ala Lys Thr Asp Lys Ile Glu 515Val Ile Asn Val Asp Gly Thr Lys Arg Arg Thr Leu Leu Glu Asp Lys 530 540 Leu Pro His Ile Phe Gly Phe Thr Leu Leu Gly Asp Phe Ile Tyr Trp 545 555 560 Thr Asp Trp Gln Arg Arg Ser Ile Glu Arg Val His Lys Val Lys Ala Ser Arg Asp Val Ile Ile Asp Gln Leu Pro Asp Leu Met Gly Leu Lys Ala Val Asn Val Ala Lys Val Val Gly Thr Asn Pro Cys Ala Asp Arg Asn Gly Gly Cys Ser His Leu Cys Phe Phe Thr Pro His Ala Thr Arg Cys Gly Cys Pro Ile Gly Leu Glu Leu Leu Ser Asp Met Lys Thr Cys Ile Val Pro Glu Ala Phe Leu Val Phe Thr Ser Arg Ala Ala Ile His Arg Ile Ser Leu Glu Thr Asn Asn Asn Asp Val Ala Ile Pro Leu Thr Gly Val Lys Glu Ala Ser Ala Leu Asp Phe Asp Val Ser Asn Asn His 675 Ile Tyr Trp Thr Asp Val Ser Leu Lys Thr Ile Ser Arg Ala Phe Met 690 700 Asn Gly Ser Ser Val Glu His Val Val Glu Phe Gly Leu Asp Tyr Pro 705 715 720 Glu Gly Met Ala Val Asp Trp Met Gly Lys Asn Leu Tyr Trp Ala Asp 725 730 735 Thr Gly Thr Asn Arg Ile Glu Val Ala Arg Leu Asp Gly Gln Phe Arg 740 745Gln Val Leu Val Trp Arg Asp Leu Asp Asn Pro Arg Ser Leu Ala Leu 755 760 Asp Pro Thr Lys Gly Tyr Ile Tyr Trp Thr Glu Trp Gly Gly Lys Pro Arg Ile Val Arg Ala Phe Met Asp Gly Thr Asn Cys Met Thr Leu Val 785 790 800 Asp Lys Val Gly Arg Ala Asn Asp Leu Thr Ile Asp Tyr Ala Asp Gln 815 Arg Leu Tŷr Trp Thr Asp Leu Asp Thr Asn Met Ile Glu Ser Ser Asn 820 Met Leu Gly Gln Glu Arg Val Val Ile Ala Asp Asp Leu Pro His Pro 840 845 Phe Gly Leu Thr Gln Tyr Ser Asp Tyr Ile Tyr Trp Thr Asp Trp Asn 850 860 Leu His Ser Ile Glu Arg Ala Asp Lys Thr Ser Gly Arg Asn Arg Thr Leu Ile Gln Gly His Leu Asp Phe Val Met Asp Ile Leu Val Phe His Ser Ser Arg Gln Asp Gly Leu Asn Asp Cys Met His Asn Asn Gly Gln 900 905Cys Gly Gln Leu Cys Leu Ala Ile Pro Gly Gly His Arg Cys Gly Cys 915 925 Ala Ser His Tyr Thr Leu Asp Pro Ser Ser Arg Asn Cys Ser Pro Pro 930 940 Thr Thr Phe Leu Leu Phe Ser Gln Lys Ser Ala Ile Ser Arg Met Ile 945 950 960 Pro Asp Asp Gln His Ser Pro Asp Leu Ile Leu Pro Leu His Gly Leu 975 Arg Asn Val Lys Ala Ile Asp Tyr Asp Pro Leu Asp Lys Phe Ile Tyr 980 985 990 Trp Val Asp Gly Arg Gln Asn Ile Lys Arg Ala Lys Asp Asp Gly Thr Gln Pro Phe Val Leu Thr Ser Leu Ser Gln Gly Gln Asn Pro Asp 1010 1020 Arg Gln Pro His Asp Leu Ser Ile Asp Ile Tyr Ser Arg Thr Leu 1025 1035 Phe Trp Thr Cys Glu Ala Thr Asn Thr Ile Asn Val His Arg Leu 1040 1050Ser Gly Glu Ala Met Gly Val Val Leu Arg Gly Asp Arg Asp Lys 1055 Pro Arg Ala Ile Val Val Asn Ala Glu Arg Gly Tyr Leu Tyr Phe 1070 1080 Thr Asn Met Gln Asp Arg Ala Ala Lys Ile Glu Arg Ala Ala Leu 1085 1095 Asp Gly Thr Glu Arg Glu Val Leu Phe Thr Thr Gly Leu Ile Arg Pro Val Ala Leu Val Val Asp Asn Thr Leu Gly Lys Leu Phe Trp

Val Asp Ala Asp Leu Lys Arg Ile Glu Ser Cys Asp Leu Ser Gly 1130 1135 Ala Asn Arg Leu Thr Leu Glu Asp Ala Asn Ile Val Gln Pro Leu 1145 1150 1155 Gly Leu Thr Ile Leu Gly Lys His Leu Tyr Trp Ile Asp Arg Gln 1160 1160 Gln Gln Met Ile Glu Arg Val Glu Lys Thr Thr Gly Asp Lys Arg 1175 1180 Thr Arg Ile Gln Gly Arg Val Ala His Leu Thr Gly Ile His Ala 1190 1200 Val Glu Glu Val Ser Leu Glu Glu Phe Ser Ala His Pro Cys Ala 1210 1215 Arg Asp Asn Gly Gly Cys Ser His Ile Cys Ile Ala Lys Gly Asp 1220 Gly Thr Pro Arg Cys Ser Cys Pro Val His Leu Val Leu Leu Gln 1235 1240 Asn Leu Leu Thr Cys Gly Glu Pro Pro Thr Cys Ser Pro Asp Gln 1250 Phe Ala Cys Ala Thr Gly Glu Ile Asp Cys Ile Pro Gly Ala Trp 1265 Arg Cys Asp Gly Phe Pro Glu Cys Asp Asp Gln Ser Asp Glu Glu 1280 Gly Cys Pro Val Cys Ser Ala Ala Gln Phe Pro Cys Ala Arg Gly Gln Cys Val Asp Leu Arg Leu Arg Cys Asp Gly Glu Ala Asp Cys 1310 1320 Gln Asp Arg Ser Asp Glu Ala Asp Cys Asp Ala Ile Cys Leu Pro 1325 Asn Gln Phe Arg Cys Ala Ser Gly Gln Cys Val Leu Ile Lys Gln 1340 1350 Gln Cys Asp Ser Phe Pro Asp Cys Ile Asp Gly Ser Asp Glu Leu 1355 Met Cys Glu Ile Thr Lys Pro Pro Ser Asp Asp Ser Pro Ala His Ser Ser Ala Ile Gly Pro Val Ile Gly Ile Ile Leu Ser Leu Phe 1385 Val Met Gly Gly Val Tyr Phe Val Cys Gln Arg Val Val Cys Gln 1400 Arg Tyr Ala Gly Ala Asn Gly Pro Phe Pro His Glu Tyr Val Ser

Gly Thr Pro His Val Pro Leu Asn Phe Ile Ala Pro Gly Gly Ser 1430 1440 Gln His Gly Pro Phe Thr Gly Ile Ala Cys Gly Lys Ser Met Met Ser Ser Val Ser Leu Met Gly Gly Arg Gly Gly Val Pro Leu Tyr 1460 1470 Thr Lys Ala Thr Leu Tyr Pro Pro Ile Leu Asn Pro Pro Pro Ser 1490 1495 Pro Ala Thr Asp Pro Ser Leu Tyr Asn Met Asp Met Phe Tyr Ser 1505 1510 Ser Asn Ile Pro Ala Thr Val Arg Pro Tyr Arg Pro Tyr Ile Ile 1520 1530 Arg Gly Met Ala Pro Pro Thr Thr Pro Cys Ser Thr Asp Val Cys 1535 Asp Ser Asp Tyr Ser Ala Ser Arg Trp Lys Ala Ser Lys Tyr Tyr 1550 1560 Leu Asp Leu Asn Ser Asp Ser Asp Pro Tyr Pro Pro Pro Pro Thr Pro His Ser Gln Tyr Leu Ser Ala Glu Asp Ser Cys Pro Pro Ser 1580 1590 Pro Ala Thr Glu Arg Ser Tyr Phe His Leu Phe Pro Pro Pro Pro 1595 Ser Pro Cys Thr Asp Ser Ser 1610 1615

51 2479 DNA Homo sapiens

CDS (6)..(1892)

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Leu	G]u 65	Trp	Phe	Leu	Thr	Asp 70	Arg	ser	Gly	Ala	Arg 75	Pro	Arg	Leu	Ala	
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cgg Arg	ggc Gly	cgc Arg	agt Ser	ccc Pro 100	cca Pro	tac Tyr	cag Gln	ctg Leu	gac Asp 105	tcc Ser	cag Gln	ggg GTy	cgc Arg	ctg Leu 110	gtg Val	338
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ggc Gly	agc Ser	ccg Pro	tcc Ser 275	acc Thr	cca Pro	gca Ala	ggc Gly	tgg Trp 280	gta Val	cgc Arg	gag Glu	ggt Gly	gac Asp 285	act Thr	gtc Val	866
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ggc Gly	CCC Pro	tgc Cys	tgc Cys	cgc Arg 580	cag Gln	cgg Arg	cgg Arg	gag Glu	aag Lys 585	ggg Gly	gct Ala	ccg Pro	ccg Pro	cca Pro 590	gag Gly	1778
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															catca	2342
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<213> Homo sapiens

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Leu Leu Cys Arg Gly Asp Gly Ser Pro Ser Pro Glu Tyr Thr Leu Phe Arg Leu Gln Asp Glu Gln Glu Glu Val Leu Asn Val Asn Leu Glu Gly 305 315 320 Asn Leu Thr Leu Glu Gly Val Thr Arg Gly Gln Ser Gly Thr Tyr Gly 325 330Cys Arg Val Glu Asp Tyr Asp Ala Ala Asp Asp Val Gln Leu Ser Lys Thr Leu Glu Leu Arg Val Ala Tyr Leu Asp Pro Leu Glu Leu Ser Glu 355 , 360 365 Gly Lys Val Leu Ser Leu Pro Leu Asn Ser Ser Ala Val Val Asn Cys 370 Ser Val His Gly Leu Pro Thr Pro Ala Leu Arg Trp Thr Lys Asp Ser 385 395 . 400 Thr Pro Leu Gly Asp Gly Pro Met Leu Ser Leu Ser Ser Ile Thr Phe Asp Ser Asn Gly Thr Tyr Val Cys Glu Ala Ser Leu Pro Thr Val Pro 420 430 Val Leu Ser Arg Thr Gln Asn Phe Thr Leu Leu Val Gln Gly Ser Pro Glu Leu Lys Thr Ala Glu Ile Glu Pro Lys Ala Asp Gly Ser Trp Arg Glu Gly Asp Glu Val Thr Leu Ile Cys Ser Ala Arg Gly His Pro Asp 465 475 480 Pro Lys Leu Ser Trp Ser Gln Leu Gly Gly Ser Pro Ala Glu Pro Ile 485 490 490 Pro Gly Arg Gln Gly Trp Val Ser Ser Ser Leu Thr Leu Lys Val Thr Ser Ala Leu Ser Arg Asp Gly Ile Ser Cys Glu Ala Ser Asn Pro His 515 Gly Asn Lys Arg His Val Phe His Phe Gly Thr Val Ser Pro Gln Thr Ser Gln Ala Gly Val Ala Val Met Ala Val Ala Val Ser Val Gly Leu 545 550 560 Leu Leu Leu Val Val Ala Val Phe Tyr Cys Val Arg Arg Lys Gly 575 Pro Cys Cys Arg Gln Arg Arg Glu Lys Gly Ala Pro Pro Gly Glu
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914

tat gaa cat gcc tcc att cac ctg tgg gac ctg ctg gaa ggg aag gaa

Tyr	Glu	His	Ala 210	ser	Ile	His	Leu	Trp 215	Asp	Leu	Leu	Glu	Gly 220	Lys	Glu	
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ccc Pro	ttg Leu	tac Tyr	cag Gln 450	cct Pro	tct Ser	cat His	gct Ala	aca Thr 455	gag Glu	caa Gln	cga Arg	cca Pro	cag Gln 460	aag Lys	gaa Glu	1634
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3488

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<213> Homo sapiens

<400> 54

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Pro Glu Ala Glu Pro Glu Pro Ala Glu Glu Tyr Thr Glu Gln Ser Glu 280 285 Val Glu Ser Thr Glu Tyr Val Asn Arg Gln Phe Met Ala Glu Thr Gln 290 300 Phe Thr Ser Gly Glu Lys Glu Gln Val Asp Glu Trp Thr Val Glu Thr 305 Val Glu Val Val Asn Ser Leu Gln Gln Gln Pro Gln Ala Ala Ser Pro Ser Val Pro Glu Pro His Ser Leu Thr Pro Val Ala Gln Ala Asp Pro Leu Val Arg Arg Gln Arg Val Gln Asp Leu Met Ala Gln Met Gln Gly 365 Pro Tyr Asn Phe Ile Gln Asp Ser Met Leu Asp Phe Glu Asn Gln Thr 370 Leu Asp Pro Ala Ile Val Ser Ala Gln Pro Met Asn Pro Thr Gln Asn 385 400 Met Asp Met Pro Gln Leu Val Cys Pro Pro Val His Ser Glu Ser Arg Leu Ala Gln Pro Asn Gln Val Pro Val Gln Pro Glu Ala Thr Gln Val Pro Leu Val Ser Ser Thr Ser Glu Gly Tyr Thr Ala Ser Gln Pro Leu 435 Tyr Gln Pro Ser His Ala Thr Glu Gln Arg Pro Gln Lys Glu Pro Ile 450 460 Asp Gln Ile Gln Ala Thr Ile Ser Leu Asn Thr Asp Gln Thr Thr Ala 465 470 480 Ser Ser Ser Leu Pro Ala Ala Ser Gln Pro Gln Val Phe Gln Ala Gly Thr Ser Lys Pro Leu His Ser Ser Gly Ile Asn Val Asn Ala Ala Pro 500 510 Phe Gln Ser Met Gln Thr Val Phe Asn Met Asn Ala Pro Val Pro Pro 515 525 Val Asn Glu Pro Glu Thr Leu Lys Gln Gln Asn Gln Tyr Gln Ala Ser 530 540 Tyr Asn Gln Ser Phe Ser Ser Gln Pro His Gln Val Glu Gln Thr Glu 545 550 560 Leu Gln Gln Glu Gln Leu Gln Thr Val Val Gly Thr Tyr His Gly Ser Pro Asp Gln Ser His Gln Val Thr Gly Asn His Gln Gln Pro Pro Gln 580 590

Gln Asn Thr Gly Phe Pro Arg Ser Asn Gln Pro Tyr Tyr Asn Ser Arg Gly Val Ser Arg Gly Gly Ser Arg Gly Ala Arg Gly Leu Met Asn Gly 610 620 Tyr Arg Gly Pro Ala Asn Gly Phe Arg Gly Gly Tyr Asp Gly Tyr Arg 625 635 640 Pro Ser Phe Ser Asn Thr Pro Asn Ser Gly Tyr Thr Gln Ser Gln Phe 645 655 Ser Ala Pro Arg Asp Tyr Ser Gly Tyr Gln Arg Asp Gly Tyr Gln Gln 660 665 Asn Phe Lys Arg Gly Ser Gly Gln Ser Gly Pro Arg Gly Ala Pro Arg Gly Arg Gly Gly Pro Pro Arg Pro Asn Arg Gly Met Pro Gln Met Asn 690 700 Thr Gln Gln Val Asn

55 2131 DNA

Homo sapiens

CDS (374)..(1528)

<400> 55

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<210> 56 <211> 384

<213> Homo sapiens

<400> 56

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Pro Ala Ala Asn Pro Pro Ala Pro Ala Pro Ala Pro Pro Ala Pro Ala Pro Ala Pro Ala Pro Ser Ala Pro Ala Pro Ala Ala Ala Ala Ala Ala Pro Ala Pro Ala Pro Ala Pro Ala Pro Ala Ser Ser Thr Pro His Gly His Cys Pro Asn Gly Val Thr Ala Gly Pro Arg Gly Pro Yal Ala Gly Pro Ala Lys Pro Ser Pro Val Gly Ala Gly Pro Arg Gly Arg His Trp Ala Gly Trp Asp Ala Gly His His Tyr Ala Ser Trp Ala Ala Arg Ala Arg Ala Ser Trp Ala Arg Ala Pro Ala Ala Arg Ala Pro Ala Arg Ala Pro Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Ala Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Arg Ala Ala Ala Arg Arg Ala Ar

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<sup>&</sup>lt;213> Homo sapiens

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ggc GTy	gac Asp	ctg Leu	gct Ala 130	aac Asn	tcc Ser	cat His	ctg Leu	ccc Pro 135	tct Ser	gaa Glu	gtg Val	ctt Leu	tat Tyr 140	atg Met	ctc Leu	672
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ctc Leu	tgc Cys 160	cgc Arg	cac His	atg Met	gtc Val	ttc Phe 165	cag Gln	cgg Arg	ctg Leu	ggc Gly	cag Gln 170	ggt Gly	gac Asp	tac Tyr	gtc Val	768
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gtg Val	gac Asp 960	ctg Leu	gtg Val	ggc Gly	ggc Gly	acg Thr 965	tcc Ser	att Ile	ggc Gly	tct Ser	ttc Phe 970	atc Ile	gga Gly	gcg Ala	ttg Leu	3168
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tgg Trp	gcc Ala	aag Lys	agc Ser	atg Met 995	act Thr	tcg Ser	gtg Val	ctg Leu	gaa Glu 1000	cct Pro	gto Val	ttg Lei	g gad u Asp	c cto Leu 100	acg Thr )5	3264
tac Tyr	cca Pro	gtç Val	acc Thr 1010	tco Ser	ato Met	tto Phe	act Thr	990 Gly 101	to 'Se	t go	c tt a Ph	t aa ie As	ac co sn Ai 10	2c 3 2g 3 20	igc Ser	3309
atc Ile	cat His	cgg Arg	gtc Val 1025	tto Phe	caç Glr	gat Asp	aag Lys	cac Glr 103	at III IO	t ga le Gl	ıg ga lu As	ic ct sp Le	g to eu Ti 10	gg ( p L )35	tg .eu	3354
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ggc tca gcc aca gat gcc tga ggacctcgac aggggtcacc ccctcctcc . Gly Ser Ala 1375

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<213> Homo sapiens

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Val Arg Leu Gln Arg Val Thr Phe Leu Ala Leu His Asn Tyr Leu Gly 285 Leu Thr Asn Glu Leu Phe Ser His Glu Ile Gln Pro Leu Arg Leu Phe 290 300 Pro Ser Pro Gly Leu Pro Thr Arg Thr Ser Pro Val Arg Gly Ser Lys 305 310 315 Arg Met Val Ser Thr Ser Ala Thr Asp Glu Pro Arg Glu Thr Pro Gly 325 Arg Pro Pro Asp Pro Thr Gly Ala Pro Leu Pro Gly Pro Thr Gly Asp 340 Pro Val Lys Pro Thr Ser Leu Glu Thr Pro Ser Pro Pro Leu Leu Ser Arg Cys Val Ser Met Pro Gly Asp Ile Ser Gly Leu Gln Gly Gly Pro Arg Ser Asp Phe Asp Met Ala Tyr Glu Arg Gly Arg Ile Ser Val Ser 385 395 400 Leu Gln Glu Glu Ala Ser Gly Gly Ser Leu Ala Ala Pro Ala Arg Thr 405 410 415 Pro Thr Gln Glu Pro Arg Glu Gln Pro Ala Gly Ala Cys Glu Tyr Ser Tyr Cys Glu Asp Glu Ser Ala Thr Gly Gly Cys Pro Phe Gly Pro Tyr
435 Gln Gly Arg Gln Thr Ser Ser Ile Phe Glu Ala Ala Lys Gln Glu Leu 450 460 Ala Lys Leu Met Arg Ile Glu Asp Pro Ser Leu Leu Asn Ser Arg Val 465 470 475 Leu Leu His His Ala Lys Ala Gly Thr Ile Ile Ala Arg Gln Gly Asp 485 490 491 Gln Asp Val Ser Leu His Phe Val Leu Trp Gly Cys Leu His Val Tyr
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Ala Asp Arg His Ser Asp Phe Ser Arg Leu Ala Arg Val Leu Thr Gly 915 Asn Thr Ile Ala Leu Val Leu Gly Gly Gly Ala Arg Gly Cys Ser His Ile Gly Val Leu Lys Ala Leu Glu Glu Ala Gly Val Pro Val Asp 945 950 960 Leu Val Gly Gly Thr Ser Ile Gly Ser Phe Ile Gly Ala Leu Tyr Ala 975 Glu Glu Arg Ser Ala Ser Arg Thr Arg Gln Arg Ala Arg Glu Trp Ala Lys Ser Met Thr Ser Val Leu Glu Pro Val Leu Asp Leu Thr Tyr Pro Val Thr Ser Met Phe Thr Gly Ser Ala Phe Asn Arg Ser Ile His Arg Val Phe Gln Asp Lys Gln Ile Glu Asp Leu Trp Leu Pro Tyr Phe Asn Val Thr Thr Asp Ile Thr Ala Ser Ala Met Arg Val His Lys Asp Gly Ser Leu Trp Arg Tyr Val Arg Ala Ser Met Thr Leu Ser Gly Tyr Leu Pro Pro Leu Cys Asp Pro Lys Asp Gly His Leu 1070 Leu Met Asp Gly Gly Tyr Ile Asn Asn Leu Pro Ala Asp Ile Ala 1085 Arg Ser Met Gly Ala Lys Thr Val Ile Ala Ile Asp Val Gly Ser 1100 1110 Gln Asp Glu Thr Asp Leu Ser Thr Tyr Gly Asp Ser Leu Ser Gly 1115 Trp Trp Leu Leu Trp Lys Arg Leu Asn Pro Trp Ala Asp Lys Val Lys Val Pro Asp Met Ala Glu Ile Gln Ser Arg Leu Ala Tyr Val 1145 1150 Ser Cys Val Arg Gln Leu Glu Val Val Lys Ser Ser Ser Tyr Cys 1160 1170 Glu Tyr Leu Arg Pro Pro Ile Asp Cys Phe Lys Thr Met Asp Phe 1175 1180 Met Asp Phe 1185Gly Lys Phe Asp Gln Ile Tyr Asp Val Gly Tyr Gln Tyr Gly Lys Ala Val Phe Gly Gly Trp Ser Arg Gly Asn Val Ile Glu Lys Met 1205

L'eu Thr' Asp Arg Arg Ser Thr Asp Leu Asn Glu Ser Arg Arg Ala 1220 1230

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Ile Val Ser Arg Ile Glu Pro Pro Thr Ser Tyr Val Ser Asp Gly 1250 1260

Cys Ala Asp Gly Glu Glu Ser Asp Cys Leu Thr Glu Tyr Glu Glu 1265

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Ala Thr Asp Ala 1325

59 4445

DNA Homo sapiens

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3573

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W	O 200	4/076	682										::	P	CT/US2	2004/006020
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												ggg GTy				912
aag Lys 305	gaa Glu	ttt Phe	gcc Ala	acg Thr	aac Asn 310	ctc Leu	acg Thr	gag Glu	agc Ser	999 Gly 315	gta Val	cac His	ggg Gly	gca Ala	ctg Leu 320	960
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CCT ( Pro l	ctc Leu /	cgc Arg	aag Lys 420	ctg Leu	cag Gln	cca Pro	gaa Glu	ggc GTy 425	cag Gln	act Thr	tct Ser	ggg GTy	agt Ser 430	tcc Ser	cgg Arg	1296
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Gly A	rg L	eu P	he G	ily L	.ys L	ys G	ilu į	ys (	Sly A	\rg	Met (	Gly F	Pro F	ro (	Пy	
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Thr A	sp P 0	ro L	eu G	ју г	.eu A 5	]а L 5	ys L	eu ד.	hr c	ily F	Pro 6	Пу А	sp L	ys A	sp	
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Gly Le	eu P	ro P	he A	la A	<b>Та</b> Т	rp A	sp G	ly p	го т	hr v	'al v	al s	er T	rp L	eu	

4

95

Glu Leu Trp Val Gly Met Pro Ala Trp Tyr Val Ala Ala Cys Arg Ala 100 105 110 Asn Val Lys Ser Gly Ala Ile Met Ala Asn Leu Ser Asp Thr Lys Ile 115 125 Gln Arg Glu Ile Gly Ile Ser Asn Pro Leu His Arg Leu Lys Leu Arg 130 140 Leu Ala Ile Gln Glu Met Val Ser Leu Thr Ser Pro Ser Ala Pro Ala 145 150 160 Ser Ser Arg Thr Ser Thr Gly Asn Val Trp Met Thr His Glu Glu Met Glu Ser Leu Thr Ala Thr Thr Lys Pro Glu Thr Lys Glu Ile Ser Trp 180 185 Glu Gln Ile Leu Ala Tyr Gly Asp Met Asn His Glu Trp Val Gly Asn Asp Trp Leu Pro Ser Leu Gly Leu Pro Gln Tyr Arg Ser Tyr Phe Met 210 220 Glu Ser Leu Val Asp Ala Arg Met Leu Asp His Leu Asn Lys Lys Glu Leu Arg Gly Gln Leu Lys Met Val Asp Ser Phe His Arg Val Ser Leu His Tyr Gly Val Met Cys Leu Lys Arg Leu Asn Tyr Asp Arg Lys Asp 260 270 Leu Glu Arg Arg Glu Glu Ser Gln Thr Gln Ile Arg Asp Val Met 275 280 Val Trp Ser Asn Glu Arg Val Met Gly Trp Val Ser Gly Leu Gly Leu 290 295 Lys Glu Phe Ala Thr Asn Leu Thr Glu Ser Gly Val His Gly Ala Leu 305 310 320 Leu Ala Leu Asp Glu Thr Phe Asp Tyr Ser Asp Leu Ala Leu Leu Leu 335 Gln Ile Pro Thr Gln Asn Ala Gln Ala Arg Gln Leu Leu Glu Lys Glu 340 345 Phe Ser Asn Leu Ile Ser Leu Gly Thr Asp Arg Arg Leu Asp Glu Asp 365 Ser Ala Lys Ser Phe Ser Arg Ser Pro Ser Trp Arg Lys Met Phe Arg Glu Lys Asp Leu Arg Gly Val Thr Pro Asp Ser Ala Glu Met Leu Pro Pro Asn Phe Arg Ser Ala Ala Ala Gly Ala Leu Gly Ser Pro Gly Leu

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	ttc Phe																634
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gct acc ctc ctg ctt ttc ctc att att tct gtg aaa tat gac cat cat Ala Thr Leu Leu Leu Phe Leu Ile Ile Ser Val Lys Tyr Asp His His 545 550 560	1738
cga gac cat cag cga tca aga gcc aat ggc gtg ccc acc agc agg agg Arg Asp His Gln Arg Ser Arg Ala Asn Gly Val Pro Thr Ser Arg Arg 565 570	1786
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<sup>64</sup> 577 PRT Homo sapiens

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1211

1259

1307

1355

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ctg att ttg ctg ctg ggc aac ttc ttc tgc att agg aag aag ccc aaa Leu Ile Leu Leu Gly Asn Phe Phe Cys Ile Arg Lys Lys Pro Lys 400 410 415

gag cca cag cct gag gtg gcg gcc gcg gag gag gag aag ctc cac aag Glu Pro Gln Pro Glu Val Ala Ala Ala Glu Glu Glu Lys Leu His Lys 420 425 430

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Leu Joe Gln Glu Phe Gly Joe Gly Tyr Ser Asp Thr Ala Trp Ile Ser
Ser Ile Leu Leu Ala Met Leu Tyr Gly Thr Gly Pro Leu Cys Ser Wal
Cys Val Asn Arg Phe Gly Cys Arg Pro Yal Met Leu Val Gly Gly Glu
Phe Ala Ser Leu Gly Met Val Ala Ala Ser Phe Cys Arg Ser Ile Ile
Gln Val Tyr Leu Thr Thr Gly Yal Ile Thr Gly Leu Gly Leu Ala Leu
Asn Phe Gln Pro Ser Leu Joe Met Ala Asn Gly Ceu Ala Ala Ala Gly Ser Pro Val Phe Leu
Cys Ala Leu Ser Pro Leu Gly Gln Leu Gly Gln Asp Arg Tyr Gly Trp

<sup>&</sup>lt;213> Homo sapiens

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Va 1 465

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	aac Asn 260			•		265					270					1412
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355	acg Thr				360					365					gtg Val 370	1700
	aag Lys															1748
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Glň	ctc Leu	ser 405	Glň	Glň	Ser	Leū	Asp 410	Phe	۷al	Lyš	Ile	1]e 415	Thr	Ile	Leū	1844
gtc Val	acg Thr 420	gcc Ala	aca Thr	gcg Ala	tcc Ser	agc Ser 425	gtg Val	ggg Gly	gca Ala	gcg Ala	ggc Gly 430	atc Ile	cct Pro	gct Ala	gga Gly	1892
ggt Gly 435	gtc Val	ctc Leu	act Thr	ctg Leu	gcc Ala 440	atc Ile	atc Ile	ctc Leu	gaa Glu	gca Ala 445	gtc Val	aac Asn	ctc Leu	ccg Pro	gtc Val 450	1940
gac Asp	cat His	atc Ile	tcc Ser	ttg Leu 455	atc Ile	ctg Leu	gct Ala	gtg Val	gac Asp 460	tgg Trp	cta Leu	gtc Val	gac Asp	cgg Arg 465	tcc Ser	1988
tgt Cys	acc Thr	gtc Val	ctc Leu 470	aat Asn	gta Val	gaa Glu	ggt Gly	gac Asp 475	gct Ala	ctg Leu	ggg Gly	gca Ala	gga Gly 480	ctc Leu	ctc Leu	2036
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gag Glu	gaa Glu	gga Gly	aac Asn	ccc Pro	ctc Leu	ctc Leu	aaa Lys	cac His	tat Tyr	cgg Arg	ggg GTy	ccc Pro	gca Ala	ggg GTy	gat Asp	2180

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<sup>&</sup>lt;213> Homo sapiens

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Pro Thr Glu Glu Gly Asn Pro Leu Leu Lys His Tyr Arg Gly Pro Ala Gly Asp Ala Thr Val Ala Ser Glu Lys Glu Ser Val Met 535

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<b>33</b> /4	O 200	1/076	682		•									D <i>C</i>	T/HS204	)4/006020
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gcc Ala 180	tgt Cys	gta Val	ctg Leu	gtg Val	ttc Phe 185	tcc Ser	ctg Leu	gcc Ala	ctc Leu	cac His 190	tcc Ser	gtg Val	ttc Phe	gag Glu	ggg GTy 195	1051
ctg Leu	gcg Ala	gta Val	ggg GTy	ctg Leu 200	cag Gln	cga Arg	gac Asp	cgg Arg	gct Ala 205	cgg Arg	gcc Ala	atg Met	gag Glu	ctg Leu 210	tgc Cys	1099
ctg Leu	gct Ala	ttg Leu	ctg Leu 215	ctc Leu	cac His	aag Lys	ggc Gly	atc Ile 220	ctg Leu	gct Ala	gtc Val	agc Ser	ctg Leu 225	tcc Ser		1147
cgg Arg	ctg Leu	ttg Leu 230	cag Gln	agc Ser	cac His	ctt Leu '	agg Arg 235	gça Ala	cag Gln	gtg Val	gtg Val	gct Ala 240	ggc GTy	tgt Cys	ggg Gly	1195
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ctg Leu	ccc Pro	cag Gln	gag Glu 295	ctg Leu	gcc Ala	agt Ser	tct Ser	gag Glu 300	caa Gln	agg Arg	atc Ile	ctc Leu	aag Lys 305	gtc Val	att Ile	1387
ctg Leu	ctc Leu	cta Leu 310	gca Ala	ggc Gly	ttt Phe	gcc Ala	ctg Leu 315	ctc Leu	act Thr	ggc GTy	ctg Leu	ctc Leu 320	ttc Phe	atc Ile	caa Gln	1435
atc Ile	tag	<b>9</b> 999	cttc	aa g	agag	gggc	a gg	ggag	attg	atg	jatca	ggt	gccc	ctgt	tc	1491
tccc	ttcc	ct c	cccc	agtt	g tg	ggga	atag	gaa	ggaa	agg	ggaa	ggga	aa t	actg	aggac	1551
caaa	aagt	tc t	ctgg	gagc	t aa	agat	agag	cct	ttgg	ggc	tatc	tgac	ta a	tgag	aggga	1611
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<213> Homo sapiens

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Ala Pro Arg Arg Gln Asp Ser Glu Asp His Ser Ser Asp Met Phe Asn 115

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73 2380 DNA Homo sapiens

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Cys Pro Ala Ile Arg Glu Met Val Ser Gin Leu Gin Arg Giu Cys Tyr 125 130 135 140	
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gtg gag atg atc cat ttc aag gac ttg ctg ctg cac gaa ccc tac gtg Val Glu Met Ile His Phe Lys Asp Leu Leu His Glu Pro Tyr Val 160 165 170	650
gac ctc gtg aac ttg ctg ctg acc tgt ggg gag gag gtg aag gag gcc Asp Leu Val Asn Leu Leu Eu Thr Cys Gly Glu Glu Val Lys Glu Ala 175 180	698
atc acc cac agc gtg cag gtt cag tgt gag cag aac tgg gga agc ctg Ile Thr His Ser Val Gln Val Gln Cys Glu Gln Asn Trp Gly Ser Leu 190 195 200	746
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<213> Homo sapiens

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170 cgg ggt acc ttc tca gat gtg cct tct agt gtg atg aaa tgc aaa gca Arg Gly Thr Phe Ser Asp Val Pro Ser Ser Val Met Lys Cys Lys Ala 175 180 185 997 1045 tac aca gac tgt ctg agt cag aac ctg gtg gtg atc aag ccg ggg acc Tyr Thr Asp Cys Leu Ser Gln Asn Leu Val Val Ile Lys Pro Gly Thr 190 200 1093 aag gag aca gac aac gtc tgt ggc aca ctc ccg tcc ttc tcc agc tcc Lys Glu Thr Asp Asn Val Cys Gly Thr Leu Pro Ser Phe Ser Ser 205 210 215 1141 acc tca cct tcc cct ggc aca gcc atc ttt cca cgc cct gag cac atg Thr Ser Pro Ser Pro Gly Thr Ala Ile Phe Pro Arg Pro Glu His Met 225 230 235 1189 gaa acc cat gaa gtc cct tcc tcc act tat gtt ccc aaa ggc atg aac Glu Thr His Glu Val Pro Ser Ser Thr Tyr Val Pro Lys Gly Met Asn 240 245 250 tca aca gaa tcc aac tct tct gcc tct gtt aga cca aag gta ctg agt Ser Thr Glu Ser Asn Ser Ser Ala Ser Val Arg Pro Lys Val Leu Ser 255 260 265 1237 agc atc cag gaa ggg aca gtc cct gac aac aca agc tca gca agg ggg Ser Ile Gln Glu Gly Thr Val Pro Asp Asn Thr Ser Ser Ala Arg Gly 270 275 280 1285 1333 aag gaa gac gtg aac aag acc ctc cca aac ctt cag gta gtc aac cac Lys Glu Asp Val Asn Lys Thr Leu Pro Asn Leu Gln Val Val Asn His 285 290 295 300 1381 cag caa ggc ccc cac cac aga cac atc ctg aag ctg ctg ccg tcc atg Gln Gln Gly Pro His His Arg His Ile Leu Lys Leu Leu Pro Ser Met 305 310 gag gcc act ggg ggc gag aag tcc agc acg ccc atc aag ggc ccc aag Glu Ala Thr Gly Glu Lys Ser Ser Thr Pro Ile Lys Gly Pro Lys 320 3251429 1477 agg gga cat cct aga cag aac cta cac aag cat ttt gac atc aat gag Arg Gly His Pro Arg Gln Asn Leu His Lys His Phe Asp Ile Asn Glu 335 340 345 1525 1573 att gtg gtg tgc agt atc cgg aaa agc tcg agg act ctg aaa aag ggg Ile Val Val Cys Ser Ile Arg Lys Ser Ser Arg Thr Leu Lys Lys Gly 365 370 375 1621 ccc cgg cag gat ccc agt gcc att gtg gaa aag gca ggg ctg aag aaa Pro Arg Gln Asp Pro Ser Ala Ile Val Glu Lys Ala Gly Leu Lys Lys 385 390 395 tcc atg act cca acc cag aac cgg gag aaa tgg atc tac tac tgc aat Ser Met Thr Pro Thr Gln Asn Arg Glu Lys Trp Ile Tyr Tyr Cys Asn 400 405 4101669 ggc cat ggt atc gat atc ctg aag ctt gta gca gcc caa gtg gga agc Gly His Gly Ile Asp Ile Leu Lys Leu Val Ala Ala Gln Val Gly Ser 415 420 425 1717 1765 cag tgg aaa gat atc tat cag ttt ctt tgc aat gcc agt gag agg gag Gln Trp Lys Asp Ile Tyr Gln Phe Leu Cys Asn Ala Ser Glu Arg Glu 430 435 440 1813 gtt gct gct ttc tcc aat ggg tac aca gcc gac cac gag cgg gcc tac Val Ala Ala Phe Ser Asn Gly Tyr Thr Ala Asp His Glu Arg Ala Tyr 445 450 455 1861 gca gct ctg cag cac tgg acc atc cgg ggc ccc gag gcc agc ctc gcc Ala Ala Leu Gln His Trp Thr Ile Arg Gly Pro Glu Ala Ser Leu Ala
465
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3641 3662

Homo sapiens

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Asn Ser Ser Ala Ser Val Arg Pro Lys Val Leu Ser Ser Ile Gin Giu 265 270 Gly Thr Val Pro Asp Asn Thr Ser Ser Ala Arg Gly Lys Glu Asp Val Asn Lys Thr Leu Pro Asn Leu Gln Val Val Asn His Gln Gln Gly Pro His His Arg His Ile Leu Lys Leu Leu Pro Ser Met Glu Ala Thr Gly 305 Gly Glu Lys Ser Ser Thr Pro Ile Lys Gly Pro Lys Arg Gly His Pro Arg Gln Asn Leu His Lys His Phe Asp Ile Asn Glu His Leu Pro Trp Met Ile Val Leu Phe Leu Leu Leu Val Leu Val Val Ile Val Val Cys 355 360 365 Ser Ile Arg Lys Ser Ser Arg Thr Leu Lys Lys Gly Pro Arg Gln Asp 370 380 Pro Ser Ala Ile Val Glu Lys Ala Gly Leu Lys Lys Ser Met Thr Pro 385 400 Thr Gln Asn Arg Glu Lys Trp Ile Tyr Tyr Cys Asn Gly His Gly Ile Asp Ile Leu Lys Leu Val Ala Ala Gln Val Gly Ser Gln Trp Lys Asp
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Ile Thr Lys Glu Lys Lys Asp Thr Val Leu Arg Gln Val Arg Leu Asp 580 585 Pro Cys Asp Leu Gln Pro Ile Phe Asp Asp Met Leu His Phe Leu Asn 595 Pro Glu Glu Leu Arg Val Ile Glu Glu Ile Pro Gln Ala Glu Asp Lys 610 620 Leu Asp Arg Leu Phe Glu Ile Ile Gly Val Lys Ser Gln Glu Ala Ser 625 635 640 Gln Thr Leu Leu Asp Ser Val Tyr Ser His Leu Pro Asp Leu Leu 655

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gtc Val	ggt Gly	ttc Phe	tat Tyr 200	tcc Ser	gcc Ala	ttc Phe	ctt Leu	gta Val 205	gca Ala	gat Asp	aag Lys	gtt Val	att Ile 210	gtc Val	act Thr	741
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2697

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<213> Homo sapiens

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Lys Glu Gly Val Lys Phe Asp Glu Ser Glu Lys Thr Lys Glu Ser Arg 595 600 Glu Ala Val Glu Lys Glu Phe Glu Pro Leu Leu Asn Trp Met Lys Asp 610 620 Lys Ala Leu Lys Asp Lys Ile Glu Lys Ala Val Ser Gln Arg Leu 625 630 640 Thr Glu Ser Pro Cys Ala Leu Val Ala Ser Gln Tyr Gly Trp Ser Gly 655 Asn Met Glu Arg Ile Met Lys Ala Gln Ala Tyr Gln Thr Gly Lys Asp 665 670 Ile Ser Thr Asn Tyr Tyr Ala Ser Gln Lys Lys Thr Phe Glu Ile Asn 675 685 Pro Arg His Pro Leu Ile Arg Asp Met Leu Arg Arg Ile Lys Glu Asp 690 700 Glu Asp Asp Lys Thr Val Leu Asp Leu Ala Val Val Leu Phe Glu Thr 705 710 720 Ala Thr Leu Arg Ser Gly Tyr Leu Leu Pro Asp Thr Lys Ala Tyr Gly
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ctg Leu	gac Asp	ttc Phe 80	acg Thr	ggg GTy	gcc Ala	ggc GTy	cgc Arg 85	aag Lys	cac His	agc Ser	aat Asn	ttc Phe 90	ctc Leu	cgg Arg	ctc Leu		351
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gat Asp 670	ggg GTy	gta Val	cag Gln	tct Ser	ctg Leu 675	ctg Leu	aca Thr	cag Gln	aag Lys	tgg Trp 680	tgg Trp	gga Gly	gat Asp	atg Met	gcc Ala 685	2127

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 th           Arg         gag         gag         ceu         gag         th         phe         phe           Arg         gau         gau         ceu         gau         th         gac         atg           Arg         gau         ser         ggu         Arg         pro         gu         ceu         gu         ceu         gu         ceu         gu         ceu         gu         ceu         ceu         gu         ceu         c	Thr         Thr         Pro         690         Trp         A1a         Leu         Val         695           atc         tac         acc         acc         atc         acc         ttc         acc         ttc         acc         ttc         acc         acc         ttc         acc         ttc         acc         acc         ttc         acc         acc	Thr         Thr         Pro         6196         Trp         Ala         Leu         Val         698         Ala           afte         tac         acc         CSC         Cteu         afte         acc         tac  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acc acc acc acc acc acc ac</td> <td>11c         tyr.         7hr         Arg         Ceu         atte         atte         arg         phr         phr&lt;</td>	Thr         Thr         Fro         618         Trp         Ala         Leu         Val         695         Ala         Phe           alte         tag         tag         Arg         Cet         alte         acc         the         acc         by         ser         ser           cag         gag         gag         ceta         gag         the         gag         acg         gag         the         gag         acg         gag         the         gag         acg         gag         the         gag         gag         gag         gag         gag         the         frag         frag	The Thr Pro 690 Trp Ala Leu Val 695 Ala Phe Phe 116 Try 705 Arg Cleu 116 Thr 710 Arg Lau 124 Arg Lau 1	The The Pro 690 trp Ala Leu val 695 Ala Prie Prie Cys  ate tag acc acc acc acc acc acc acc acc acc a	The Thr Pro 696 trp Ala Leu Val 685 Ala Pre Pre 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Lys Lys Thr Cys Thr Thr Phe Ile Val Asp Ser Thr Asp Pro Gly Gly Thr Leu Cys Gln Cys Gly Arg Pro Arg Thr Ala His Pro Ala Val Ala Met Glu Asp Ala Phe Gly Ala Ala Val Val Thr Val Trp Asp Ser Asp Ala His Thr Thr Glu Lys Pro Thr Asp Ala Tyr Gly Glu Leu Asp Phe Thr Gly Ala Gly Arg Lys His Ser Asn Phe Leu Arg Leu Ser Asp Arg Thr Asp Pro Ala Ala Val Tyr Ser Leu Val Thr Arg Thr Trp Gly Phe Arg Ala Pro Asn Leu Val Val Ser Val Leu Gly Gly Ser Gly Gly Pro Val Leu Gln Thr Trp Leu Gln Asp Leu Leu Arg Arg Gly Leu Val Arg 130 140 Ala Ala Gln Ser Thr Gly Ala Trp Ile Val Thr Gly Gly Leu His Thr 145 150 160 Gly Ile Gly Arg His Val Gly Val Ala Val Arg Asp His Gln Met Ala 165 170 175 Ser Thr Gly Gly Thr Lys Val Val Ala Met Gly Val Ala Pro Trp Gly 180 185 Val Val Arg Asn Arg Asp Thr Leu Ile Asn Pro Lys Gly Ser Phe Pro Ala Arg Tyr Arg Trp Arg Gly Asp Pro Glu Asp Gly Val Gln Phe Pro Leu Asp Tyr Asn Tyr Ser Ala Phe Phe Leu Val Asp Asp Gly Thr His 235 240 Gly Cys Leu Gly Gly Glu Asn Arg Phe Arg Leu Arg Leu Glu Ser Tyr 250 Ile Ser Gln Gln Lys Thr Gly Val Gly Gly Thr Gly Ile Asp Ile Pro Val Leu Leu Leu Ile Asp Gly Asp Glu Lys Met Leu Thr Arg Ile Glu Asn Ala Thr Gln Ala Gln Leu Pro Cys Leu Leu Val Ala Gly Ser Gly Gly Ala Ala Asp Cys Leu Ala Glu Thr Leu Glu Asp Thr Leu Ala 305 310 320 Pro Gly Ser Gly Gly Ala Arg Gln Gly Glu Ala Arg Asp Arg Ile Arg

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Leu Ala Met Gln Ala Asp Ala Arg Ala Phe Phe Ala Gln Asp Gly Val Gln Ser Leu Leu Thr Gln Lys Trp Trp Gly Asp Met Ala Ser Thr Thr 675 680 Pro Ile Trp Ala Leu Val Leu Ala Phe Phe Cys Pro Pro Leu Ile Tyr 690 700 Thr Arg Leu Ile Thr Phe Arg Lys Ser Glu Glu Glu Pro Thr Arg Glu 705 715 710 720 Glu Leu Glu Phe Asp Met Asp Ser Val Ile Asn Gly Glu Gly Pro Val Gly Thr Ala Asp Pro Ala Glu Lys Thr Pro Leu Gly Val Pro Arg Gln 740 740Ser Gly Arg Pro Gly Cys Cys Gly Gly Arg Cys Gly Gly Arg Cys 760 765 Leu Arg Arg Trp Phe His Phe Trp Gly Ala Pro Val Thr Ile Phe Met 770 780 Gly Asn Val Val Ser Tyr Leu Leu Phe Leu Leu Phe Ser Arg Val Leu Leu Val Asp Phe Gln Pro Ala Pro Pro Gly Ser Leu Glu Leu Leu 810 815 Leu Tyr Phe Trp Ala Phe Thr Leu Leu Cys Glu Glu Leu Arg Gln Gly 820 825 Leu Ser Gly Gly Gly Gly Ser Leu Ala Ser Gly Gly Pro Gly 845 His Ala Ser Leu Ser Gln Arg Leu Arg Leu Tyr Leu Ala Asp Ser Trp 850 860 Asn Gln Cys Asp Leu Val Ala Leu Thr Cys Phe Leu Leu Gly Val Gly 865 870 875 Cys Arg Leu Thr Pro Gly Leu Tyr His Leu Gly Arg Thr Val Leu Cys 895Ile Asp Phe Met Val Phe Thr Val Arg Leu Leu His Ile Phe Thr Val Asn Lys Gln Leu Gly Pro Lys Ile Val Ile Val Ser Lys Met Met Lys Asp Val Phe Phe Leu Phe Phe Leu Gly Val Trp Leu Val Ala Tyr 930 940 Gly Val Ala Thr Glu Gly Leu Leu Arg Pro Arg Asp Ser Asp Phe Pro 945 955 960 Ser Ile Leu Arg Arg Val Phe Tyr Arg Pro Tyr Leu Gln Ile Phe Gly
970
975

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27

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